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(54) Title: ENZYMES

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(57) Abstract: Various embodiments of the invention provide human enzymes (ENZM) and polynucleotides which identify and encode ENZM. Embodiments of the invention also provide expression vectors, host cells, antibodies, agonists, and antagonists. Other embodiments provide methods for diagnosing, treating, or preventing disorders associated with abernant expression of ENZM.

#### ENZYMES

## TECHNICAL FIELD

The invention relates to novel nucleic acids, enzymes encoded by these nucleic acids, and to

the use of these nucleic acids and proteins in the diagnosis, treatment, and prevention of
autoimmune/inflammatory disorders, infectious disorders, immune deficiencies, disorders of
metabolism, reproductive disorders, neurological disorders, cardiovascular disorders, eye disorders, and
cell proliferative disorders, including cancer. The invention also relates to the assessment of the
effects of exogenous compounds on the expression of nucleic acids and enzymes.

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### BACKGROUND OF THE INVENTION

The cellular processes of biogenesis and biodegradation involve a number of key enzyme classes including oxidoreductases, transferases, hydrolases, lyases, isomerases, ligases, and others.

Each class of enzyme comprises many substrate-specific enzymes having precise and well regulated functions. Enzymes facilitate metabolic processes such as glycolysis, the tricarboxylic cycle, and fatty acid metabolism; synthesis or degradation of amino acids, steroids, phospholipids, and alcohols; regulation of cell signaling, proliferation, inflamation, and apoptosis; and through catalyzing critical steps in DNA replication and repair and the process of translation.

## Oxidoreductases

Many pathways of biogenesis and biodegradation require oxidoreductase (dehydrogenase or reductase) activity, coupled to reduction or oxidation of a cofactor. Potential cofactors include cytochromes, oxygen, disulfide, iron-sulfur proteins, flavin adenine dinucleotide (FAD), and the nicotinamide adenine dinucleotides NAD and NADP (Newsholme, E.A. and A.R. Leech (1983) Biochemistry for the Medical Sciences, John Wiley and Sons, Chichester, U. K. pp. 779-793).

Reductase activity catalyzes transfer of electrons between substrate(s) and cofactor(s) with concurrent oxidation of the cofactor. Reverse dehydrogenase activity catalyzes the reduction of a cofactor and consequent oxidation of the substrate. Oxidoreductase enzymes are a broad superfamily that catalyze reactions in all cells of organisms, including metabolism of sugar, certain detoxification reactions, and synthesis or degradation of fatty acids, amino acids, glucocorticoids, estrogens, androgens, and prostaglandins. Different family members may be referred to as oxidoreductases, oxidases, reductases, or dehydrogenases, and they often have distinct cellular locations such as the cytosol, the plasma membrane, mitochondrial inner or outer membrane, and peroxisomes.

Short-chain alcohol dehydrogenases (SCADs) are a family of dehydrogenases that share only

15% to 30% sequence identity, with similarity predominantly in the coenzyme binding domain and the substrate binding domain. In addition to their role in detoxification of ethanol, SCADs are involved in synthesis and degradation of fatty acids, steroids, and some prostaglandins, and are therefore implicated in a variety of disorders such as lipid storage disease, myopathy, SCAD deficiency, and 5 certain genetic disorders. For example, retinol dehydrogenase is a SCAD-family member (Simon, A. et al. (1995) J. Biol. Chem. 270:1107-1112) that converts retinol to retinal, the precursor of retinoic acid. Retinoic acid, a regulator of differentiation and apoptosis, has been shown to down-regulate genes involved in cell proliferation and inflammation (Chai, X. et al. (1995) J. Biol. Chem. 270:3900-3904). In addition, retinol dehydrogenase has been linked to hereditary eye diseases such as autosomal recessive childhood-onset severe retinal dystrophy (Simon, A. et al. (1996) Genomics 36:424-430).

Membrane-bound succinate dehydrogenases (succinate:quinone reductases, SQR) and fumarate reductases (quinol:fumarate reductases, QFR) couple the oxidation of succinate to fumarate with the reduction of quinone to quinol, and also catalyze the reverse reaction. QFR and SQR complexes are collectively known as succinate:quinone oxidoreductases (EC 1.3.5.1) and have similar compositions. The complexes consist of two hydrophilic and one or two hydrophobic, membrane-integrated subunits. The larger hydrophilic subunit A carries covalently bound flavin adenine dinucleotide; subunit B contains three iron-sulphur centers (Lancaster, C.R. and A. Kroger (2000) Biochim. Biophys. Acta 1459:422-431). The full-length cDNA sequence for the flavoprotein subunit of human heart succinate dehydrogenase (succinate: (acceptor) oxidoreductase; EC 1.3.99.1) is similar to the bovine succinate dehydrogenase in that it contains a cysteine triplet and in that the active site contains an additional cysteine that is not present in yeast or prokaryotic SQRs (Morris, A.A. et al. (1994) Biochim. Biophys. Acta 29:125-128).

Propagation of nerve impulses, modulation of cell proliferation and differentiation, induction of the immune response, and tissue homeostasis involve neurotransmitter metabolism (Weiss, B. (1991) Neurotoxicology 12:379-386; Collins, S.M. et al. (1992) Ann. N.Y. Acad. Sci. 664:415-424; Brown, J.K. and H. Imam (1991) J. Inherit. Metab. Dis. 14:436-458). Many pathways of neurotransmitter metabolism require oxidoreductase activity, coupled to reduction or oxidation of a cofactor, such as NAD¹/NADH (Newsholme and Leech, supra, pp. 779-793). Degradation of catecholamines (epinephrine or norepinephrine) requires alcohol dehydrogenase (in the brain) or aldehyde dehydrogenase (in peripheral tissue). NAD¹-dependent aldehyde dehydrogenase oxidizes 5-hydroxyindole-3-acetate (the product of 5-hydroxytryptamine (serotonin) metabolism) in the brain, blood platelets, liver and pulmonary endothelium (Newsholme and Leech, supra, p. 786). Other

neurotransmitter degradation pathways that utilize NAD'/NADH-dependent oxidoreductase activity include those of L-DOPA (precursor of dopamine, a neuronal excitatory compound), glycine (an inhibitory neurotransmitter in the brain and spinal cord), histamine (liberated from mast cells during the inflammatory response), and taurine (an inhibitory neurotransmitter of the brain stem, spinal cord and retina) (Newsholme and Leech, supra, pp. 790, 792). Epigenetic or genetic defects in neurotransmitter metabolic pathways can result in diseases including Parkinson disease and inherited myoclonus (McCance, K.L. and S.E. Huether (1994) Pathophysiology, Mosby-Year Book, Inc., St. Louis, MO pp. 402-404; Gundlach, A.L. (1990) FASEB J. 4:2761-2766).

Tetrahydrofolate is a derivatized glutamate molecule that acts as a carrier, providing activated

one-carbon units to a wide variety of biosynthetic reactions, including synthesis of purines, pyrimidines, and the amino acid methionine. Tetrahydrofolate is generated by the activity of a holoenzyme complex called tetrahydrofolate synthase, which includes three enzyme activities: tetrahydrofolate dehydrogenase, tetrahydrofolate cyclohydrolase, and tetrahydrofolate synthetase. Thus, tetrahydrofolate dehydrogenase plays an important role in generating building blocks for nucleic and amino acids, crucial to proliferating cells.

3-Hydroxyacyl-CoA dehydrogenase (3HACD) is involved in fatty acid metabolism. It catalyzes the reduction of 3-hydroxyacyl-CoA to 3-oxoacyl-CoA, with concomitant oxidation of NAD to NADH, in the mitochondria and peroxisomes of eukaryotic cells. In peroxisomes, 3HACD and enoyl-CoA hydratase form an enzyme complex called bifunctional enzyme, defects in which are associated with peroxisomal bifunctional enzyme deficiency. This interruption in fatty acid metabolism produces accumulation of very-long chain fatty acids, disrupting development of the brain, bone, and adrenal glands. Infants born with this deficiency typically die within 6 months (Watkins, P. et al. (1989) J. Clin. Invest. 83:771-777; Online Mendelian Inheritance in Man (OMIM), #261515). The neurodegeneration characteristic of Alzheimer's disease involves development of extracellular plaques in certain brain regions. A major protein component of these plaques is the peptide amyloid-β (Aβ), which is one of several cleavage products of amyloid precursor protein (APP). 3HACD has been shown to bind the Aβ peptide, and is overexpressed in neurons affected in Alzheimer's disease. In addition, an antibody against 3HACD can block the toxic effects of Aβ in a cell culture model of Alzheimer's disease (Yan, S. et al. (1997) Nature 389:689-695; OMIM, #602057).

Steroids such as estrogen, testosterone, and corticosterone are generated from a common precursor, cholesterol, and interconverted. Enzymes acting upon cholesterol include dehydrogenases. Steroid dehydrogenases, such as the hydroxysteroid dehydrogenases, are involved in hypertension, fertility, and cancer (Duax, W.L. and D. Ghosh (1997) Steroids 62:95-100). One such dehydrogenases

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is 3-oxo-5-a-steroid dehydrogenase (OASD), a microsomal membrane protein highly expressed in prostate and other androgen-responsive tissues. OASD catalyzes the conversion of testosterone into dihydrotestosterone, which is the most potent androgen. Dihydrotestosterone is essential for the formation of the male phenotype during embryogenesis, as well as for proper androgen-mediated growth of tissues such as the prostate and male genitalia. A defect in OASD leads to defective formation of the external genitalia (Andersson, S. et al. (1991) Nature 354:159-161; Labrie, F. et al. (1992) Endocrinology 131:1571-1573; OMIM #264600).

17β-hydroxysteroid dehydrogenase (17βHSD6) plays an important role in the regulation of the male reproductive hormone, dihydrotestosterone (DHTT). 17βHSD6 acts to reduce levels of DHTT by oxidizing a precursor of DHTT, 3α-diol, to androsterone which is readily glucuronidated and removed. 17βHSD6 is active with both androgen and estrogen substrates in embryonic kidney 293 cells. Isozymes of 17βHSD catalyze oxidation and/or reduction reactions in various tissues with preferences for different steroid substrates (Biswas, M.G. and D.W. Russell (1997) J. Biol. Chem. 272:15959-15966). For example, 17βHSD1 preferentially reduces estraid and is abundant in the ovary and placenta. 17βHSD2 catalyzes oxidation of androgens and is present in the endometrium and placenta. 17βHSD3 is exclusively a reductive enzyme in the testis (Geissler, W.M. et al. (1994) Nature Genet. 7:34-39). An excess of androgens such as DHTT can contribute to diseases such as benign prostatic hyperplasia and prostate cancer.

The oxidoreductase isocitrate dehydrogenase catalyzes the conversion of isocitrate to a
ketoglutarate, a substrate of the citric acid cycle. Isocitrate dehydrogenase can be either NAD or

NADP dependent, and is found in the cytosol, mitochondria, and peroxisomes. Activity of isocitrate

dehydrogenase is regulated developmentally, and by hormones, neurotransmitters, and growth factors.

Hydroxypyruvate reductase (HPR), a peroxisomal 2-hydroxyacid dehydrogenase in the
glycolate pathway, catalyzes the conversion of hydroxypyruvate to glycerate with the oxidation of both
NADH and NADPH. The reverse dehydrogenase reaction reduces NAD\* and NADP\*. HPR
recycles nucleotides and bases back into pathways leading to the synthesis of ATP and GTP, which
are used to produce DNA and RNA and to control various aspects of signal transduction and energy
metabolism. Purine nucleotide biosynthesis inhibitors are used as antiproliferative agents to treat
cancer and viral diseases. HPR also regulates biochemical synthesis of serine and cellular serine
levels available for protein synthesis.

The mitochondrial electron transport (or respiratory) chain is the series of oxidoreductase-type enzyme complexes in the mitochondrial membrane that is responsible for the transport of electrons from NADH to oxygen and the coupling of this oxidation to the synthesis of ATP (oxidative

phosphorylation). ATP provides energy to drive energy-requiring reactions. The key respiratory chain complexes are NADH:ubiquinone oxidoreductase (complex I), succinate:ubiquinone oxidoreductase (complex II), cytochrome c<sub>1</sub>-b oxidoreductase (complex III), cytochrome c oxidase (complex IV), and ATP synthase (complex V) (Alberts, B. et al. (1994) Molecular Biology of the Cell.

5 Garland Publishing, Inc., New York, NY, pp. 677-678). All of these complexes are located on the inner matrix side of the mitochondrial membrane except complex II, which is on the cytosolic side where it transports electrons generated in the citric acid cycle to the respiratory chain. Electrons released in oxidation of succinate to fumarate in the citric acid cycle are transferred through electron carriers in complex II to membrane bound ubiquinone (Q). Transcriptional regulation of these nuclear
10 encoded genes controls the biogenesis of respiratory enzymes. Defects and altered expression of enzymes in the respiratory chain are associated with a variety of disease conditions.

Other dehydrogenase activities using NAD as a cofactor include 3-hydroxyisobutyrate dehydrogenase (3HBD), which catalyzes the NAD-dependent oxidation of 3-hydroxyisobutyrate to methylmalonate semialdehyde within mitochondria. 3-hydroxyisobutyrate levels are elevated in ketoacidosis, methylmalonic acidemia, and other disorders (Rougraff, P.M. et al. (1989) J. Biol. Chem. 264:5899-5903). Another mitochondrial dehydrogenase important in amino acid metabolism is the enzyme isovaleryl-CoA-dehydrogenase (IVD). IVD is involved in leucine metabolism and catalyzes the oxidation of isovaleryl-CoA o3-methylcrotonyl-CoA. Human IVD is a tetrameric flavoprotein synthesized in the cytosol with a mitochondrial import signal sequence. A mutation in the gene encoding IVD results in isovaleric acidemia (Vockley, J. et al. (1992) J. Biol. Chem. 267:2494-2501).

The family of glutathione peroxidases encompass tetrameric glutathione peroxidases (GPx1-3) and the monomeric phospholipid hydroperoxide glutathione peroxidase (PHGPx/GPx4). Although the overall homology between the tetrameric enzymes and GPx4 is less than 30%, a pronounced similarity has been detected in clusters involved in the active site and a common catalytic triad has been defined 25 by structural and kinetic data (Epp, O. et al. (1983) Eur. J. Biochem. 133:51-69). GPx1 is ubiquitously expressed in cells, whereas GPx2 is present in the liver and colon, and GPx3 is present in plasma. GPx4 is found at low levels in all tissues but is expressed at high levels in the testis (Ursini, F. et al (1995) Meth. Enzymol. 252:38-53). GPx4 is the only monomeric glutathione peroxidase found in mammals and the only mammalian glutathione peroxidase to show high affinity for and reactivity with phospholipid hydroperoxides, and to be membrane associated. A tandem mechanism for the antioxidant activities of GPx4 and vitamin E has been suggested. GPx4 has alternative transcription and translation start sites which determine its subcellular localization (Esworthy, R.S. et al. (1994) Gene 144:317-318; and Maiorino, M. et al. (1990) Meth. Enzymol. 186:448-450).

The glutathione S-transferases (GST) are a ubiquitous family of enzymes with dual substrate specificities that perform important biochemical functions of xenobiotic biotransformation and detoxification, drug metabolism, and protection of tissues against peroxidative damage. They catalyze the conjugation of an electrophile with reduced glutathione (GSH) which results in either activation or 5 deactivation/detoxification. The absolute requirement for binding reduced GSH to a variety of chemicals necessitates a diversity in GST structures in various organisms and cell types. GSTs are homodimeric or heterodimeric proteins localized in the cytosol. The major isozymes share common structural and catalytic properties and include four major classes, Alpha, Mu, Pi, and Theta. Each GST possesses a common binding site for GSH, and a variable hydrophobic binding site specific for 10 its particular electrophilic substrates. Specific amino acid residues within GSTs have been identified as important for these binding sites and for catalytic activity. Residues Q67, T68, D101, E104, and R131 are important for the binding of GSH (Lee, H.-C. et al. (1995) J. Biol. Chem. 270:99-109). Residues R13, R20, and R69 are important for the catalytic activity of GST (Stenberg, G. et al. (1991) Biochem. J. 274:549-555).

GSTs normally deactivate and detoxify potentially mutagenic and carcinogenic chemicals. Some forms of rat and human GSTs are reliable preneoplastic markers of carcinogenesis. Dihalomethanes, which produce liver tumors in mice, are believed to be activated by GST (Thier, R, et al. (1993) Proc. Natl. Acad. Sci. USA 90:8567-8580). The mutagenicity of ethylene dibromide and ethylene dichloride is increased in bacterial cells expressing the human Alpha GST, A1-1, while the 20 mutagenicity of aflatoxin B1 is substantially reduced by enhancing the expression of GST (Simula, T.P. et al. (1993) Carcinogenesis 14:1371-1376). Thus, control of GST activity may be useful in the control of mutagenesis and carcinogenesis.

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GST has been implicated in the acquired resistance of many cancers to drug treatment, the phenomenon known as multi-drug resistance (MDR). MDR occurs when a cancer patient is treated 25 with a cytotoxic drug such as cyclophosphamide and subsequently becomes resistant to this drug and to a variety of other cytotoxic agents as well. Increased GST levels are associated with some drug resistant cancers, and it is believed that this increase occurs in response to the drug agent which is then deactivated by the GST catalyzed GSH conjugation reaction. The increased GST levels then protect the cancer cells from other cytotoxic agents for which GST has affinity. Increased levels of 30 A1-1 in tumors has been linked to drug resistance induced by cyclophosphamide treatment (Dirven, H.A. et al. (1994) Cancer Res. 54:6215-6220). Thus control of GST activity in cancerous tissues may be useful in treating MDR in cancer patients.

The reduction of ribonucleotides to the corresponding deoxyribonucleotides, needed for DNA

synthesis during cell proliferation, is catalyzed by the enzyme ribonucleotide diphosphate reductase.

Glutaredoxin is a glutathione (GSH)-dependent hydrogen donor for ribonucleotide diphosphate reductase and contains the active site consensus sequence -C-P-Y-C-. This sequence is conserved in glutaredoxins from such different organisms as Escherichia coli, vaccinia virus, yeast, plants, and mammalian cells. Glutaredoxin has inherent GSH-disulfide oxidoreductase (thioltransferase) activity in a coupled system with GSH, NADPH, and GSH-reductase, catalyzing the reduction of low molecular weight disulfides as well as proteins. Glutaredoxin has been proposed to exert a general thiol redox control of protein activity by acting both as an effective protein disulfide reductase, similar to thioredoxin, and as a specific GSH-mixed disulfide reductase (Padilla, C.A. et al. (1996) FEBS Lett. 378:69-73).

In addition to their important role in DNA synthesis and cell division, glutaredoxin and other thioproteins provide effective antioxidant defense against oxygen radicals and hydrogen peroxide (Schallreuter, K.U. and J.M. Wood (1991) Melanoma Res. 1:159-167). Glutaredoxin is the principal agent responsible for protein dethiolation in vivo and reduces dehydroascorbic acid in normal human neutrophils (Jung, C.H. and J.A. Thomas (1996) Arch. Biochem. Biophys. 335:61-72; Park, J.B. and M. Levine (1996) Biochem. J. 315:931-938).

The thioredoxin system serves as a hydrogen donor for ribonucleotide reductase and as a regulator of enzymes by redox control. It also modulates the activity of transcription factors such as NF-κB, AP-1, and steroid receptors. Several cytokines or secreted cytokine-like factors such as adult T-cell leukemia-derived factor, 3B6-interleukin-1, T-hybridoma-derived (MP-6) B cell stimulatory factor, and early pregnancy factor have been reported to be identical to thioredoxin (Holmgren, A. (1985) Annu. Rev. Biochem. 54:237-271; Abate, C. et al. (1990) Science 249:1157-1161; Tagaya, Y. et al. (1989) EMBO J. 8:757-764; Wakasugi, H. (1987) Proc. Natl. Acad. Sci. USA 84:804-808; Rosen, A. et al. (1995) Int. Immunol. 7:625-633). Thus thioredoxin secreted by stimulated lymphocytes (Yodoi, J. and T. Tursz (1991) Adv. Cancer Res. 57:381-411; Tagaya, N. et al. (1990) Proc. Natl. Acad. Sci. USA 87:8282-8286) has extracellular activities including a role as a regulator of cell growth and a mediator in the immune system (Miranda-Vizuete, A. et al. (1996) J. Biol. Chem. 271:19099-19103; Yamauchi, A. et al. (1992) Mol. Immunol. 29:263-270). Thioredoxin and thioredoxin reductase protect against cytotoxicity mediated by reactive oxygen species in disorders such as Alzheimer's disease (Lovell, M.A. (2000) Free Radic. Biol. Med. 28:418-427).

The selenoprotein thioredoxin reductase is secreted by both normal and neoplastic cells and has been implicated as both a growth factor and as a polypeptide involved in apoptosis (Soderberg, A. et al. (2000) Cancer Res. 60:2281-2289). An extracellular plasmin reductase secreted by hamster

ovary cells (HT-1080) has been shown to participate in the generation of angiostatin from plasmin. In this case, the reduction of the plasmin disulfide bonds triggers the proteolytic cleavage of plasmin which yields the angiogenesis inhibitor, angiostatin (Stathakis, P. et al. (1997) J. Biol. Chem. 272:20641-20645). Low levels of reduced sulfhydryl groups in plasma has been associated with rheumatoid arthritis. The failure of these sulfhydryl groups to scavenge active oxygen species (e.g., hydrogen peroxide produced by activated neutrophils) results in oxidative damage to surrounding

tissues and the resulting inflammation (Hall, N.D. et al. (1994) Rheumatol. Int. 4:35-38).

Another example of the importance of redox reactions in cell metabolism is the degradation of saturated and unsaturated fatty acids by mitochondrial and peroxisomal beta-oxidation enzymes which sequentially remove two-carbon units from Coenzyme A (CoA)-activated fatty acids. The main beta-oxidation pathway degrades both saturated and unsaturated fatty acids while the auxiliary pathway performs additional steps required for the degradation of unsaturated fatty acids.

The pathways of mitchondrial and peroxisomal beta-oxidation use similar enzymes, but have different substrate specificities and functions. Mitochondria oxidize short-, medium-, and long-chain fatty acids to produce energy for cells. Mitochondrial beta-oxidation is a major energy source for cardiac and skeletal muscle. In liver, it provides ketone bodies to the peripheral circulation when glucose levels are low as in starvation, endurance exercise, and diabetes (Eaton, S. et al. (1996) Biochem. J. 320:345-357). Peroxisomes oxidize medium-, long-, and very-long-chain fatty acids, dicarboxylic fatty acids, branched fatty acids, prostaglandins, xenobiotics, and bile acid intermediates.

The chief roles of peroxisomal beta-oxidation are to shorten toxic lipophilic carboxylic acids to facilitate their excretion and to shorten very-long-chain fatty acids prior to mitochondrial beta-oxidation (Mannaerts, G.P. and P.P. Van Veldhoven (1993) Biochimie 75:147-158).

The auxiliary beta-oxidation enzyme 2,4-dienoyl-CoA reductase catalyzes the following reaction:

trans-2, cis/trans-4-dienoyl-CoA + NADPH + H+---> trans-3-enoyl-CoA + NADP+

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This reaction removes even-numbered double bonds from unsaturated fatty acids prior to their entry into the main beta-oxidation pathway (Koivuranta, K.T. et al. (1994) Biochem. J. 304:787-792). The enzyme may also remove odd-numbered double bonds from unsaturated fatty acids (Smeland, T.E. et al. (1992) Proc. Natl. Acad. Sci. USA 89:6673-6677).

Rat 2,4-dienoyl-CoA reductase is located in both mitochondria and peroxisomes (Dommes, V. et al. (1981) J. Biol. Chem. 256:8259-8262). Two immunologically different forms of rat mitochondrial enzyme exist with molecular masses of 60 kDa and 120 kDa (Hakkola, E.H. and J.K.

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Hiltunen (1993) Eur. J. Biochem. 215:199-204). The 120 kDa mitochondrial rat enzyme is synthesized as a 335 amino acid precursor with a 29 amino acid N-terminal leader peptide which is cleaved to form the mature enzyme (Hirose, A. et al. (1990) Biochim. Biophys. Acta 1049:346-349). A human mitochondrial enzyme 83% similar to rat enzyme is synthesized as a 335 amino acid residue precursor with a 19 amino acid N-terminal leader peptide (Koivuranta et al., supra). These cloned human and rat mitochondrial enzymes function as homotetramers (Koivuranta et al., supra). A Saccharomyces cerevisiae peroxisomal 2,4-dienoyl-CoA reductase is 295 amino acids long, contains a C-terminal peroxisomal targeting signal, and functions as a homodimer (Coe, J.G.S. et al. (1994) Mol. Gen. Genet. 244:661-672; and Gurvitz, A. et al. (1997) J. Biol. Chem. 272:22140-22147). All 2,4-dienoyl-10 CoA reductases have a fairly well conserved NADPH binding site motif (Koivuranta et al., supra).

The main pathway beta-oxidation enzyme enovl-CoA hydratase catalyzes the reaction:

# 2-trans-enoyl-CoA + H2O <---> 3-hydroxyacyl-CoA

This reaction hydrates the double bond between C-2 and C-3 of 2-trans-enoyl-CoA, which is 15 generated from saturated and unsaturated fatty acids (Engel, C.K. et al. (1996) EMBO J. 15:5135-5145). This step is downstream from the step catalyzed by 2,4-dienoyl-reductase. Different enoyl-CoA hydratases act on short-, medium-, and long-chain fatty acids (Eaton et al., supra). Mitochondrial and peroxisomal enoyl-CoA hydratases occur as both mono-functional enzymes and as part of multi-functional enzyme complexes. Human liver mitochondrial short-chain enoyl-CoA hydratase is synthesized as a 290 amino acid precursor with a 29 amino acid N-terminal leader peptide (Kanazawa, M. et al. (1993) Enzyme Protein 47:9-13; and Janssen, U. et al. (1997) Genomics 40:470-475). Rat short-chain enoyl-CoA hydratase is 87% identical to the human sequence in the mature region of the protein and functions as a homohexamer (Kanazawa et al., supra; and Engel et al., 25 supra). A mitochondrial trifunctional protein exists that has long-chain enoyl-CoA hydratase, 3hydroxyacyl-CoA dehydrogenase, and long-chain 3-oxothiolase activities (Eaton et al., supra). In human peroxisomes, enoyl-CoA hydratase activity is found in both a 327 amino acid residue monofunctional enzyme and as part of a multi-functional enzyme, also known as bifunctional enzyme, which possesses enoyl-CoA hydratase, enoyl-CoA isomerase, and 3-hydroxyacyl-CoA hydrogenase 30 activities (FitzPatrick, D.R. et al. (1995) Genomics 27:457-466; and Hoefler, G. et al. (1994) Genomics 19:60-67). A 339 amino acid residue human protein with short-chain enoyl-CoA hydratase activity also acts as an AU-specific RNA binding protein (Nakagawa, J. et al. (1995) Proc. Natl. Acad. Sci. USA 92:2051-2055). All enoyl-CoA hydratases share homology near two active site glutamic acid

residues, with 17 amino acid residues that are highly conserved (Wu, W.-J. et al. (1997) Biochemistry 36:2211-2220).

Inherited deficiencies in mitochondrial and peroxisomal beta-oxidation enzymes are associated with severe diseases, some of which manifest soon after birth and lead to death within a few years.

5 Mitochondrial beta-oxidation associated deficiencies include, e.g., carnitine palmitoyl transferase and carnitine deficiency, very-long-chain acyl-CoA dehydrogenase deficiency, medium-chain acyl-CoA dehydrogenase deficiency, electron transport flavoprotein: ubiquinone oxidoreductase deficiency, trifunctional protein deficiency, and short-chain 3-hydroxyacyl-CoA dehydrogenase deficiency (Eaton et al., supra). Mitochondrial trifunctional protein (including enoyl-CoA hydratase) deficient patients have reduced long-chain enoyl-CoA hydratase activities and suffer from non-ketotic hypoglycemia, sudden infant death syndrome, cardiomyopathy, hepatic dysfunction, and muscle weakness, and may die at an

Defects in mitochondrial beta-oxidation are associated with Reye's syndrome, a disease

characterized by hepatic dysfunction and encephalopathy that sometimes follows viral infection in
children. Reye's syndrome patients may have elevated serum levels of free fatty acids (Cotran, R.S.
et al. (1994) Robbins Pathologic Basis of Disease, W.B. Saunders Co., Philadelphia PA, p.866).
Patients with mitochondrial short-chain 3-hydroxyacyl-CoA dehydrogenase deficiency and mediumchain 3-hydroxyacyl-CoA dehydrogenase deficiency also exhibit Reye-like illnesses (Eaton et al.,

supra; and Egidio, R.J. et al. (1989) Am. Fam. Physician 39:221-226).

early age (Eaton et al., supra).

Inherited conditions associated with peroxisomal beta-oxidation include Zellweger syndrome, neonatal adrenoleukodystrophy, infantile Refsum's disease, acyl-CoA oxidase deficiency, peroxisomal thiolase deficiency, and bifunctional protein deficiency (Suzuki, Y. et al. (1994) Am. J. Hum. Genet. 54:36-43; Hoefler et al., supra). Patients with peroxisomal bifunctional enzyme deficiency, including that of enoyl-CoA hydratase, suffer from hypotonia, seizures, psychomotor defects, and defective neuronal migration; accumulate very-long-chain fatty acids; and typically die within a few years of birth (Watkins, P.A. et al. (1989) J. Clin. Invest. 83:771-777).

Peroxisomal beta-oxidation is impaired in cancerous tissue. Although neoplastic human breast epithelial cells have the same number of peroxisomes as do normal cells, fatty acyl-CoA oxidase activity is lower than in control tissue (el Bouhtoury, F. et al. (1992) J. Pathol. 166:27-35). Human colon carcinomas have fewer peroxisomes than normal colon tissue and have lower fatty-acyl-CoA oxidase and bifunctional enzyme (including enoyl-CoA hydratase) activities than normal tissue (Cable, S. et al. (1992) Virchows Arch. B Cell Pathol. Incl. Mol. Pathol. 62:221-226).

6-phosphogluconate dehydrogenase (6-PGDH) catalyses the NADP\*-dependent oxidative decarboxylation of 6-phosphogluconate to ribulose 5-phosphate with the production of NADPH. The absence or inhibition of 6-PGDH results in the accumulation of 6-phosphogluconate to toxic levels in eukaryotic cells. 6-PGDH is the third enzyme of the pentose phosphate pathway (PPP) and is 5 ubiquitous in nature. In some heterofermentatative species, NAD+ is used as a cofactor with the subsequent production of NADH.

The reaction proceeds through a 3-keto intermediate which is decarboxylated to give the enol of ribulose 5-phosphate, then converted to the keto product following tautomerization of the enol (Berdis A.J. and P.F. Cook (1993) Biochemistry 32:2041-2046). 6-PGDH activity is regulated by the 10 inhibitory effect of NADPH, and the activating effect of 6-phosphogluconate (Rippa, M. et al. (1998) Biochim, Biophys, Acta 1429:83-92). Deficiencies in 6-PGDH activity have been linked to chronic hemolytic anemia.

The targeting of specific forms of 6-PGDH (e.g., enzymes found in trypanosomes) has been suggested as a means for controlling parasitic infections (Tetaud, E. et al. (1999) Bjochem. J. 338:55-15 60). For example, the Trypanosoma brucei enzyme is markedly more sensitive to inhibition by the substrate analogue 6-phospho-2-deoxygluconate and the coenzyme analogue adenosine 2',5'-bisphosphate, compared to the mammalian enzyme (Hanau, S. et al. (1996) Eur. J. Bjochem. 240:592-599).

Ribonucleotide diphosphate reductase catalyzes the reduction of ribonucleotide diphosphates 20 (i.e., ADP, GDP, CDP, and UDP) to their corresponding deoxyribonucleotide diphosphates (i.e., dADP, dGDP, dCDP, and dUDP) which are used for the synthesis of DNA. Ribonucleotide diphosphate reductase thereby performs a crucial role in the de novo synthesis of deoxynucleotide precursors. Deoxynucleotides are also produced from deoxynucleosides by nucleoside kinases via the salvage pathway.

Mammalian ribonucleotide diphosphate reductase comprises two components, an effectorbinding component (E) and a non-heme iron component (F). Component E binds the nucleoside triphosphate effectors while component F contains the iron radical necessary for catalysis. Molecular weight determinations of the E and F components, as well as the holoenzyme, vary according to the methods used in purification of the proteins and the particular laboratory. Component E is 30 approximately 90-100 kDa, component F is approximately 100-120 kDa, and the holoenzyme is 200-250 kDa.

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Ribonucleotide diphosphate reductase activity is adversely effected by iron chelators, such as thiosemicarbazones, as well as EDTA. Deoxyribonucleotide diphosphates also appear to be negative

allosteric effectors of ribonucleotide diphosphate reductase. Nucleotide triphosphates (both ribo- and deoxyribo-) appear to stimulate the activity of the enzyme. 3-methyl-4-nitrophenol, a metabolite of widely used organophosphate pesticides, is a potent inhibitor of ribonucleotide diphosphate reductase in mammalian cells. Some evidence suggests that ribonucleotide diphosphate reductase activity in DNA virus (e.g., herpes virus)-infected cells and in cancer cells is less sensitive to regulation by allosteric regulators and a correlation exists between high ribonucleotide diphosphate reductase activity levels and high rates of cell proliferation (e.g., in hepatomas). This observation suggests that virus-encoded ribonucleotide diphosphate reductases, and those present in cancer cells, are capable of maintaining an increased supply deoxyribonucleotide pool for the production of virus genomes or for the increased DNA synthesis which characterizes cancers cells. Ribonucleotide diphosphate reductase is thus a target for therapeutic intervention (Nutter, L.M. and Y.-C. Cheng (1984) Pharmac. Ther. 26:191-207; and Wright, J.A. (1983) Pharmac. Ther. 22:81-102).

Dihydrodiol dehydrogenases (DD) are monomeric, NAD(P)\*-dependent, 34-37 kDa enzymes responsible for the detoxification of trans-dihydrodiol and anti-diol epoxide metabolites of polycyclic aromatic hydrocarbons (PAH) such as benzo[a]yrene, benz[a]anthracene, 7-methyl-benz[a]anthracene, 7-methyl-benz[a]anthracene, 7-methyl-chrysene. In mammalian cells, an environmental PAH toxin such as benzo[a]yrene is initially epoxidated by a microsomal cytochrome P450 to yield 7R,8R-arene-oxide and subsequently (-)-7R,8R-dihydrodiol ((-)-trans-7,8-dihydroxy-7,8-dihydrobenzo[a]pyrene or (-)-trans-B[a]P-diol) This latter compound is further transformed to the anti-diol epoxide of benzo[a]pyrene (i.e., (±)-anti-7β,8α-dihydroxy-9α,10α-epoxy-7,8,9,10-tetrahydrobenzo[a]pyrene), by the same enzyme or a different enzyme, depending on the species. This resulting anti-diol epoxide of benzo[a]yrene, or the corresponding derivative from another PAH compound, is highly mutagenic.

DD efficiently oxidizes the precursor of the anti-diol epoxide (i.e., trans-dihydrodiol) to
transient catechols which auto-oxidize to quinones, also producing hydrogen peroxide and semiquinone
radicals. This reaction prevents the formation of the highly carcinogenic anti-diol. Anti-diols are not
themselves substrates for DD yet the addition of DD to a sample comprising an anti-diol compound
results in a significant decrease in the induced mutation rate observed in the Ames test. In this
instance, DD is able to bind to and sequester the anti-diol, even though it is not oxidized. Whether
through oxidation or sequestration, DD plays an important role in the detoxification of metabolites of
xenobiotic polyevelic compounds (Penning, T.M. (1993) Chemico-Biological Interactions 89:1-34).

15-oxoprostaglandin 13-reductase (PGR) and 15-hydroxyprostaglandin dehydrogenase (15-PGDH) are enzymes present in the lung that are responsible for degrading circulating prostaglandins.

Oxidative catabolism via passage through the pulmonary system is a common means of reducing the concentration of circulating prostaglandins. 15-PGDH oxidizes the 15-hydroxyl group of a variety of prostaglandins to produce the corresponding 15-oxo compounds. The 15-oxo derivatives usually have reduced biological activity compared to the 15-hydroxyl molecule. PGR further reduces the 13,14 5 double bond of the 15-oxo compound which typically leads to a further decrease in biological activity. PGR is a monomer with a molecular weight of approximately 36 kDa. The enzyme requires NADH or NADPH as a cofactor with a preference for NADH. The 15-oxo derivatives of prostaglandins PGE1, PGE2, and PGE2a are all substrates for PGR; however, the non-derivatized prostaglandins (i.e., PGE, PGE, and PGE, are not substrates (Ensor, C.M. et al. (1998) Biochem. J. 330:103-108).

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15-PGDH and PGR also catalyze the metabolism of lipoxin A4 (LXA4). Lipoxins (LX) are autacoids, lipids produced at the sites of localized inflammation, which down-regulate polymorphonuclear leukocyte (PMN) function and promote resolution of localized trauma. Lipoxin production is stimulated by the administration of aspirin in that cells displaying cyclooxygenase II (COX II) that has been acetylated by aspirin and cells that possess 5-lipoxygenase (5-LO) interact 15 and produce lipoxin. 15-PGDH generates 15-oxo-LXA4 with PGR further converting the 15-oxo compound to 13,14-dihydro-15-oxo-LXA4 (Clish, C.B. et al. (2000) J. Biol. Chem. 275:25372-25380). This finding suggests a broad substrate specificity of the prostaglandin dehydrogenases and has implications for these enzymes in drug metabolism and as targets for therapeutic intervention to regulate inflammation.

The GMC (glucose-methanol-choline) oxidoreductase family of enzymes was defined based on sequence alignments of Drosophila melanogaster glucose dehydrogenase, Escherichia coli choline dehydrogenase, Aspergillus niger glucose oxidase, and Hansenula polymorpha methanol oxidase. Despite their different sources and substrate specificities, these four flavoproteins are homologous, being characterized by the presence of several distinctive sequence and structural features. Each molecule contains a canonical ADP-binding, beta-alpha-beta mononucleotide-binding motif close to the amino terminus. This fold comprises a four-stranded parallel beta-sheet sandwiched between a three-stranded antiparallel beta-sheet and alpha-helices. Nucleotides bind in similar positions relative to this chain fold (Cavener, D.R. (1992) J. Mol. Biol. 223:811-814; Wierenga, R.K. et al. (1986) J. Mol. Biol. 187:101-107). Members of the GMC oxidoreductase family also share a consensus sequence near the central region of the polypeptide. Additional members of the GMC oxidoreductase family include cholesterol oxidases from Brevibacterium sterolicum and Streptomyces; and an alcohol dehydrogenase from Pseudomonas oleovorans (Cavener, supra; Henikoff, S. and J.G. Henikoff (1994) Genomics 19:97-107; van Beilen, J.B. et al. (1992) Mol.

Microbiol, 6:3121-3136).

IMP dehydrogenase and GMP reductase are two oxidoreductases which share many regions of sequence similarity. IMP dehydrogenase (BC 1.1.1.205) catalyes the NAD-dependent reduction of IMP (inosine monophosphate) into XMP (xanthine monophosphate) as part of *de novo* GTP biosynthesis (Collart, F.R. and E. Huberman (1988) J. Biol. Chem. 263:15769-15772). GMP reductase catalyzes the NADPH-dependent reductive deamination of GMP into IMP, helping to maintain the intracellular balance of adenine and guanine nucleotides (Andrews, S.C. and J.R. Guest (1988) Biochem. J. 255:35-43).

Pyridine nucleotide-disulphide oxidoreductases are FAD flavoproteins involved in the transfer of reducing equivalents from FAD to a substrate. These flavoproteins contain a pair of redox-active cysteines contained within a consensus sequence which is characteristic of this protein family (Kurlyan, J. et al. (1991) Nature 352:172-174). Members of this family of oxidoreductases include glutathione reductase (EC 1.6.4.2); thioredoxin reductase of higher eukaryotes (EC 1.6.4.5); trypanothione reductase (EC 1.6.4.8); lipoamide dehydrogenase (EC 1.8.1.4), the E3 component of alpha-ketoacid dehydrogenase complexes; and mercuric reductase (EC 1.16.1.1).

Transferases

Transferases are enzymes that catalyze the transfer of molecular groups. The reaction may involve an oxidation, reduction, or cleavage of covalent bonds, and is often specific to a substrate or to particular sites on a type of substrate. Transferases participate in reactions essential to such functions as synthesis and degradation of cell components, and regulation of cell functions including cell signaling, cell proliferation, inflammation, apoptosis, secretion and excretion. Transferases are involved in key steps in disease processes involving these functions. Transferases are frequently classified according to the type of group transferace. For example, methyl transferases transfer one-carbon methyl groups, amino transferases transfer nitrogenous amino groups, and similarly denominated enzymes transfer aldehyde or ketone, acyl, glycosyl, alkyl or aryl, isoprenyl, saccharyl, phosphorous-containing, sulfur-containing, or selenium-containing groups, as well as small enzymatic groups such as Coerzyme A.

Acyl transferases include peroxisomal camitine octanoyl transferase, which is involved in the fatty acid beta-oxidation pathway, and mitochondrial carnitine palmitoyl transferases, involved in fatty acid metabolism and transport. Choline O-acetyl transferase catalyzes the biosynthesis of the neurotransmitter acetylcholine. N-acyltransferase enzymes catalyze the transfer of an amino acid conjugate to an activated carboxylic group. Endogenous compounds and xenobiotics are activated by acyl-CoA synthetases in the cytosol, microsomes, and mitochondria. The acyl-CoA intermediates are

then conjugated with an amino acid (typically glycine, glutamine, or taurine, but also ornithine, arginine, histidine, serine, aspartic acid, and several dipeptides) by N-acyltransferases in the cytosol or mitochondria to form a metabolite with an amide bond. One well-characterized enzyme of this class is the bile acid-CoA:amino acid N-acyltransferase (BAT) responsible for generating the bile acid 5 conjugates which serve as detergents in the gastrointestinal tract (Falany, C.N. et al. (1994) J. Biol. Chem. 269:19375-19379; Johnson, M.R. et al. (1991) J. Biol. Chem. 266:10227-10233). BAT is also useful as a predictive indicator for prognosis of hepatocellular carcinoma patients after partial hepatectomy (Furutani, M. et al. (1996) Hepatology 24:1441-1445). Acetyltransferases

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Acetyltransferases have been extensively studied for their role in histone acetylation. Histone acetylation results in the relaxing of the chromatin structure in eukaryotic cells, allowing transcription factors to gain access to promoter elements of the DNA templates in the affected region of the genome (or the genome in general). In contrast, histone deacetylation results in a reduction in transcription by closing the chromatin structure and limiting access of transcription factors. To this 15 end, a common means of stimulating cell transcription is the use of chemical agents that inhibit the deacetylation of histones (e.g., sodium butyrate), resulting in a global (albeit artifactual) increase in gene expression. The modulation of gene expression by acetylation also results from the acetylation of other proteins, including but not limited to, p53, GATA-1, MyoD, ACTR, TFIIE, TFIIF and the high mobility group proteins (HMG). In the case of p53, acetylation results in increased DNA binding, 20 leading to the stimulation of transcription of genes regulated by p53. The prototypic histone acetylase (HAT) is Gcn5 from Saccharomyces cerevisiae. Gcn5 is a member of a family of acetylases that includes Tetrahymena p55, human Gcn5, and human p300/CBP. Histone acetylation is reviewed in (Cheung, W.L. et al. (2000) Curr. Opin. Cell Biol. 12:326-333 and Berger, S.L (1999) Curr. Opin. Cell Biol. 11:336-341). Some acetyltransferase enzymes possess the alpha/beta hydrolase fold (Center of 25 Applied Molecular Engineering Inst. of Chemistry and Biochemistry - University of Salzburg, http://predict.sanger.ac.uk/irbm-course97/Docs/ms/) common to several other major classes of enzymes, including but not limited to, acetylcholinesterases and carboxylesterases (Structural Classification of Proteins, http://scop.mrc-lmb.cam.ac.uk/scop/index.html).

N-acetyltransferases are cytosolic enzymes which utilize the cofactor acetyl-coenzyme A 30 (acetyl-CoA) to transfer the acetyl group to aromatic amines and hydrazine containing compounds. In humans, there are two highly similar N-acetyltransferase enzymes, NAT1 and NAT2; mice appear to have a third form of the enzyme, NAT3. The human forms of N-acetyltransferase have independent regulation (NAT1 is widely-expressed, whereas NAT2 is in liver and gut only) and overlapping

substrate preferences. Both enzymes appear to accept most substrates to some extent, but NAT1 does prefer some substrates (para-aminobenzoic acid, para-aminosalicylic acid, sulfamethoxazole, and sulfamilamide), while NAT2 prefers others (isoniazid, hydralazine, procainamide, dapsone, aminoglutethimide, and sulfamethazine). A recently isolated human gene, tubedown-1, is homologous to the yeast NAT-1 N-acetyltransferases and encodes a protein associated with acetyltransferase activity. The expression patterns of tubedown-1 suggest that it may be involved in regulating vascular and hematopoietic development (Gendron, R.L. et al. (2000) Dev. Dyn. 218:300-315).

Amino transferases comprise a family of pyridoxal 5'-phosphate (PLP) -dependent enzymes that catalyze transformations of amino acids. Amino transferases play key roles in protein synthesis and degradation, and they contribute to other processes as well. For example, GABA aminotransferase (GABA-T) catalyzes the degradation of GABA, the major inhibitory amino acid neurotransmitter. The activity of GABA-T is correlated to neuropsychiatric disorders such as alcoholism, epilepsy, and Alzheimer's disease (Sherif, F.M. and S.S. Ahmed (1995) Clin. Biochem. 28:145-154). Other members of the family include pyruvate aminotransferase, branched-chain amino acid aminotransferase, tyrosine aminotransferase, aminotransferase, alanine:glyoxylate aminotransferase (AGT), and kynurenine aminotransferase (Vacca, R.A. et al. (1997) J. Biol. Chem. 272:21932-21937). Kynurenine aminotransferase catalyzes the irreversible transamination of the L-tryptophan metabolite L-kynurenine to form kynurenic acid. The enzyme may also catalyzes the reversible transamination reaction between L-2-aminoadipate and 2-oxoglutarate to produce 20 2-oxoadipate and L-glutamate. Kynurenic acid is a putative modulator of glutamatergic neurotransmission, thus a deficiency in kynurenine aminotransferase may be associated with pleiotropic effects (Buchli, R. et al. (1995) J. Biol. Chem. 270:29330-29335).

Glycosyl transferases include the mammalian UDP-glucouronosyl transferases, a family of membrane-bound microsomal enzymes catalyzing the transfer of glucouronic acid to lipophilic substates in reactions that play important roles in detoxification and excretion of drugs, carcinogens, and other foreign substances. Another mammalian glycosyl transferase, mammalian UDP-galactose-ceramide galactosyl transferase, catalyzes the transfer of galactose to ceramide in the synthesis of galactoerebrosides in myelin membranes of the nervous system. The UDP-glycosyl transferases share a conserved signature domain of about 50 amino acid residues (PROSITE: PDOC00359, http://expasy.hcuge.ch/sprot/prosite.html).

Methyl transferases are involved in a variety of pharmacologically important processes.

Nicotinamide N-methyl transferase catalyzes the N-methylation of nicotinamides and other pyridines, an important step in the cellular handling of drugs and other foreign compounds. Phenylethanolamine

N-methyl transferase catalyzes the conversion of noradrenalin to adrenalin. 6-O-methylguanine-DNA methyl transferase reverses DNA methylation, an important step in carcinogenesis. Uroporphyrin-III C-methyl transferase, which catalyzes the transfer of two methyl groups from S-adenosyl-Lmethionine to uroporphyrinogen III, is the first specific enzyme in the biosynthesis of cobalamin, a 5 dietary enzyme whose uptake is deficient in pernicious anemia. Protein-arginine methyl transferases catalyze the posttranslational methylation of arginine residues in proteins, resulting in the mono- and dimethylation of arginine on the guanidino group. Substrates include histones, myelin basic protein, and heterogeneous nuclear ribonucleoproteins involved in mRNA processing, splicing, and transport. Protein-arginine methyl transferase interacts with proteins upregulated by mitogens, with proteins 10 involved in chronic lymphocytic leukemia, and with interferon, suggesting an important role for methylation in cytokine receptor signaling (Lin, W.-J. et al. (1996) J. Biol. Chem. 271:15034-15044; Abramovich, C. et al. (1997) EMBO J. 16:260-266; and Scott, H. S. et al. (1998) Genomics 48:330-340).

Phospho transferases catalyze the transfer of high-energy phosphate groups and are important 15 in energy-requiring and -releasing reactions. The metabolic enzyme creatine kinase catalyzes the reversible phosphate transfer between creatine/creatine phosphate and ATP/ADP. Glycocyamine kinase catalyzes phosphate transfer from ATP to guanidoacetate, and arginine kinase catalyzes phosphate transfer from ATP to arginine. A cysteine-containing active site is conserved in this family (PROSITE: PDOC00103).

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Prenyl transferases are heterodimers, consisting of an alpha and a beta subunit, that catalyze the transfer of an isoprenyl group. The Ras farnesyltransferase (FTase) enzyme transfers a farnesyl moiety from cytosolic farnesylpyrophosphate to a cysteine residue at the carboxyl terminus of the Ras oncogene protein. This modification is required to anchor Ras to the cell membrane so that it can perform its role in signal transduction. FTase inhibitors block Ras function and demonstrate antitumor 25 activity\_(Buolamwini, J.K. (1999) Curr. Opin. Chem. Biol. 3:500-509). Ftase, which shares structural similarity with geranylgeranyl transferase, or Rab GG transferase, prenylates Rab proteins, allowing them to perform their roles in regulating vesicle transport (Seabra, M.C. (1996) J. Biol. Chem. 271:14398-14404).

Saccharyl transferases are glycating enzymes involved in a variety of metabolic processes. 30 Oligosaccharyl transferase-48, for example, is a receptor for advanced glycation endproducts, which accumulate in vascular complications of diabetes, macrovascular disease, renal insufficiency, and Alzheimer's disease (Thornalley, P. J. (1998) Cell Mol. Biol. (Noisy-Le-Grand) 44:1013-1023).

Coenzyme A (CoA) transferase catalyzes the transfer of CoA between two carboxylic acids.

Succinyl CoA:3-oxoacid CoA transferase, for example, transfers CoA from succinyl-CoA to a recipient such as acetoacetate. Acetoacetate is essential to the metabolism of ketone bodies, which accumulate in tissues affected by metabolic disorders such as diabetes (PROSITE: PDOC00980).

Transglutaminase transferases (Tgases) are Ca<sup>2+</sup> dependent enzymes capable of forming

5 isopeptide bonds by catalyzing the transfer of the γ-carboxy group from protein-bound glutamine to the
ε-amino group of protein-bound lysine residues or other primary amines. Tgases are the enzymes
responsible for the cross-linking of cornified envelope (CE), the highly insoluble protein structure on
the surface of corneccytes, into a chemically and mechanically resistant protein polymer. Seven
known human Tgases have been identified. Individual transglutaminase gene products are specialized

10 in the cross-linking of specific proteins or tissue structures, such as factor XIIIa which stabilizes the
fibrin clot in hemostasis, prostrate transglutaminase which functions in semen coagulation, and tissue
transglutaminase which is involved in GTP-binding in receptor signaling. Four (Tgases 1, 2, 3, and X)
are expressed in terminally differentiating epithelia such as the epidermis. Tgases are critical for the
proper cross-linking of the CE as seen in the pathology of patients suffering from one form of the skin

15 diseases referred to as congenital ichthyosis which has been linked to mutations in the keratinocyte
transglutaminase (TG<sub>R</sub>) gene (Nemes, Z. et al. (1999) Proc. Natl. Acad. Sci. U.S.A. 96:8402-8407,
Aeschlimann, D. et al. (1998) J. Biol. Chem. 273:3452-3460.)

## **Hydrolases**

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Hydrolases are a class of enzymes that catalyze the cleavage of various covalent bonds in a substrate by the introduction of a molecule of water. The reaction involves a nucleophilic attack by the water molecule's oxygen atom on a target bond in the substrate. The water molecule is split across the target bond, breaking the bond and generating two product molecules. Hydrolases participate in reactions essential to such functions as synthesis and degradation of cell components, and for regulation of cell functions including cell signaling, cell proliferation, inflamation, apoptosis, secretion and excretion. Hydrolases are involved in key steps in disease processes involving these functions. Hydrolytic enzymes, or hydrolases, may be grouped by substrate specificity into classes including phosphatases, peptidases, lysophospholipases, phosphodiesterases, glycosidases, glyoxalases, aminohydrolases, carboxylesterases, sulfatases, phosphohydrolases, nucleotidases, lysozymes, and many others.

Phosphatases hydrolytically remove phosphate groups from proteins, an energy-providing step that regulates many cellular processes, including intracellular signaling pathways that in turn control cell growth and differentiation, cell-cell contact, the cell cycle, and oncogenesis.

Peptidases, also called proteases, cleave peptide bonds that form the backbone of peptide or protein chains. Proteolytic processing is essential to cell growth, differentiation, remodeling, and homeostasis as well as inflammation and the immune response. Since typical protein half-lives range from hours to a few days, peptidases are continually cleaving precursor proteins to their active form, removing signal sequences from targeted proteins, and degrading aged or defective proteins. Peptidases function in bacterial, parasitic, and viral invasion and replication within a host. Examples of peptidases include trypsin and chymotrypsin (components of the complement cascade and the blood-clotting cascade) lysosomal cathepsins, calpains, pepsin, renin, and chymosin (Beynon, R.J. and J.S. Bond (1994) Proteolytic Enzymes: A Practical Approach, Oxford University Press, New York, 10 NY, pp. 1-5).

Lysophospholipases (LPLs) regulate intracellular lipids by catalyzing the hydrolysis of ester bonds to remove an acyl group, a key step in lipid degradation. Small LPL isoforms, approximately 15-30 kD, function as hydrolases; larger isoforms function both as hydrolases and transacylases. A particular substrate for LPLs, lysophosphatidylcholine, causes lysis of cell membranes. LPL activity is regulated by signaling molecules important in numerous pathways, including the inflammatory response.

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The phosphodiesterases catalyze the hydrolysis of one of the two ester bonds in a phosphodiester compound. Phosphodiesterases are therefore crucial to a variety of cellular processes. Phosphodiesterases include DNA and RNA endo- and exo-nucleases, which are essential to cell growth and replication as well as protein synthesis. Endonuclease V (deoxyinosine 3'-endonuclease) is an example of a type II site-specific deoxyribonuclease, a putative DNA repair enzyme that cleaves DNAs containing hypoxanthine, uracil, or mismatched bases. Escherichia coli endonuclease V has been shown to cleave DNA containing deoxyxanthosine at the second phosphodiester bond 3' to deoxyxanthosine, generating a 3'-hydroxyl and a 5'-phosphoryl group at the nick site (He. B. et al. (2000) Mutat. Res. 459:109-114). It has been suggested that Escherichia coli endonuclease V plays a role in the removal of deaminated guanine, i.e., xanthine, from DNA, thus helping to protect the cell against the mutagenic effects of nitrosative deamination (Schouten, K.A. and B. Weiss (1999) Mutat. Res. 435:245-254). In eukaryotes, the process of tRNA splicing requires the removal of small tRNA introns that interrupt the anticodon loop 1 base 3' to the anticodon. This process requires the stepwise action of an endonuclease, a ligase, and a phosphotransferase (Hong, L. et al. (1998) Science 280:279-284). Ribonuclease P (RNase P) is a ubiquitous RNA processing endonuclease that is required for generating the mature tRNA 5'-end during the tRNA splicing process. This is accomplished through the catalysis of the cleavage of P-3'O bonds to produce 5'-phosphate and 3'-hydroxyl end groups at

a specific site on pre-tRNA. Catalysis by RNase P is absolutely dependent on divalent cations such as Mg<sup>2+</sup> or Mn<sup>2+</sup> (Kyzz, J.C. et al. (2000) Curr. Opin. Chem. Biol. 4:553-558). Substrate recognition mechanisms of RNase P are well conserved among eukaryotes and bacteria (Fabbri, S, et al. (1998) Science 280:284-286). In Saccharomyces cerevisiae, POP1 ('processing of precursor RNAs') 5 encodes a protein component of both RNase P and RNase MRP, another RNA processing protein.

Mutations in yeast POP1 are lethal (Lygerou, Z. et al. (1994) Genes Dev. 8:1423-1433). Another phosphodiesterase, acid sphingomyelinase, hydrolyzes the membrane phospholipid sphingomyelin to ceramide and phosphorylcholine. Phosphorylcholine functions in synthesis of phosphatidylcholine. which is involved in intracellular signaling pathways. Ceramide is an essential precursor for the generation of gangliosides, membrane lipids found in high concentration in neural tissue. Defective acid sphingomyelinase phosphodiesterase leads to Niemann-Pick disease.

"Glycosidases catalyze the cleavage of hemiacetyl bonds of glycosides, which are compounds that contain one or more sugar. Mammalian lactase-phlorizin hydrolase, for example, is an intestinal enzyme that splits lactose. Mammalian beta-galactosidase removes the terminal galactose from gangliosides, glycoproteins, and glycosaminoglycans, and deficiency of this enzyme is associated with a gangliosidosis known as Morquio disease type B (PROSITE PCDOC00910). Vertebrate lysosomal alpha-glucosidase, which hydrolyzes glycogen, maltose, and isomaltose, and vertebrate intestinal sucrase-isomaltase, which hydrolyzes sucrose, maltose, and isomaltose, are widely distributed members of this family with highly conserved sequences at their active sites.

The glyoxylase system is involved in gluconeogenesis, the production of glucose from storage compounds in the body. It consists of glyoxylase I, which catalyzes the formation of S-Dlactoylglutathione from methyglyoxal, a side product of triose-phosphate energy metabolism, and glyoxylase II, which hydrolyzes S-D-lactoylglutathione to D-lactic acid and reduced glutathione. Glyoxylases are involved in hyperglycemia, non-insulin-dependent diabetes mellitus, the detoxification 25 of bacterial toxins, and in the control of cell proliferation and microtubule assembly.

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NG,NG-dimethylarginine dimethylaminohydrolase (DDAH) is an enzyme that hydrolyzes the endogenous nitric oxide synthase (NOS) inhibitors, NG-monomethyl-arginine and NG,NG-dimethyl-Larginine, to L-citrulline. Inhibiting DDAH can cause increased intracellular concentration of NOS inhibitors to levels sufficient to inhibit NOS. Therefore, DDAH inhibition may provide a method of 30 NOS inhibition, and changes in the activity of DDAH could play a role in pathophysiological alterations in nitric oxide generation (MacAllister, R.J. et al. (1996) Br. J. Pharmacol. 119:1533-1540). DDAH was found in neurons displaying cytoskeletal abnormalities and oxidative stress in Alzheimer's disease. In age-matched control cases, DDAH was not found in neurons. This suggests that oxidative stress-

and nitric oxide-mediated events play a role in the pathogenesis of Alzheimer's disease (Smith, M.A. et al. (1998) Free Rad. Biol. Med. 25:898-902).

Acyl-CoA thioesterase is another member of the carboxylesterase family (Alexson, S.E. et al. (1993) Eur. J. Biochem. 214:719-727). Evidence suggests that acyl-CoA thioesterase has a regulatory role in steroidogenic tissues (Finkielstein, C. et al. (1998) Eur. J. Biochem. 256:60-66).

The alpha/beta hydrolase protein fold is common to several hydrolases of diverse phylogenetic origin and catalytic function. Enzymes with the alpha/beta hydrolase fold have a common core structure consisting of eight beta-sheets connected by alpha-helices. The most conserved structural feature of this fold is the loops of the nucleophile-histidine-acid catalytic triad. The histidine in the catalytic triad is completely conserved, while the nucleophile and acid loops accommodate more than one type of amino acid (Ollis, D.L. et al. (1992) Protein Eng. 5:197-211).

Sulfatases are members of a highly conserved gene family that share extensive sequence homology and a high degree of structural similarity. Sulfatases catalyze the cleavage of sulfate esters.

To perform this function, sulfatases undergo a unique post-translational modification in the endoplasmic reticulum that involves the oxidation of a conserved cysteine residue. A human disorder called multiple sulfatase deficiency is due to a defect in this post-translational modification step, leading to inactive sulfatases (Recksiek, M. et al. (1998) J. Biol. Chem. 273:6096-6103).

Phosphohydrolases are enzymes that hydrolyze phosphate esters. Some phosphohydrolases
contain a mutT domain signature sequence. MutT is a protein involved in the GO system responsible
for removing an exidatively damaged form of guanine from DNA. A region of about 40 amino acid
residues, found in the N-terminus of mutT, is also found in other proteins, including some
phosphohydrolases (PROSITE PDOC00695).

Serine hydrolases are a large functional class of hydrolytic enzymes that contain a serine
residue in their active site. This class of enzymes contains proteinases, esterases, and lipases which
hydrolyze a variety of substrates and, therefore, have different biological roles. Proteins in this
superfamily can be further grouped into subfamilies based on substrate specificity or amino acid
similarities (Puente, X.S. and C. Lopez-Otin (1995) J. Biol. Chem. 270:12926-12932).

Neuropathy target esterase (NTE) is an integral membrane protein present in all neurons and in some non-neural-cell types of vertebrates. NTE is involved in a cell-signaling pathway controlling interactions between neurons and accessory glial cells in the developing nervous system. NTE has serine esterase activity and efficiently catalyses the hydrolysis of phenyl valerate (PV) in vitro, but its physiological substrate is unknown. NTE is not related to either the major serine esterase family, which includes acetylcholinesterase, nor to any other known serine hydrolases. NTE contains at least

two functional domains: an N-terminal putative regulatory domain and a C-terminal effector domain which contains the esterase activity and is, in part, conserved in proteins found in bacteria, yeast, nematodes and insects. NTE's effector domain contains three predicted transmembrane segments, and the active-site serine residue lies at the center of one of these segments. The isolated recombinant domain shows PV hydrolase activity only when incorporated into phospholipid liposomes. NTE's esterase activity is largely redundant in adult vertebrates, but organophosphates which react with NTE in vivo initiate unknown events which lead to a neuropathy with degeneration of long axons. These neuropathic organophosphates leave a negatively charged group covalently attached to the active-site serine residue, which causes a toxic gain of function in NTE (Glynn, P. (1999))

10 Biochem. J. 344:625-631). Further, the Drosophila neurodegeneration gene swiss-cheese encodes a neuronal protein involved in glia-neuron interaction and is homologous to the above human NTE (Moser, M. et al. (2000) Mech. Dev. 90:279-282).

Chitinases are chitin-degrading enzymes present in a variety of organisms and participate in processes including cell wall remodeling, defense and catabolism. Chitinase activity has been found in 15 human serum, leukocytes, granulocytes, and in association with fertilized oocytes in mammals (Escott, G.M. (1995) Infect. Immunol. 63:4770-4773; DeSouza, M.M. (1995) Endrocrinology 136:2485-2496). Glycolytic and proteolytic molecules in humans are associated with tissue damage in lung diseases and with increased tumorigenicity and metastatic potential of cancers (Mulligan, M.S. (1993) Proc. Natl. Acad. Sci. 90:11523-11527; Matrisian, L.M. (1991) Am. J. Med. Sci. 302:157-162; Witty, J.P. (1994) 20 Cancer Res. 54:4805-4812). The discovery of a human enzyme with chitinolytic activity is noteworthy given the lack of endogenous chitin in the human body (Raghavan, N. (1994) Infect. Immun. 62:1901-1908). However, there is a group of mammalian proteins that share homology with chitinases from various non-mammalian organisms, such as bacteria, fungi, plants, and insects. The members of this family differ in their ability to hydrolyze chitin or chitin-like substrates. Some of the mammalian 25 members of the family, such as a bovine whey chitotriosidase and human cartilage proteins which do not demonstrate specific chitinolytic activity, are expressed in association with tissue remodeling events (Rejman, J.J. (1988) Biochem. Biophys. Res. Commun. 150:329-334, Nyirkos, P. (1990) Biochem. J. 268:265-268). Elevated levels of human cartilage proteins have been reported in the synovial fluid and cartilage of patients with rheumatoid arthritis, a disease which produces a severe degradation of the cartilage and a proliferation of the synovial membrane in the affected joints (Hakala, B.E. (1993) J. Biol. Chem. 268:25803-25810).

A small subclass of hydrolases acting on ether bonds includes the thioether hydrolases. Sadenosyl-L-homocysteine hydrolase, also known as AdoHcyase or SAHH (PROSITE PDOC00603;

EC 3.3.1.1), is a thioether hydrolase first described in rat liver extracts as the activity responsible for the reversible hydrolysis of S-adenosyl-L-homocysteine (AdoHey) to adenosine and homocysteine (Sganga, M.W. et al. (1992) PNAS 89:6328-6332). SAHH is a cytosolic enzyme that has been found in all cells that have been tested, with the exception of Escherichia coli and certain related bacteria 5 (Walker, R.D. et al. (1975) Can. J. Biochem. 53:312-319; Shimizu, S. et al. (1988) FEMS Microbiol. Lett. 51:177-180; Shimizu, S. et al. (1984) Eur. J. Biochem. 141:385-392). SAHH activity is dependent on NAD+ as a cofactor. Deficiency of SAHH is associated with hypermethioninemia (Online Mendelian Inheritance in Man (OMIM) #180960 Hypermethioninemia), a pathologic condition characterized by neonatal cholestasis, failure to thrive, mental and motor retardation, facial dysmorphism with abnormal hair and teeth, and myocaridopathy (Labrune, P. et al. (1990) J. Pediat. 117:220-2260.

Another subclass of hydrolases includes those enzymes which act on carbon-nitrogen (C-N) bonds other than peptide bonds. To this subclass belong those enzymes hydrolyzing amides, amidines, and other C-N bonds. This subclass is further subdivided on the basis of substrate specificity such as 15 linear amides, cyclic amides, linear amidines, cyclic amidines, nitriles and other compounds. A hydrolase belonging to the sub-subclass of enzymes acting on the cyclic amidines is adenosine deaminase (ADA). ADA catalyzes the breakdown of adenosine to inosine. ADA is present in many mammalian tissues, including placenta, muscle, lung, stomach, digestive diverticulum, spleen, erythrocytes, thymus, seminal plasma, thyroid, T-cells, bone marrow stem cells, and liver. A subclass-20 of ADAs, ADAR, act on RNA and are classified as RNA editases. An ADAR from Drosophila, dADAR, expressed in the developing nervous system, may act on para voltage-gated Na+ channel transcripts in the central nervous system (Palladino, M.J. et al. (2000) RNA 6:1004-1018). ADA deficiency causes profound lymphopenia with severe combined immunodeficiency (SCID). Cells from patients with ADA deficiency contain low, sometimes undetectable, amounts of ADA catalytic activity and ADA protein. ADA deficiency stems from genetic mutations in the ADA gene (Hershfield, M.S. (1998) Semin. Hematol. 4:291-298). Metabolic consequences of ADA deficiency are associated with defects in alveogenesis, pulmonary inflammation, and airway obstruction (Blackburn, M.R. et al. (2000) J. Exp. Med. 192:159-170).

Pancreatic ribonucleases (RNase) are pyrimidine-specific endonucleases found in high
quantity in the pancreas of certain mammalian taxa and of some reptiles (Beintema, J.J. et al (1988)
Prog. Biophys. Mol. Biol. 51:165-192). Proteins in the mammalian pancreatic RNase superfamily are
noncytosolic endonucleases that degrade RNA through a two-step transphosphorolytic-hydrolytic
reaction (Beintema, J.J. et al. (1986) Mol. Biol. Evol. 3:262-275). Specifically, the enzymes are

involved in endonucleolytic cleavage of 3'-phosphomononucleotides and 3'-phosphooligonucleotides ending in C-P or U-P with 2',3'-cyclic phosphate intermediates. Ribonucleases can unwind the DNA helix by complexing with single-stranded DNA; the complex arises by an extended multi-site cation-anion interaction between lysine and arginine residues of the enzyme and phosphate groups of 5 the nucleotides. Some of the enzymes belonging to this family appear to play a purely digestive role, whoreas others exhibit potent and unusual biological activities (D'Alessio, G. (1993) Trends Cell Biol. 3:106-109). Proteins belonging to the pancreatic RNase family include: bovine seminal vesicle and brain ribonucleases; kidney non-secretory ribonucleases (Beintema, J.J. et al (1986) FEBS Lett. 194:338-343); liver-type ribonucleases (Rosenberg, H.F. et al. (1989) PNAS U.S.A. 86:4460-4464); angiogenin, which induces vascularisation of normal and malignant tissues; eosinophil cationic protein (Hofsteenge, J. et al. (1989) Biochemistry 28:9806-9813), a cytotoxin and helminthotoxin with ribonuclease activity; and frog liver ribonuclease and frog sialic acid-binding lectin. The sequences of pancreatic RNases contain 4 conserved disulfide bonds and 3 amino acid residues involved in the catalytic activity.

ADP-ribosylation is a reversible post-translational protein modification in which an ADP-ribose moiety is transferred from β-NAD to a target amino acid such as arginine or cysteine. ADP-ribosylation hydrolases regenerate arginine by removing ADP-ribose from the protein, completing the ADP-ribosylation cycle (Moss, J. et al. (1997) Adv. Exp. Med. Biol. 419:25-33). ADP-ribosylation is a well-known reaction among bacterial toxins. Cholera toxin, for example, disrupts the adenylyl cyclase system by ADP-ribosylating the α-subunit of the stimulatory G-protein, causing an increase in intracellular cAMP (Moss, J. and M. Vaughan (Eds) (1990) ADP-ribosylating Toxins and G-Proteins: Insights into Signal Transduction, American Society for Microbiology, Washington, D.C.). ADP-ribosylation may also have a regulatory function in eukaryotes, affecting such processes as cytoskeletal assembly (Zhou, H. et al. (1996) Arch. Biochem. Biophys. 334:214-222) and cell proliferation in evitotxic T-cells (Wans, J. et al. (1996) J. Immunol. 156:2819-2827).

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Nucleotidases catalyze the formation of free nucleosides from nucleotides. The cytosolic nucleotidase cN-I (5' nucleotidase-I) cloned from pigeon heart catalyzes the formation of adenosine from AMP generated during ATP hydrolysis (Sala-Newby, G.B. et al. (1999) J. Biol. Chem. 274:17789-17793). Increased adenosine concentration is thought to be a signal of metabolic stress, and adenosine receptors mediate effects including vasodilation, decreased stimulatory neuron firing and ischemic preconditioning in the heart (Schrader, J. (1990) Circulation 81:389-391; Rubino, A. et al. (1992) Eur. J. Pharmacol. 220:95-98; de Jong, J.W. et al. (2000) Pharmacol. Ther. 87:141-149). Deficiency of pyrimidine 5'-nucleotidase can result in hereditary hemolytic anemia (OMIM #266120).

The lysozyme c superfamily consists of conventional lysozymes c, calcium-binding lysozymes c, and α-lactalbumin (Prager, E.M. and P. Jolles (1996) EXS 75:9-31). The proteins in this superfamily have 35-40% sequence homology and share a common three-dimensional fold, but can have different functions. Lysozymes c are ubiquitous in a variety of tissues and secretions and can lyse the cell walls of certain bacteria (McKenzie, H.A. (1996) EXS 75:365-409). Alpha-lactalbumin is a metallo-protein that binds calcium and participates in the synthesis of lactose (Iyer, L.K. and P.K. Qasba (1999) Protein Eng. 12:129-139). Alpha-lactalbumin occurs in mammalian milk and colostrum (McKenzie, supra).

Lysozymes catalyze the hydrolysis of certain mucopolysaccharides of bacterial cell walls,

specifically, the beta (1-4) glycosidic linkages between N-acetylmuramic acid and Nacetylglucosamine, and cause bacterial lysis. Lysozymes occur in diverse organisms including viruses,
birds, and mammals. In humans, lysozymes are found in spleen, lung, kidney, white blood cells,
plasma, saliva, milk, tears, and cartilage (OMIM #153450 Lysozyme; Weaver, L.H. et al. (1985) J.

Mol. Biol. 184:739-741). Lysozyme c functions in ruminants as a digestive enzyme, releasing proteins

from ingested bacterial cells, and may perform the same function in human newborns (Braun, O.H. et
al. (1995) Klin. Pediatr. 207:4-7).

The two known forms of lysozymes, chicken-type and goose-type, were originally isolated from chicken and goose egg white, respectively. Chicken-type and goose-type lysozymes have similar three-dimensional structures, but different amino acid sequences (Nakano, T. and T. Graf (1991)

Biochim. Biophys. Acta 1090:273-276). In chickens, both forms of lysozyme are found in neutrophil granulocytes (heterophils), but only chicken-type lysozyme is found in egg white. Generally, chicken-type lysozyme mRNA is found in both adherent monocytes and macrophages and nonadherent promyelocytes and granulocytes as well as in cells of the bone marrow, spleen, bursa, and oviduct. Goose-type lysozyme mRNA is found in non-adherent cells of the bone marrow and lung. Several isozymes have been found in rabbits, including leukocytic, gastrointestinal, and possibly lymphoepithelial forms (OMIM #153450, supra; Nakano and Graf, supra; and GenBank GI 1310929). A human lysozyme gene encoding a protein similar to chicken-type lysozyme has been cloned (Yoshimura, K. et al. (1988) Biochem. Biophys. Res. Commun. 150:794-801). A consensus motif featuring regularly spaced cysteine residues has been derived from the lysozyme C enzymes of various species (PROSITE PS00128). Lysozyme C shares about 40% amino acid sequence identity with a-lactalbumin.

Lysozymes have several disease associations. Lysozymuria is observed in diabetic nephropathy (Shima, M. et al. (1986) Clin. Chem. 32:1818-1822), endemic nephropathy (Bruckner, I.

et al. (1978) Med. Interne. 16:117-125), urinary tract infections (Heidegger, H. (1990) Minerva Ginecol. 42:243-250), and acute monocytic leukemia (Shaw, M.T. (1978) Am. J. Hematol. 4:97-103). Nakano and Graf (supra) suggested a role for lysozyme in host defense systems. Older rabbits with an inherited lysozyme deficiency show increased susceptibility to infections, such as subcutaneous 5 abscesses (OMIM #153450, supra). Human lysozyme gene mutations cause hereditary systemic amyloidosis, a rare autosomal dominant disease in which amyloid deposits form in the viscera. including the kidney, adrenal glands, spleen, and liver. This disease is usually fatal by the fifth decade. The amyloid deposits contain variant forms of lysozyme. Renal amyloidosis is the most common and potentially the most serious form of organ involvement (Pepys, M.B. et al. (1993) Nature 362:553-557; 10 OMIM #105200 Familial Visceral Amyloidosis; Cotran, R.S. et al. (1994) Robbins Pathologic Basis of Disease, W.B. Saunders Company, Philadelphia PA, pp. 231-238). Increased levels of lysozyme and lactate have been observed in the cerebrospinal fluid of patients with bacterial meningitis (Ponka, A. et al. (1983) Infection 11:129-131). Acute monocytic leukemia is characterized by massive lysozymuria (Den Tandt, W.R. (1988) Int. J. Biochem, 20:713-719).

Lyases

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Lyases are a class of enzymes that catalyze the cleavage of C-C, C-O, C-N, C-S, C-(halide), P-O, or other bonds without hydrolysis or oxidation to form two molecules, at least one of which contains a double bond (Stryer, L. (1995) Biochemistry, W.H. Freeman and Co., New York NY, 20 p.620). Under the International Classification of Enzymes (Webb, E. C. (1992) Enzyme Nomenclature 1992: Recommendations of the Nomenclature Committee of the International Union of Biochemistry and Molecular Biology on the Nomenclature and Classification of Enzymes, Academic Press, San Diego CA), lyases form a distinct class designated by the numeral 4 in the first digit of the enzyme number (i.e., EC 4.x.x.x).

Further classification of lyases reflects the type of bond cleaved as well as the nature of the cleaved group. The group of C-C lyases includes carboxyl-lyases (decarboxylases), aldehyde-lyases (aldolases), oxo-acid-lyases, and other lyases. The C-O lyase group includes hydro-lyases, lyases acting on polysaccharides, and other lyases. The C-N lyase group includes ammonia-lyases, amidinelyases, amine-lyases (deaminases), and other lyases. Lyases are critical components of cellular 30 biochemistry, with roles in metabolic energy production, including fatty acid metabolism and the tricarboxylic acid cycle, as well as other diverse enzymatic processes.

One important family of lyases are the carbonic anhydrases (CA), also called carbonate dehydratases, which catalyze the hydration of carbon dioxide in the reaction H<sub>2</sub>O + CO<sub>2</sub> ≠ HCO<sub>3</sub> +

H\*. CA accelerates this reaction by a factor of over 106 by virtue of a zinc ion located in a deep cleft about 15Å below the protein's surface and co-ordinated to the imidazole groups of three His residues. Water bound to the zinc ion is rapidly converted to HCO<sub>3</sub>.

Eight enzymatic and evolutionarily related forms of carbonic anhydrase are currently known to 5 exist in humans: three cytosolic isozymes (CAI, CAII, and CAIII), two membrane-bound forms (CAIV and CAVII), a mitochondrial form (CAV), a secreted salivary form (CAVI) and a yet uncharacterized isozyme (PROSITE PDOC00146 Eukaryotic-type carbonic anhydrases signature). Though the isoenzymes CAI, CAII, and bovine CAIII have similar secondary structures and polypeptide-chain folds, CAI has 6 tryptophans, CAII has 7 and CAIII has 8 (Boren, K. et al. (1996) 10 Protein Sci. 5:2479-2484). CAll is the predominant CA isoenzyme in the brain of mammals.

CAs participate in a variety of physiological processes that involve pH regulation, CO, and HCO3 transport, ion transport, and water and electrolyte balance. For example, CAII contributes to H+ secretion by gastric parietal cells, by renal tubular cells, and by osteoclasts that secrete H+ to acidify the bone-resorbing compartment. In addition, CAII promotes HCO3 secretion by pancreatic 15 duct cells, cilary body epithelium, choroid plexus, salivary gland acinar cells, and distal colonal epithelium, thus playing a role in the production of pancreatic juice, aqueous humor, cerebrospinal fluid, and saliva, and contributing to electrolyte and water balance. CAII also promotes CO, exchange in proximal tubules in the kidney, in erythrocytes, and in lung. CAIV has roles in several tissues: it facilitates HCO3 reabsorption in the kidney; promotes CO2 flux in tissues including brain, skeletal 20 muscle, and heart muscle; and promotes CO2 exchange from the blood to the alveoli in the lung. CAVI probably plays a role in pH regulation in saliva, along with CAII, and may have a protective effect in the esophagus and stomach. Mitochondrial CAV appears to play important roles in gluconeogenesis and ureagenesis, based on the effects of CA inhibitors on these pathways. (Sly, W.S. and P.Y. Hu (1995) Ann. Rev. Biochem. 64:375-401.)

A number of disease states are marked by variations in CA activity. Mutations in CAII which lead to CAII deficiency are the cause of osteopetrosis with renal tubular acidosis (OMIM #259730 Osteopetrosis with Renal Tubular Acidosis). The concentration of CAII in the cerebrospinal fluid (CSF) appears to mark disease activity in patients with brain damage. High CA concentrations have been observed in patients with brain infarction. Patients with transient ischemic attack, multiple 30 sclerosis, or epilepsy usually have CAII concentrations in the normal range, but higher CAII levels have been observed in the CSF of those with central nervous system infection, dementia, or trigeminal neuralgia (Parkkila, A.K. et al. (1997) Eur. J. Clin. Invest. 27:392-397). Colonic adenomas and adenocarcinomas have been observed to fail to stain for CA, whereas non-neoplastic controls showed

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CAI and CAII in the cytoplasm of the columnar cells lining the upper half of colonic crypts. The neoplasms show staining patterns similar to less mature cells lining the base of normal crypts (Gramlich T.L. et al. (1990) Arch. Pathol. Lab. Med. 114:415-419).

Therapeutic interventions in a number of diseases involve altering CA activity. CA inhibitors such as acetazolamide are used in the treatment of glaucoma (Stewart, W.C. (1999) Curr. Opin. Opthamol. 10:99-108), essential tremor and Parkinson's disease (Uitti, R.J. (1998) Geriatrics 53:46-48, 53-57), intermittent ataxia (Singhvi, J.P. et al. (2000) Neurology India 48:78-80), and altitude related illnesses (Klocke, D.L. et al. (1998) Mayo Clin. Proc. 73:988-992).

CA activity can be particularly useful as an indicator of long-term disease conditions, since the
enzyme reacts relatively slowly to physiological changes. CAI and zinc concentrations have been
observed to decrease in hyperthyroid Graves' disease (Yoshida, K. (1996) Tohoku J. Exp. Med.
178:345-356) and glycosylated CAI is observed in diabetes mellitus (Kondo, T. et al. (1987) Clin.
Chim. Acta 166:227-236). A positive correlation has been observed between CAI and CAII reactivity
and endometriosis (Brinton, D.A. et al. (1996) Ann. Clin. Lab. Sci. 26:409-420; D'Cruz, O.J. et al.
(1996) Fertil. Steril. 66:547-556).

Another important member of the lyase family is omithine decarboxylase (ODC), the initial rate-limiting enzyme in polyamine biosynthesis. ODC catalyses the transformation of ornithine into putrescine in the reaction L-ornithine → putrescine + CO<sub>2</sub>. Polyamines, which include putrescine and the subsequent metabolic pathway products spermidine and spermine, are ubiquitous cell components essential for DNA synthesis, cell differentiation, and proliferation. Thus the polyamines play a key role in tumor proliferation (Medina, M.A. et al. (1999) Biochem. Pharmacol. 57:1341-1344).

ODC is a pyridoxal-5'-phosphate (PLP)-dependent enzyme which is active as a homodimer.

Conserved residues include those at the PLP binding site and a stretch of glycine residues thought to
be part of a substrate binding region (PROSITE PDOC00685 Orn/DAP/Arg decarboxylase family 2
25 signatures). Mammalian ODCs also contain PEST regions, sequence fragments enriched in proline,
glutamic acid, serine, and threonine residues that act as signals for intracellular degradation (Medina et
al., supra).

Many chemical carcinogens and tumor promoters increase ODC levels and activity. Several known oncogenes may increase ODC levels by enhancing transcription of the ODC gene, and ODC itself may act as an oncogene when expressed at very high levels. A high level of ODC is found in a number of precancerous conditions, and elevation of ODC levels has been used as part of a screen for tumor-promoting compounds (Pegg, A.E. et al. (1995) J. Cell. Biochem. Suppl. 22:132-138).

Inhibitors of ODC have been used to treat tumors in animal models and human clinical trials,

and have been shown to reduce development of tumors of the bladder, brain, esophagus, gastrointestinal tract, lung, oral cavity, mammary gland, stomach, skin and trachea (Pegg et al., supra; McCann, P.P. and A.E. Pegg (1992) Pharmac. Ther. 54:195-215). ODC also shows promise as a target for chemoprevention (Pegg et al., supra). ODC inhibitors have also been used to treat infections by African trypanosomes, malaria, and Pneumocystis carinii, and are potentially useful for

treatment of autoimmune diseases such as lupus and rheumatoid arthritis (McCann and Pegg, supra).

Another family of pyridoxal-dependent decarboxylases are the group II decarboxylases. This family includes glutamate decarboxylase (GAD) which catalyzes the decarboxylation of glutamate into the neurotransmitter GABA; histidine decarboxylase (HDC), which catalyzes the decarboxylation of histidine to histamine; aromatic-L-amino-acid decarboxylase (DDC), also known as L-dopa decarboxylase or tryptophan decarboxylase, which catalyzes the decarboxylation of tryptophan to tryptamine and also acts on 5-hydroxy-tryptophan and dihydroxyphenylalanine (L-dopa); and cysteine sulfinic acid decarboxylase (CSD), the rate-limiting enzyme in the synthesis of taurine from cysteine (PROSITE PDOC00329 DDC/GAD/HDC/TyrDC pyridoxal-phosphate attachment site). Taurine is an abundant sulfonic amino acid in brain and is thought to act as an osmoregulator in brain cells (Bitoun, M. and M. Tappaz (2000) J. Neurochem. 75:919-924).

#### Isomerases

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Isomerases are a class of enzymes that catalyze geometric or structural changes within a molecule to form a single product. This class includes racemases and epimerases, cis-transisomerases, intramolecular oxidoreductases, intramolecular transferases (mutases) and intramolecular lyases. Isomerases are critical components of cellular biochemistry with roles in metabolic energy production including glycolysis, as well as other diverse enzymatic processes (Stryer, supra, pp.483-507).

Racemases are a subset of isomerases that catalyze inversion of a molecule's configuration around the asymmetric carbon atom in a substrate having a single center of asymmetry, thereby interconverting two racemers. Epimerases are another subset of isomerases that catalyze inversion of configuration around an asymmetric carbon atom in a substrate with more than one center of symmetry, thereby interconverting two epimers. Racemases and epimerases can act on amino acids and derivatives, hydroxy acids and derivatives, and carbohydrates and derivatives. The interconversion of UDP-galactose and UDP-glucose is catalyzed by UDP-galactose-4'-epimerase. Proper regulation and function of this epimerase is essential to the synthesis of glycoproteins and glycolipids. Elevated blood galactose levels have been correlated with UDP-galactose-4'-epimerase.

deficiency in screening programs of infants (Gitzelmann, R. (1972) Helv. Paediat. Acta 27:125-130).

Correct folding of newly synthesized proteins is assisted by molecular chaperones and folding catalysts, two unrelated groups of helper molecules. Chaperones suppress non-productive side reactions by stoichiometric binding to folding intermediates, whereas folding enzymes catalyze some of the multiple folding steps that enable proteins to attain their final functional configurations (Kern, G. et al. (1994) FEBS Lett. 348:145-148). One class of folding enzymes, the peptidyl prolyl cis-trans isomerases (PPlases), isomerizes certain proline imidic bonds in what is considered to be a rate limiting step in protein maturation and export. PPlases catalyze the cis to trans isomerization of certain proline imidic bonds in proteins. There are three evolutionarily unrelated families of PPlases: the cyclophilins, the FK506 binding proteins, and the newly characterized parvulin family (Rahfeld, J.U. et al. (1994) FEBS Lett. 352:180-184).

The cyclophilins (CvP) were originally identified as major receptors for the immunosuppressive drug cyclosporin A (CsA), an inhibitor of T-cell activation (Handschumacher, R.E. et al. (1984) Science 226:544-547; Harding, M.W. et al. (1986) J. Biol. Chem. 261:8547-8555). 15 Thus, the peptidyl-prolyl isomerase activity of CyP may be part of the signaling pathway that leads to T-cell activation. Subsequent work demonstrated that CyP's isomerase activity is essential for correct protein folding and/or protein trafficking, and may also be involved in assembly/disassembly of protein complexes and regulation of protein activity. For example, in Drosophila, the CyP NinaA is required for correct localization of rhodopsins, while a mammalian CyP (Cyp40) is part of the Hsp90/Hsp70 20 complex that binds steroid receptors. The mammalian CyP (CypA) has been shown to bind the gag protein from human immunodeficiency virus 1 (HIV-1), an interaction that can be inhibited by cyclosporin. Since cyclosporin has potent anti-HIV-1 activity, CypA may play an essential function in HTV-1 replication. Finally, Cvp40 has been shown to bind and inactivate the transcription factor c-Myb, an effect that is reversed by cyclosporin. This effect implicates CyP in the regulation of 25 transcription, transformation, and differentiation (Bergsma, D.J. et al (1991) J. Biol. Chem. 266:23204-23214; Hunter, T. (1998) Cell 92;141-143; and Leverson, J.D. and S.A. Ness (1998) Mol. Cell. 1:203-211).

One of the major rate limiting steps in protein folding is the thiol:disulfide exchange that is necessary for correct protein assembly. Although incubation of reduced, unfolded proteins in buffers with defined ratios of oxidized and reduced thiols can lead to native conformation, the rate of folding is slow and the attainment of native conformation decreases proportionately with the size and number of cysteines in the protein. Certain cellular compartments such as the endoplasmic reticulum of eukaryotes and the periplasmic space of prokaryotes are maintained in a more oxidized state than the

surrounding cytosol. Correct disulfide formation can occur in these compartments, but at a rate that is insufficient for normal cell processes and inadequate for synthesizing secreted proteins. The protein disulfide isomerases, thioredoxins and glutaredoxins are able to catalyze the formation of disulfide bonds and regulate the redox environment in cells to enable the necessary thiol:disulfide exchanges

(Loferer, H. (1995) J. Biol. Chem. 270:26178-26183).

Each of these proteins has somewhat different functions, but all belong to a group of disulfidecontaining redox proteins that contain a conserved active-site sequence and are ubiquitously distributed
in eukaryotes and prokaryotes. Protein disulfide isomerases are found in the endoplasmic reticulum of
eukaryotes and in the periplasmic space of prokaryotes. They function by exchanging their own

disulfide for a thiol in a folding peptide chain. In contrast, the reduced thioredoxins and glutaredoxins
are generally found in the cytoplasm and function by directly reducing disulfides in the substrate
proteins.

Oxidoreductases can be isomerases as well. Oxidoreductases catalyze the reversible transfer of electrons from a substrate that becomes oxidized to a substrate that becomes reduced. This class of enzymes includes dehydrogenases, hydroxylases, oxidases, oxygenases, peroxidases, and reductases. Proper maintenance of oxidoreductase levels is physiologically important. For example, genetically-linked deficiencies in lipoamide dehydrogenase can result in lactic acidosis (Robinson, B. H. et al. (1977) Pediat. Res. 11:1198-1202).

Another subgroup of isomerases are the transferases (or mutases). Transferases transfer a

20 chemical group from one compound (the donor) to another compound (the acceptor). The types of
groups transferred by these enzymes include acyl groups, amino groups, phosphate groups
(phosphotransferases or phosphomutases), and others. The transferase carnitine palmitoyltransferase
is an important component of fatty acid metabolism. Genetically-linked deficiencies in this transferase
can lead to myopathy (Scriver, C. et al. (1995) The Metabolic and Molecular Basis of Inherited
25 Disease, McGraw-Hill, New York NY, pp. 1501-1533).

Yet another subgroup of isomerases are the topoisomerases. Topoisomerases are enzymes
that affect the topological state of DNA. For example, defects in topoisomerases or their regulation
can affect normal physiology. Reduced levels of topoisomerase II have been correlated with some of
the DNA processing defects associated with the disorder ataxia-telangiectasia (Singh, S.P. et al.

30 (1988) Nucleic Acids Res. 16:3919-3929).

## Ligases

Ligases catalyze the formation of a bond between two substrate molecules. The process

involves the hydrolysis of a pyrophosphate bond in ATP or a similar energy donor. Ligases are classified based on the nature of the type of bond they form, which can include carbon-oxygen, carbon-sulfur, carbon-nitrogen, carbon-carbon and phosphoric ester bonds.

Ligases forming carbon-oxygen bonds include the aminoacyl-transfer RNA (tRNA) 5 synthetases which are important RNA-associated enzymes with roles in translation. Protein biosynthesis depends on each amino acid forming a linkage with the appropriate tRNA. The aminoacyl-tRNA synthetases are responsible for the activation and correct attachment of an amino acid with its cognate tRNA. The 20 aminoacyl-tRNA synthetase enzymes can be divided into two structural classes, and each class is characterized by a distinctive topology of the catalytic domain. 10 Class I enzymes contain a catalytic domain based on the nucleotide-binding "Rossman fold". Class II enzymes contain a central catalytic domain, which consists of a seven-stranded antiparallel ß-sheet motif, as well as N- and C- terminal regulatory domains. Class II enzymes are separated into two groups based on the heterodimeric or homodimeric structure of the enzyme; the latter group is further subdivided by the structure of the N- and C-terminal regulatory domains (Hartlein, M. and S. Cusack, 15 (1995) J. Mol. Evol. 40:519-530). Autoantibodies against aminoacyl-tRNAs are generated by patients with dermatomyositis and polymyositis, and correlate strongly with complicating interstitial lung disease (ILD). These antibodies appear to be generated in response to viral infection, and coxsackie virus has been used to induce experimental viral myositis in animals.

Ligases forming carbon-sulfur bonds (acid-thiol ligases) mediate a large number of cellular biosynthetic intermediary metabolism processes involving intermolecular transfer of carbon atom-containing substrates (carbon substrates). Examples of such reactions include the tricarboxylic acid cycle, synthesis of fatty acids and long-chain phospholipids, synthesis of alcohols and aldehydes, synthesis of intermediary metabolites, and reactions involved in the amino acid degradation pathways. Some of these reactions require input of energy, usually in the form of conversion of ATP to either 25 ADP or AMP and pyrophosphate.

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In many cases, a carbon substrate is derived from a small molecule containing at least two carbon atoms. The carbon substrate is often covalently bound to a larger molecule which acts as a carbon substrate carrier molecule within the cell. In the biosynthetic mechanisms described above, the carrier molecule is coenzyme A. Coenzyme A (CoA) is structurally related to derivatives of the 30 nucleotide ADP and consists of 4'-phosphopantetheine linked via a phosphodiester bond to the alpha phosphate group of adenosine 3',5'-bisphosphate. The terminal thiol group of 4'-phosphopantetheine acts as the site for carbon substrate bond formation. The predominant carbon substrates which utilize CoA as a carrier molecule during biosynthesis and intermediary metabolism in the cell are acetyl,

succinyl, and propionyl moieties, collectively referred to as acyl groups. Other carbon substrates include enoyl lipid, which acts as a fatty acid oxidation intermediate, and carnitine, which acts as an acetyl-CoA flux regulator/mitochondrial acyl group transfer protein. Acyl-CoA and acetyl-CoA are synthesized in the cell by acyl-CoA synthetase and acetyl-CoA synthetase, respectively.

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Activation of fatty acids is mediated by at least three forms of acyl-CoA synthetase activity: i) acetyl-CoA synthetase, which activates acetate and several other low molecular weight carboxylic acids and is found in muscle mitochondria and the cytosol of other tissues; ii) medium-chain acyl-CoA synthetase, which activates fatty acids containing between four and eleven carbon atoms (predominantly from dietary sources), and is present only in liver mitochondria; and iii) acyl CoA synthetase, which is specific for long chain fatty acids with between six and twenty carbon atoms, and is found in microsomes and the mitochondria. Proteins associated with acyl-CoA synthetase activity have been identified from many sources including bacteria, yeast, plants, mouse, and man. The activity of acyl-CoA synthetase may be modulated by phosphorylation of the enzyme by cAMP-dependent protein kinase.

Ligases forming carbon-nitrogen bonds include amide synthases such as glutamine synthetase (glutamate-ammonia ligase) that catalyzes the amination of glutamic acid to glutamine by ammonia using the energy of ATP hydrolysis. Glutamine is the primary source for the amino group in various amide transfer reactions involved in de novo pyrimidine nucleotide synthesis and in purine and pyrimidine ribonucleotide interconversions. Overexpression of glutamine synthetase has been observed in primary liver cancer (Christa, L. et al. (1994) Gastroent. 106:1312-1320).

Acid-amino-acid ligases (peptide synthases) are represented by the ubiquitin conjugating enzymes which are associated with the ubiquitin conjugation system (UCS), a major pathway for the degradation of cellular proteins in eukaryotic cells and some bacteria. The UCS mediates the elimination of abnormal proteins and regulates the half-lives of important regulatory proteins that control cellular processes such as gene transcription and cell cycle progression. In the UCS pathway, proteins targeted for degradation are conjugated to ubiquitin (Ub), a small heat stable protein. Ub is first activated by a ubiquitin-activating enzyme (E1), and then transferred to one of several Ub-conjugating enzymes (E2). E2 then links the Ub molecule through its C-terminal glycine to an internal lysine (acceptor lysine) of a target protein. The ubiquitinated protein is then recognized and degraded by proteasome, a large, multisubunit proteolytic enzyme complex, and ubiquitin is released for reutilization by ubiquitin protease. The UCS is implicated in the degradation of mitotic cyclic kinases, oncoproteins, tumor suppressor genes such as p53, viral proteins, cell surface receptors associated with signal transduction, transcriptional regulators, and mutated or damaged proteins (Ciechanover, A.

(1994) Cell 79:13-21).

Cyclo-ligases and other carbon-nitrogen ligases comprise various enzymes and enzyme
complexes that participate in the *de novo* pathways of purine and pyrimidine biosynthesis. Because
these pathways are critical to the synthesis of nucleotides for replication of both RNA and DNA,
many of these enzymes have been the targets of clinical agents for the treatment of cell proliferative
disorders such as cancer and infectious diseases.

Purine biosynthesis occurs de novo from the amino acids glycine and glutamine, and other small molecules. Three of the key reactions in this process are catalyzed by a trifunctional enzyme composed of glycinamide-ribonucleotide synthetase (GARS), aminoimidazole ribonucleotide synthetase (GARS), aminoimidazole ribonucleotide synthetase (GART). Together these three enzymes combine ribosylamine phosphate with glycine to yield phosphoribosyl aminoimidazole, a precursor to both adenylate and guanylate nucleotides. This trifunctional protein has been implicated in the pathology of Downs synthema (Aimi, J. et al. (1990) Nucleic Acid Res. 18:6665-6672).

Adenylosuccinate synthetase catalyzes a later step in purine biosynthesis that converts inosinic acid to adenylosuccinate, a key step on the path to ATP synthesis. This enzyme is also similar to another carbon-nitrogen ligase, argininosuccinate synthetase, that catalyzes a similar reaction in the urea cycle (Powell, S.M. et al. (1992) FEBS Lett. 303:4-10).

Adenylosuccinate synthetase, adenylosuccinate lyase, and AMP deaminase may be considered as a functional unit, the purine nucleotide cycle. This cycle converts AMP to inosine monophosphate (IMP) and reconverts IMP to AMP via adenylosuccinate, thereby producing NH<sub>3</sub> and forming fumarate from aspartate. In muscle, the purine nucleotide cycle functions, during intense exercise, in the regeneration of ATP by pulling the adenylate kinase reaction in the direction of ATP formation and by providing Krebs cycle intermediates. In kidney, the purine nucleotide cycle accounts for the release of NH<sub>3</sub> under normal acid-base conditions. In brain, the purine nucleotide cycle may contribute to ATP recovery. Adenylosuccinate lyase deficiency provokes psychomotor retardation, often accompanied by autistic features (Van den Berghe, G. et al. (1992) Prog Neurobiol.: 39:547-561). A marked imbalance in the enzymic pattern of purine metabolism is linked with transformation and/or progression in cancer cells. In rat hepatomas the specific activities of the anabolic enzymes, IMP dehydrogenase, GMP synthetase, adenylosuccinate synthetase, adenylosuccinase, AMP deaminase and amidophosphoribosyltransferase, increased to 13.5-, 3.7-, 3.1-, 1.8-, 5.5- and 2.8-fold, respectively, of those in normal liver (Weber, G. (1983) Clin. Biochem. 16:57-63).

Like the de novo biosynthesis of purines, de novo synthesis of the pyrimidine nucleotides

uridylate and cytidylate also arises from a common precursor, in this instance the nucleotide orotidylate derived from orotate and phosphoribosyl pyrophosphate (PPRP). Again a trifunctional enzyme comprising three carbon-nitrogen ligases plays a key role in the process. In this case the enzymes aspartate transcarbamylase (ATCase), carbamyl phosphate synthetase II, and dihydrocrotase 5 (DHOase) are encoded by a single gene called CAD. Together these three enzymes combine the initial reactants in pyrimidine biosynthesis, glutamine, CO2 and ATP to form dihydrocrotate, the precursor to orotate and orotidylate (Iwahana, H. et al. (1996) Biochem. Biophys. Res. Commun. 219:249-255). Further steps then lead to the synthesis of uridine nucleotides from orotidylate. Cytidine nucleotides are derived from uridine-5'-triphosphate (UTP) by the amidation of UTP using 10 glutamine as the amino donor and the enzyme CTP synthetase. Regulatory mutations in the human CTP synthetase are believed to confer multi-drug resistance to agents widely used in cancer therapy (Yamauchi, M. et al. (1990) EMBO J. 9:2095-2099).

Ligases forming carbon-carbon bonds include the carboxylases acetyl-CoA carboxylase and pyruvate carboxylase. Acetyl-CoA carboxylase catalyzes the carboxylation of acetyl-CoA from CO2 15 and H<sub>2</sub>O using the energy of ATP hydrolysis. Acetyl-CoA carboxylase is the rate-limiting enzyme in the biogenesis of long-chain fatty acids. Two isoforms of acetyl-CoA carboxylase, types I and types II, are expressed in human in a tissue-specific manner (Ha, J. et al. (1994) Eur, J. Biochem. 219:297-306). Pyruvate carboxylase is a nuclear-encoded mitochondrial enzyme that catalyzes the conversion of pyruvate to oxaloacetate, a key intermediate in the citric acid cycle.

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Ligases forming phosphoric ester bonds include the DNA ligases involved in both DNA replication and repair. DNA ligases seal phosphodiester bonds between two adjacent nucleotides in a DNA chain using the energy from ATP hydrolysis to first activate the free 5'-phosphate of one nucleotide and then react it with the 3'-OH group of the adjacent nucleotide. This resealing reaction is used in DNA replication to join small DNA fragments called "Okazaki" fragments that are transiently 25 formed in the process of replicating new DNA, and in DNA repair. DNA repair is the process by which accidental base changes, such as those produced by oxidative damage, hydrolytic attack, or uncontrolled methylation of DNA, are corrected before replication or transcription of the DNA can occur. Bloom's syndrome is an inherited human disease in which individuals are partially deficient in DNA ligation and consequently have an increased incidence of cancer (Alberts et al., supra, p. 247).

Pantothenate synthetase (D-pantoate; beta-alanine ligase (AMP-forming); EC 6.3.2.1) is the last enzyme of the pathway of pantothenate (vitamin B(5)) synthesis. It catalyzes the condensation of pantoate with beta-alanine in an ATP-dependent reaction. The enzyme is dimeric, with two well-defined domains per protomer: the N-terminal domain, a Rossmann fold, contains the active site

cavity, with the C-terminal domain forming a hinged lid. The N-terminal domain is structurally very similar to class I aminoacyl-tRNA synthetases and is thus a member of the cytidylyltransferase superfamily (von Delft, F. et al. (2000) Structure (Camb) 9:439-450).

Farnesyl diphosphate synthase (FPPS) is an essential enzyme that is required both for

cholesterol synthesis and protein prenylation. The enzyme catalyzes the formation of farnesyl
diphosphate from dimethylallyl diphosphate and isopentyl diphosphate. FPPS is inhibited by nitrogencontaining biphosphonates, which can lead to the inhibition of osteoclast-mediated bone resorption by
preventing protein prenylation (Dunford, J.E. et al. (2001) J. Pharmacol. Exp. Ther. 296:235-242).

5-aminolevulinate synthase (ALAS; delta-aminolevulinate synthase; EC 2.3.1.37) catalyzes

the rate-limiting step in heme biosynthesis in both crythroid and non-crythroid tissues. This enzyme is
unique in the heme biosynthetic pathway in being encoded by two genes, the first encoding ALAS1,
the non-crythroid specific enzyme which is ubiquitously expressed, and the second encoding ALAS2,
which is expressed exclusively in crythroid cells. The genes for ALAS1 and ALAS2 are located,
respectively, on chromosome 3 and on the X chromosome. Defects in the gene encoding ALAS2

result in X-linked sideroblastic anemia. Elevated levels of ALAS are seen in acute hepatic porphyrias
and can be lowered by zine mesoporphyrin.

# Drug Metabolizing Enzymes (DMEs)

The metabolism of a drug and its movement through the body (pharmacokinetics) are
important in determining its effects, toxicity, and interactions with other drugs. The three processes
governing pharmacokinetics are the absorption of the drug, distribution to various tissues, and
elimination of drug metabolites. These processes are intimately coupled to drug metabolism, since a
variety of metabolic modifications alter most of the physicochemical and pharmacological properties of
drugs, including solubility, binding to receptors, and excretion rates. The metabolic pathways which
modify drugs also accept a variety of naturally occurring substrates such as steroids, fatty acids,
prostaglandins, leukotrienes, and vitamins. The enzymes in these pathways are therefore important
sites of biochemical and pharmacological interaction between natural compounds, drugs, carcinogens,
mutagens, and xenobiotics. It has long been appreciated that inherited differences in drug metabolism
lead to drastically different levels of drug efficacy and toxicity among individuals. Advances in
pharmacogenomics research, of which DMEs constitute an important part, are promising to expand
the tools and information that can be brought to bear on questions of drug efficacy and toxicity (See
Evans, W.E. and R.V. Relling (1999) Science 286:487-491). DMEs have broad substrate specificities,
unlike antibodies, for example, which are diverse and highly specific. Since DMEs metabolize a wide

variety of molecules, drug interactions may occur at the level of metabolism so that, for example, one compound may induce a DME that affects the metabolism of another compound.

Drug metabolic reactions are categorized as Phase I, which prepare the drug molecule for functioning and further metabolism, and Phase II, which are conjugative. In general, Phase I reaction products are partially or fully inactive, and Phase II reaction products are the chief excreted species. However, Phase I reaction products are sometimes more active than the original administered drugs; this metabolic activation principle is exploited by pro-drugs (e.g. L-dopa). Additionally, some nontoxic compounds (e.g. aflatoxin, benzo[a]pyrene) are metabolized to toxic intermediates through these pathways. Phase I reactions are usually rate-limiting in drug metabolism. Prior exposure to the compound, or other compounds, can induce the expression of Phase I enzymes however, and thereby increase substrate flux through the metabolic pathways. (See Klaassen, C.D. et al. (1996) <u>Casarett and Doull's Toxicology: The Basic Science of Poisons.</u> McGraw-Hill, New York, NY, pp. 113-186; Katzung, B.G. (1995) <u>Basic and Clinical Pharmacology</u>, Appleton and Lange, Norwalk, CT, pp. 48-59; Gibson, G.G. and P. Skett (1994) <u>Introduction to Drug Metabolism</u>. Blackie Academic and

The major classes of Phase I enzymes include, but are not limited to, cytochrome P450 and flavin-containing monooxygenase. Other enzyme classes involved in Phase I-type catalytic cycles and reactions include, but are not limited to, NADPH cytochrome P450 reductase (CPR), the microsomal cytochrome b5/NADH cytochrome b5 reductase system, the ferredoxin/ferredoxin reductase redox pair, aldo/keto reductases, and alcohol dehydrogenases. The major classes of Phase II enzymes include, but are not limited to, UDP glucuronyltransferase, sulfotransferase, glutathione S-transferase, N-acyltransferase, and N-acetyl transferase.

### Cytochrome P450 and P450 catalytic cycle-associated enzymes

Members of the cytochrome P450 superfamily of enzymes catalyze the oxidative metabolism

of a variety of substrates, including natural compounds such as steroids, fatty acids, prostaglandins,
leukotrienes, and vitamins, as well as drugs, carcinogens, mutagens, and xenobiotics. Cytochromes

P450, also known as P450 heme-thiolate proteins, usually act as terminal oxidases in multi-component
electron transfer chains, called P450-containing monooxygenase systems. Specific reactions
catalyzed include hydroxylation, epoxidation, N-oxidation, sulfooxidation, N-, S-, and O-dealkylations,
desulfation, deamination, and reduction of azo, nitro, and N-oxide groups. These reactions are
involved in steroidogenesis of glucocorticoids, cortisols, estrogens, and androgens in animals;
insecticide resistance in insects; herbicide resistance and flower coloring in plants; and environmental
bioremediation by microorganisms. Cytochrome P450 actions on drugs, carcinogens, mutagens, and

xenobiotics can result in detoxification or in conversion of the substance to a more toxic product. Cytochromes P450 are abundant in the liver, but also occur in other tissues; the enzymes are located in microsomes. (See ExPASY ENZYME EC 1.14.14.1; Prosite PDOC00081 Cytochrome P450 cysteine heme-iron ligand signature; PRINTS EP450I E-Class P450 Group I signature; Graham-5 Lorence, S. and J.A. Peterson (1996) FASEB J. 10:206-214.)

Four hundred cytochromes P450 have been identified in diverse organisms including bacteria, fungi, plants, and animals (Graham-Lorence and Peterson, supra). The B-class is found in prokaryotes and fungi, while the E-class is found in bacteria, plants, insects, vertebrates, and mammals. Five subclasses or groups are found within the larger family of E-class cytochromes P450 10 (PRINTS EP450I E-Class P450 Group I signature).

All cytochromes P450 use a heme cofactor and share structural attributes. Most cytochromes P450 are 400 to 530 amino acids in length. The secondary structure of the enzyme is about 70% alpha-helical and about 22% beta-sheet. The region around the heme-binding site in the Cterminal part of the protein is conserved among cytochromes P450. A ten amino acid signature 15 sequence in this heme-iron ligand region has been identified which includes a conserved cysteine involved in binding the heme iron in the fifth coordination site. In eukaryotic cytochromes P450, a membrane-spanning region is usually found in the first 15-20 amino acids of the protein, generally consisting of approximately 15 hydrophobic residues followed by a positively charged residue. (See Prosite PDOC00081, supra; Graham-Lorence and Peterson, supra.)

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Cytochrome P450 enzymes are involved in cell proliferation and development. The enzymes have roles in chemical mutagenesis and carcinogenesis by metabolizing chemicals to reactive intermediates that form adducts with DNA (Nebert, D.W. and F.J. Gonzalez (1987) Ann. Rev. Biochem. 56:945-993). These adducts can cause nucleotide changes and DNA rearrangements that lead to oncogenesis. Cytochrome P450 expression in liver and other tissues is induced by xenobiotics 25 such as polycyclic aromatic hydrocarbons, peroxisomal proliferators, phenobarbital, and the glucocorticoid dexamethasone (Dogra, S.C. et al. (1998) Clin. Exp. Pharmacol. Physiol. 25:1-9). A cytochrome P450 protein may participate in eye development as mutations in the P450 gene CYP1B1 cause primary congenital glaucoma (OMIM #601771 Cytochrome P450, subfamily I (dioxin-inducible), polypeptide 1; CYP1B1).

Cytochromes P450 are associated with inflammation and infection. Hepatic cytochrome P450 activities are profoundly affected by various infections and inflammatory stimuli, some of which are suppressed and some induced (Morgan, E.T. (1997) Drug Metab. Rev. 29:1129-1188). Effects observed in vivo can be mimicked by proinflammatory cytokines and interferons. Autoantibodies to

two cytochrome P450 proteins were found in patients with autoimmune polyenodocrinopathycandidiasis-ectodermal dystrophy (APECED), a polyglandular autoimmune syndrome (OMIM #240300 Autoimmune polyenodocrinopathy-candidiasis-ectodermal dystrophy).

Mutations in cytochromes P450 have been linked to metabolic disorders, including congenital 5 adrenal hyperplasia, the most common adrenal disorder of infancy and childhood; pseudovitamin Ddeficiency rickets; cerebrotendinous xanthomatosis, a lipid storage disease characterized by progressive neurologic dysfunction, premature atherosclerosis, and cataracts; and an inherited resistance to the anticoagulant drugs coumarin and warfarin (Isselbacher, K.J. et al. (1994) Harrison's Principles of Internal Medicine, McGraw-Hill, Inc. New York, NY, pp. 1968-1970; Takeyama, K. et 10 al. (1997) Science 277:1827-1830; Kitanaka, S. et al. (1998) N. Engl. J. Med. 338:653-661; OMIM #213700 Cerebrotendinous xanthomatosis; and OMIM #122700 Coumarin resistance). Extremely high levels of expression of the cytochrome P450 protein aromatase were found in a fibrolamellar hepatocellular carcinoma from a boy with severe gynecomastia (feminization) (Agarwal, V.R. (1998) J. Clin. Endocrinol. Metab. 83:1797-1800).

The cytochrome P450 catalytic cycle is completed through reduction of cytochrome P450 by NADPH cytochrome P450 reductase (CPR). Another microsomal electron transport system consisting of cytochrome b5 and NADPH cytochrome b5 reductase has been widely viewed as a minor contributor of electrons to the cytochrome P450 catalytic cycle. However, a recent report by Lamb, D.C. et al. (1999; FEBS Lett. 462:283-288) identifies a Candida albicans cytochrome P450 20 (CYP51) which can be efficiently reduced and supported by the microsomal cytochrome b5/NADPH cytochrome b5 reductase system. Therefore, there are likely many cytochromes P450 which are supported by this alternative electron donor system.

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Cytochrome b5 reductase is also responsible for the reduction of oxidized hemoglobin (methemoglobin, or ferrihemoglobin, which is unable to carry oxygen) to the active hemoglobin 25 (ferrohemoglobin) in red blood cells. Methemoglobinemia results when there is a high level of oxidant drugs or an abnormal hemoglobin (hemoglobin M) which is not efficiently reduced. Methemoglobinemia can also result from a hereditary deficiency in red cell cytochrome b5 reductase (Reviewed in Mansour, A. and A.A. Lurie (1993) Am. J. Hematol. 42:7-12).

Members of the cytochrome P450 family are also closely associated with vitamin D synthesis 30 and catabolism. Vitamin D exists as two biologically equivalent prohormones, ergocalciferol (vitamin D2), produced in plant tissues, and cholecalciferol (vitamin D3), produced in animal tissues. The latter form, cholecalciferol, is formed upon the exposure of 7-dehydrocholesterol to near ultraviolet light (i.e., 290-310 nm), normally resulting from even minimal periods of skin exposure to sunlight (reviewed in

Miller, W.L. and A.A. Portale (2000) Trends Endocrinol. Metab. 11:315-319).

Both prohormone forms are further metabolized in the liver to 25-hydroxyvitamin D (25(OH)D) by the enzyme 25-hydroxylase. 25(OH)D is the most abundant precursor form of vitamin D which must be further metabolized in the kidney to the active form, 1a,25-dihydroxyvitamin D

by which must be tuturer measonized in the kidney to the active form, 1α,25-dihydroxylasmin D
 (1α,25(OH)<sub>2</sub>D), by the enzyme 25-hydroxyvitamin D 1α-hydroxylase (1α-hydroxylase). Regulation of 1α,25(OH)<sub>2</sub>D production is primarily at this final step in the synthetic pathway. The activity of 1α-hydroxylase depends upon several physiological factors including the circulating level of the enzyme product (1α,25(OH)<sub>2</sub>D) and the levels of parathyroid hormone (PTH), calcitonin, insulin, calcium, phosphorus, growth hormone, and prolactin. Furthermore, extrarenal 1α-hydroxylase activity
 has been reported, suggesting that tissue-specific, local regulation of 1α,25(OH)<sub>2</sub>D production may also be biologically important. The catalysis of 1α,25(OH)<sub>2</sub>D to 24,25-dihydroxyvitamin D (24,25(OH)<sub>2</sub>D), involving the enzyme 25-hydroxyvitation D 24-hydroxylase (24-hydroxylase), also occurs in the kidney. 24-hydroxylase can also use 25(OH)D as a substrate (Shinki, T. et al. (1997) Proc. Natl. Acad. Sci. U.S.A. 94:12920-12925; Miller and Portale, supra; and references within).

Vitamin D 25-hydroxylase, 1\(\alpha\)-hydroxylase, and 24-hydroxylase are all NADPH-dependent, type I (mitochondrial) cytochrome P450 enzymes that show a high degree of homology with other members of the family. Vitamin D 25-hydroxylase also shows a broad substrate specificity and may also perform 26-hydroxylation of bile acid intermediates and 25, 26, and 27-hydroxylation of cholesterol (Dilworth, F.J. et al. (1995) J. Biol. Chem. 270:16766-16774; Miller and Portale, supra; and references within).

The active form of vitamin D (1α,25(OH)<sub>2</sub>D) is involved in calcium and phosphate homeostasis and promotes the differentiation of myeloid and skin cells. Vitamin D deficiency resulting from deficiencies in the enzymes involved in vitamin D metabolism (e.g., 1α-hydroxylase) causes hypocalcemia, hypophosphatemia, and vitamin D-dependent (sensitive) rickets, a disease characterized by loss of bone density and distinctive clinical features, including bandy or bow leggedness accompanied by a waddling gait. Deficiencies in vitamin D 25-hydroxylase cause cerebrotendinous xanthomatosis, a lipid-storage disease characterized by the deposition of cholesterol and cholestanol in the Achilles' tendons, brain, lungs, and many other tissues. The disease presents with progressive neurologic dysfunction, including postpubescent cerebellar ataxia, atherosclerosis, and cataracts.

Vitamin D 25-hydroxylase deficiency does not result in rickets, suggesting the existence of alternative pathways for the synthesis of 25(OH)D (Griffin, J.E. and J.E. Zerwekh (1983) J. Clin. Invest. 72:1190-1199; Gamblin, G.T. et al. (1985) J. Clin. Invest. 75:954-960; and Miller and Portale, supra).

Ferredoxin and ferredoxin reductase are electron transport accessory proteins which support

at least one human cytochrome P450 species, cytochrome P450c27 encoded by the CYP27 gene (Dilworth, F.J. et al. (1996) Biochem. J. 320:267-71). A Streptomyces griseus cytochrome P450, CYP104D1, was heterologously expressed in Scherichia coti and found to be reduced by the endogenous ferredoxin and ferredoxin reductase enzymes (Taylor, M. et al. (1999) Biochem. Biophys. 5 Res. Commun. 263:838-842), suggesting that many cytochrome P450 species may be supported by the ferredoxin/ferredoxin reductase pair. Ferredoxin reductase has also been found in a model drug metabolism system to reduce actinomycin D, an antitumor antibiotic, to a reactive free radical species (Flitter, W.D. and R.P. Mason (1988) Arch. Biochem. Biophys. 267:632-639). Flaviu-containing monocoxygenase (FMO)

Flavin-containing monooxygenases oxidize the nucleophilic nitrogen, sulfur, and phosphorus heteroatom of an exceptional range of substrates. Like cytochromes P450, FMOs are microsomal and use NADPH and O<sub>2</sub>; there is also a great deal of substrate overlap with cytochromes P450. The tissue distribution of FMOs includes liver, kidney, and lung.

Isoforms of FMO in mammals include FMO1, FMO2, FMO3, FMO4, and FMO5, which are
expressed in a tissue-specific manner. The isoforms differ in their substrate specificities and
properties such as inhibition by various compounds and stereospecificity of reaction. FMOs have a 13
amino acid signature sequence, the components of which span the N-terminal two-thirds of the
sequences and include the FAD binding region and the FATGY motif found in many N-hydroxylating
enzymes (Stehr, M. et al. (1998) Trends Biochem. Sci. 23:56-57; PRINTS FMOXYGENASE Flavincontaining monoxygenase signature). Specific reactions include oxidation of nucleophilic tertiary
amines to N-oxides, secondary amines to hydroxylamines and nitrones, primary amines to
hydroxylamines and oximes, and sulfur-containing compounds and phosphines to S- and P-oxides.
Hydrazines, iodides, selenides, and boron-containing compounds are also substrates. FMOs are more
heat labile and less detergent-sensitive than cytochromes P450 in vitro though FMO isoforms vary in
thermal stability and detergent sensitivity.

FMOs play important roles in the metabolism of several drugs and xenobiotics. FMO (FMO3 in liver) is predominantly responsible for metabolizing (S)-nicotine to (S)-nicotine N-1'-oxide, which is excreted in urine. FMO is also involved in S-oxygenation of cimetidine, an H<sub>2</sub>-antagonist widely used for the treatment of gastric ulcers. Liver-expressed forms of FMO are not under the same regulatory control as cytochrome P450. In rats, for example, phenobarbital treatment leads to the induction of cytochrome P450, but the repression of FMO1.

### Lysyl oxidase

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Lysyl oxidase (lysine 6-oxidase, LO) is a copper-dependent amine oxidase involved in the

formation of connective tissue matrices by crosslinking collagen and elastin. LO is secreted as an Nglycosylated precursor protein of approximately 50 kDa and cleaved to the mature form of the enzyme by a metalloprotease, although the precursor form is also active. The copper atom in LO is involved in the transport of electrons to and from oxygen to facilitate the oxidative deamination of lysine residues 5 in these extracellular matrix proteins. While the coordination of copper is essential to LO activity, insufficient dietary intake of copper does not influence the expression of the apoenzyme. However, the absence of the functional LO is linked to the skeletal and vascular tissue disorders that are associated with dietary copper deficiency. LO is also inhibited by a variety of semicarbazides, hydrazines, and amino nitrites, as well as heparin. Beta-aminopropionitrile is a commonly used inhibitor. LO activity is increased in response to ozone, cadmium, and elevated levels of hormones released in response to local tissue trauma, such as transforming growth factor-beta, platelet-derived growth factor, angiotensin II, and fibroblast growth factor. Abnormalities in LO activity have been linked to Menkes syndrome and occipital horn syndrome. Cytosolic forms of the enzyme have been implicated in abnormal cell proliferation (reviewed in Rucker, R.B. et al. (1998) Am. J. Clin. Nutr. 15 67:996S-1002S and Smith-Mungo, L.I. and H.M. Kagan (1998) Matrix Biol. 16:387-398). Dihydrofolate reductases

Dihydrofolate reductases (DHFR) are ubiquitous enzymes that catalyze the

NADPH-dependent reduction of dihydrofolate to tetrahydrofolate, an essential step in the *de novo*synthesis of glycine and purines as well as the conversion of deoxyuridine monophosphate (dUMP) to

deoxythymidine monophosphate (dTMP). The basic reaction is as follows:

7,8-dihydrofolate + NADPH → 5,6,7,8-tetrahydrofolate + NADP+

The enzymes can be inhibited by a number of dihydrofolate analogs, including trimethroprim and methotrexate. Since an abundance of dTMP is required for DNA synthesis, rapidly dividing cells require the activity of DHFR. The replication of DNA viruses (i.e., herpesvirus) also requires high levels of DHFR activity. As a result, drugs that target DHFR have been used for cancer chemotherapy and to inhibit DNA virus replication. (For similar reasons, thymidylate synthetases are also target enzymes.) Drugs that inhibit DHFR are preferentially cytotoxic for rapidly dividing cells (or DNA virus-infected cells) but have no specificity, resulting in the indiscriminate destruction of dividing cells. Furthermore, cancer cells may become resistant to drugs such as methotrexate as a result of acquired transport defects or the duplication of one or more DHFR genes (Stryer, L (1988) <u>Biochemistry</u>. W.H Freeman and Co., Inc. New York. pp. 511-519).

#### Aldo/keto reductases

Aldo/keto reductases are monomeric NADPH-dependent oxidoreductases with broad substrate specificities (Bohren, K.M. et al. (1989) J. Biol. Chem. 264:9547-9551). These enzymes catalyze the reduction of carbonyl-containing compounds, including carbonyl-containing sugars and aromatic compounds, to the corresponding alcohols. Therefore, a variety of carbonyl-containing drugs and xenobiotics are likely metabolized by enzymes of this class.

One known reaction catalyzed by a family member, aldose reductase, is the reduction of glucose to sorbitol, which is then further metabolized to fructose by sorbitol dehydrogenase. Under normal conditions, the reduction of glucose to sorbitol is a minor pathway. In hyperglycemic states, 10 however, the accumulation of sorbitol is implicated in the development of diabetic complications (OMIM #103880 Aldo-keto reductase family 1, member B1). Members of this enzyme family are also highly expressed in some liver cancers (Cao, D. et al. (1998) J. Biol. Chem. 273:11429-11435).

Alcohol dehydrogenases

Alcohol dehydrogenases (ADHs) oxidize simple alcohols to the corresponding aldehydes.

ADH is a cytosolic enzyme, prefers the cofactor NAD\*, and also binds zinc ion. Liver contains the highest levels of ADH, with lower levels in kidney, lung, and the gastric mucosa.

Known ADH isoforms are dimeric proteins composed of 40 kDa subunits. There are five known gene loci which encode these subunits (a, b, g, p, c), and some of the loci have characterized allelic variants (b<sub>1</sub>, b<sub>2</sub>, b<sub>3</sub>, g<sub>3</sub>). The subunits can form homodimens and heterodimers; the subunit composition determines the specific properties of the active enzyme. The holoeuzymes have therefore been categorized as Class I (subunit compositions aa, ab, ag, bg, gg), Class II (pp), and Class III (cc).

Class I ADH isozymes oxidize ethanol and other small aliphatic alcohols, and are inhibited by pyrazole.

Class II isozymes prefer longer chain aliphatic and aromatic alcohols, are unable to oxidize methanol, and are not inhibited by pyrazole. Class III isozymes prefer even longer chain aliphatic alcohols (five carbons and longer) and aromatic alcohols, and are not inhibited by pyrazole.

The short-chain alcohol dehydrogenases include a number of related enzymes with a variety of substrate specificities. Included in this group are the mammalian enzymes D-beta-hydroxybutyrate dehydrogenase, (R)-3-hydroxybutyrate dehydrogenase, 15-hydroxyprostaglandin dehydrogenase, NADPH-dependent carbonyl reductase, corticosteroid 11-beta-dehydrogenase, and estradiol 17-beta-dehydrogenase, as well as the bacterial enzymes acetoacetyl-CoA reductase, glucose 1-dehydrogenase, 3-beta-hydroxysteroid dehydrogenase, 20-beta-hydroxysteroid dehydrogenase, ribitol dehydrogenase, 3-oxoacyl reductase, 2,3-dihydro-2,3-dihydroxybenzoate dehydrogenase, sorbitol-6-phosphate 2-dehydrogenase, 7-alpha-hydroxysteroid dehydrogenase, cis-1,2-dihydroxy-3,4-

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cyclohexadiene-1-carboxylate dehydrogenase, cis-toluene dihydrodiol dehydrogenase, cis-benzene glycol dehydrogenase, biphenyl-2,3-dihydro-2,3-diol dehydrogenase, N-acylmannosamine 1-dehydrogenase, and 2-deoxy-D-gluconate 3-dehydrogenase (Krozowski, Z. (1994) J. Steroid Biochem. Mol. Biol. 51:125-130; Krozowski, Z. (1992) Mol. Cell Endocrinol. 84:C25-31; and Marks, 5 A.R. et al. (1992) J. Biol. Chem. 267:15459-15463).

### Sulfotransferases

Sulfate conjugation occurs on many of the same substrates which undergo O-glucuronidation to produce a highly water-soluble sulfuric acid ester. Sulfotransferases (ST) catalyze this reaction by transferring SO<sub>3</sub>: from the cofactor 3'-phosphoadenosine-5'-phosphosulfate (PAPS) to the substrate.

ST substrates are predominantly phenols and aliphatic alcohols, but also include aromatic amines and aliphatic amines, which are conjugated to produce the corresponding sulfamates. The products of these reactions are excreted mainly in urine.

STs are found in a wide range of tissues, including liver, kidney, intestinal tract, lung, platelets, and brain. The enzymes are generally cytosolic, and multiple forms are often co-expressed. For example, there are more than a dozen forms of ST in rat liver cytosol. These biochemically characterized STs fall into five classes based on their substrate preference: arylsulfotransferase, alcohol sulfotransferase, estrogen sulfotransferase, tyrosine ester sulfotransferase, and bile salt sulfotransferase.

ST enzyme activity varies greatly with sex and age in rats. The combined effects of

developmental cues and sex-related hormones are thought to lead to these differences in ST

expression profiles, as well as the profiles of other DMEs such as cytochromes P450. Notably, the

high expression of STs in cats partially compensates for their low level of UDP glucuronyltransferase

activity.

Several forms of ST have been purified from human liver cytosol and cloned. There are two phenol sulfotransferases with different thermal stabilities and substrate preferences. The thermostable enzyme catalyzes the sulfation of phenols such as para-nitrophenol, minoxidil, and acetaminophen; the thermolabile enzyme prefers monoamine substrates such as dopamine, epinephrine, and levadopa. Other cloned STs include an estrogen sulfotransferase and an N-acetylglucosamine-6-Co-sulfotransferase. This last enzyme is illustrative of the other major role of STs in cellular biochemistry, the modification of carbohydrate structures that may be important in cellular differentiation and maturation of proteoglycans. Indeed, an inherited defect in a sulfotransferase has been implicated in macular corneal dystrophy, a disorder characterized by a failure to synthesize mature keratan sulfate proteoglycans (Nakazawa, K. et al. (1984) J. Biol. Chem. 259:13751-13757; OMIM #217800 Macular

dystrophy, corneal).

#### Galactosyltransferases

Galactosyltransferases are a subset of glycosyltransferases that transfer galactose (Gal) to the terminal N-acetylglucosamine (GlcNAc) oligosaccharide chains that are part of glycoproteins or 5 glycolipids that are free in solution (Kolbinger, F. et al. (1998) J. Biol. Chem. 273:433-440; Amado, M. et al. (1999) Biochim. Biophys. Acta 1473:35-53). Galactosyltransferases have been detected on the cell surface and as soluble extracellular proteins, in addition to being present in the Golgi. 61.3galactosyltransferases form Type I carbohydrate chains with Gal (β1-3)GlcNAc linkages. Known human and mouse \$1,3-galactosyltransferases appear to have a short cytosolic domain, a single transmembrane domain, and a catalytic domain with eight conserved regions. (Kolbinger et al., supra; and Hennet, T. et al. (1998) J. Biol. Chem. 273:58-65). In mouse UDP-galactose:β-Nacetylglucosamine β1,3-galactosyltransferase-I region 1 is located at amino acid residues 78-83, region 2 is located at amino acid residues 93-102, region 3 is located at amino acid residues 116-119, region 4 is located at amino acid residues 147-158, region 5 is located at amino acid residues 172-183, region 6 15 is located at amino acid residues 203-206, region 7 is located at amino acid residues 236-246, and region 8 is located at amino acid residues 264-275. A variant of a sequence found within mouse UDP-galactose:β-N-acetylglucosamine β1,3-galactosyltransferase-I region 8 is also found in bacterial galactosyltransferases, suggesting that this sequence defines a galactosyltransferase sequence motif (Hennet et al., supra). Recent work suggests that brainiac protein is a β1,3-galactosyltransferase 20 (Yuan, Y. et al. (1997) Cell 88:9-11; and Hennet et al., supra).

UDP-Gal:GicNAc-1,4-galactosyltransferase (-1,4-GaIT) (Sato, T. et al., (1997) EMBO J.

16:1850-1857) catalyzes the formation of Type II carbohydrate chains with Gal (β1-4)GlcNAc
linkages. As is the case with the β1,3-galactosyltransferase, a soluble form of the enzyme is formed
by cleavage of the membrane-bound form. Amino acids conserved among β1,425 galactosyltransferases include two cysteines linked through a disulfide-bond and a putative UDPgalactose-binding site in the catalytic domain (Yadav, S. and K. Brew (1990) J. Biol. Chem.
265:14163-14169; Yadav, S.P. and K. Brew (1991) J. Biol. Chem. 266:698-703; and Shaper, N.L. et
al. (1997) J. Biol. Chem. 272:31389-31399). β1,4-galactosyltransferases have several specialized
roles in addition to synthesizing carbohydrate chains on glycoproteins or glycolipids. In mammals a
30 β1,4-galactosyltransferase, as part of a heterodimer with α-lactalbumin, functions in lactating
mammary gland lactose production. A β1,4-galactosyltransferase on the surface of sperm functions
as a receptor that specifically recognizes the egg. Cell surface β1,4-galactosyltransferases also
function in cell adhesion, cell/basal lamina interaction, and normal and metastatic cell migration. (Shur,

B. (1993) Curr. Opin. Cell Biol. 5:854-863; and Shaper, J. (1995) Adv. Exp. Med. Biol. 376:95-104). Gamma-alutamyl transpeptidase

Gamma-glutamyl transpeptidases are ubiquitously expressed enzymes that initiate extracellular glutathione (GSH) breakdown by cleaving gamma-glutamyl amide bonds. The breakdown of GSH provides cells with a regional cysteine pool for biosynthetic pathways. Gamma-glutamyl transpeptidases also contribute to cellular antioxidant defenses and expression is induced by oxidative stress. The cell surface-localized glycoproteins are expressed at high levels in cancer cells. Studies have suggested that the high level of gamma-glutamyl transpeptidase activity present on the surface of cancer cells could be exploited to activate precursor drugs, resulting in high local concentrations of anti-cancer therapeutic agents (Hanigan, M.H. (1998) Chem. Biol. Interact. 111-112:333-342;
Taniguchi, N. and Y. Ikeda (1998) Adv. Enzymol. Relat. Areas Mol. Biol. 72:239-278; Chikhi, N. et al. (1999) Comp. Biochem. Physiol. B. Biochem. Mol. Biol. 122:367-380).

### Aminotransferases

Aminotransferases comprise a family of pyridoxal 5'-phosphate (PLP) -dependent enzymes
that catalyze transformations of amino acids. Aspartate aminotransferase (AspAT) is the most
extensively studied PLP-containing enzyme. It catalyzes the reversible transamination of dicarboxylic
L-amino acids, aspartate and glutamate, and the corresponding 2-oxo acids, oxalacetate and
2-oxoglutarate. Other members of the family include pyruvate aminotransferase, branched-chain
amino acid aminotransferase, tyrosine aminotransferase, aromatic aminotransferase, alanine:glyoxylate
aminotransferase (AGT), and kynurenine aminotransferase (Vacca, R.A. et al. (1997) J. Biol. Chem.
272:21932-21937).

Primary hyperoxaluria type-1 is an autosomal recessive disorder resulting in a deficiency in the liver-specific peroxisomal enzyme, alanine:glyoxylate aminotransferase-1. The phenotype of the disorder is a deficiency in glyoxylate metabolism. In the absence of AGT, glyoxylate is oxidized to oxalate rather than being transaminated to glycine. The result is the deposition of insoluble calcium oxalate in the kidneys and urinary tract, ultimately causing renal failure (Lumb, M.J. et al. (1999) J. Biol. Chem. 274:20587-20596).

Kynurenine aminotransferase catalyzes the irreversible transamination of the L-tryptophan metabolite L-kynurenine to form kynurenic acid. The enzyme may also catalyze the reversible transamination reaction between L-2-aminoadipate and 2-oxoglutarate to produce 2-oxoadipate and L-glutamate. Kynurenic acid is a putative modulator of glutamatergic neurotransmission; thus a deficiency in kynurenine aminotransferase may be associated with pleotrophic effects (Buchli, R. et al. (1995) J. Biol. Chem. 270:29330-29335).

#### Catechol-O-methyltransferase

Catechol-O-methyltransferase (COMT) catalyzes the transfer of the methyl group of Sadenosyl-L-methionine (AdoMet; SAM) donor to one of the hydroxyl groups of the catechol substrate
(e.g., L-dopa, dopamine, or DBA). Methylation of the 3'-hydroxyl group is favored over methylation
of the 4'-hydroxyl group and the membrane bound isoform of COMT is more regiospecific than the
soluble form. Translation of the soluble form of the enzyme results from utilization of an internal start
codon in a full-length mRNA (1.5 kb) or from the translation of a shorter mRNA (1.3 kb), transcribed
from an internal promoter. The proposed S<sub>N</sub>2-like methylation reaction requires Mg\*+ and is inhibited
by Ca\*+. The binding of the donor and substrate to COMT occurs sequentially. AdoMet first binds

COMT in a Mg\*+-independent manner, followed by the binding of Mg\*+ and the binding of the catechol
substrate.

The amount of COMT in tissues is relatively high compared to the amount of activity normally required, thus inhibition is problematic. Nonetheless, inhibitors have been developed for in vitro use (e.g., gallates, tropolone, U-0521, and 3',4'-dihydroxy-2-methyl-propiophetropolone) and for clinical use 15 (e.g., nitrocatechol-based compounds and tolcapone). Administration of these inhibitors results in the increased half-life of L-dopa and the consequent formation of dopamine. Inhibition of COMT is also likely to increase the half-life of various other catechol-structure compounds, including but not limited to epinephrine/norepinephrine, isoprenaline, rimiterol, dobutamine, fenoldopam, apomorphine, and α-methyldopa. A deficiency in nonepinephrine has been linked to clinical depression, hence the use of 20 COMT inhibitors could be usefull in the treatment of depression. COMT inhibitors are generally well tolcrated with minimal side effects and are ultimately metabolized in the liver with only minor accumulation of metabolites in the body (Männistö, P.T. and S. Kaakkola (1999) Pharmacol. Rev. 51:593-628).

#### Copper-zinc superoxide dismutases

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Copper-zinc superoxide dismutases are compact homodimeric metalloenzymes involved in cellular defenses against oxidative damage. The enzymes contain one atom of zinc and one atom of copper per subunit and catalyze the dismutation of superoxide anions into O<sub>2</sub> and H<sub>2</sub>O<sub>2</sub>. The rate of dismutation is diffusion-limited and consequently enhanced by the presence of favorable electrostatic interactions between the substrate and enzyme active site. Examples of this class of enzyme have been identified in the cytoplasm of all the eukaryotic cells as well as in the periplasm of several bacterial species. Copper-zinc superoxide dismutases are robust enzymes that are highly resistant to proteolytic digestion and denaturing by urea and SDS. In addition to the compact structure of the enzymes, the presence of the metal ions and intrasubunit disulfide bonds is believed to be responsible

for enzyme stability. The enzymes undergo reversible denaturation at temperatures as high as 70 °C (Battistoni, A. et al. (1998) J. Biol. Chem. 273:5655-5661).

Overexpression of superoxide dismutase has been implicated in enhancing freezing tolerance of transgenic alfalfa as well as providing resistance to environmental toxins such as the diphenyl ether 5 herbicide, acifluorfen (McKersie, B.D. et al. (1993) Plant Physiol. 103:1155-1163). In addition, yeast cells become more resistant to freeze-thaw damage following exposure to hydrogen peroxide which causes the yeast cells to adapt to further peroxide stress by upregulating expression of superoxide dismutases. In this study, mutations to yeast superoxide dismutase genes had a more detrimental effect on freeze-thaw resistance than mutations which affected the regulation of glutathione 10 metabolism, long suspected of being important in determining an organism's survival through the process of cryopreservation (Jong-In Park, J.-I. et al. (1998) J. Biol. Chem. 273:22921-22928).

Expression of superoxide dismutase is also associated with Mycobacterium tuberculosis, the organism that causes tuberculosis. Superoxide dismutase is one of the ten major proteins secreted by M. tuberculosis and its expression is upregulated approximately 5-fold in response to oxidative stress.

15 M. tuberculosis expresses almost two orders of magnitude more superoxide dismutase than the nonpathogenic mycobacterium M. smegmatis, and secretes a much higher proportion of the expressed enzyme. The result is the secretion of ~350-fold more enzyme by M. tuberculosis than M. smegmatis, providing substantial resistance to oxidative stress (Harth, G. and M.A. Horwitz (1999) J. Biol. Chem. 274-4281-4292).

The reduced expression of copper-zinc superoxide dismutases, as well as other enzymes with anti-oxidant capabilities, has been implicated in the early stages of cancer. The expression of copper-zinc superoxide dismutases is reduced in prostatic intraepithelial neoplasia and prostate carcinomas, (Bostwick, D.G. (2000) Cancer 89:123-134).

### Phosphoesterases

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Phosphotriesterases (PTE, paraoxonases) are enzymes that hydrolyze toxic organophosphorus compounds and have been isolated from a variety of tissues. Phosphotriesterases play a central role in the detoxification of insecticides by mammals. Birds and insects lack PTE, and as a result have reduced tolerance for organophosphorus compounds (Vilanova, E. and M.A. Sogorb (1999) Crit. Rev. Toxicol. 29:21-57). Phosphotriesterase activity varies among individuals and is lower in infants than adults. PTE knockout mice are markedly more sensitive to the organophosphate-based toxins diazoxon and chlorpyrifos oxon (Furlong, C.E., et al. (2000) Neurotoxicology 21:91-100). Phosphotriesterase is also implicated in atherosclerosis and diseases involving lipoprotein metabolism.

Glycerophosphoryl diester phosphodiesterase (also known as glycerophosphodiester

phosphodiesterase) is a phosphodiesterase which hydrolyzes deacetylated phospholipid glycerophosphodiesters to produce sn-glycerol-3-phosphate and an alcohol. Glycerophosphocholine, glycerophosphoethanolamine, glycerophosphoglycerol, and glycerophosphoinositol are examples of substrates for glycerophosphoryl diester phosphodiesterases. A glycerophosphoryl diester 5 phosphodiesterase from E. coli has broad specificity for glycerophosphodiester substrates (Larson, T.J. et al. (1983) J. Biol. Chem. 248:5428-5432).

Cyclic nucleotide phosphodiesterases (PDEs) are crucial enzymes in the regulation of the cyclic nucleotides cAMP and cGMP. cAMP and cGMP function as intracellular second messengers to transduce a variety of extracellular signals including hormones, light, and neurotransmitters. PDEs 10 degrade cyclic nucleotides to their corresponding monophosphates, thereby regulating the intracellular concentrations of cyclic nucleotides and their effects on signal transduction. Due to their roles as regulators of signal transduction, PDEs have been extensively studied as chemotherapeutic targets (Perry, M.J. and G.A. Higgs (1998) Curr. Opin. Chem. Biol. 2:472-481; Torphy, J.T. (1998) Am. J. Resp. Crit. Care Med. 157:351-370).

Families of mammalian PDEs have been classified based on their substrate specificity and affinity, sensitivity to cofactors, and sensitivity to inhibitory agents (Beavo, J.A. (1995) Physiol. Rev. 75:725-748; Conti, M. et al. (1995) Endocrine Rev. 16:370-389). Several of these families contain distinct genes, many of which are expressed in different tissues as splice variants. Within PDE families, there are multiple isozymes and multiple splice variants of these isozymes (Conti, M. and S.-20 L.C. Jin (1999) Prog. Nucleic Acid Res. Mol. Biol. 63:1-38). The existence of multiple PDE families, isozymes, and splice variants is an indication of the variety and complexity of the regulatory pathways involving cyclic nucleotides (Houslay, M.D. and G. Milligan (1997) Trends Biochem. Sci. 22:217-224).

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Type 1 PDEs (PDE1s) are Ca2+/calmodulin-dependent and appear to be encoded by at least three different genes, each having at least two different splice variants (Kakkar, R. et al. (1999) Cell 25 Mol. Life Sci. 55:1164-1186). PDE1s have been found in the lung, heart, and brain. Some PDE1 isozymes are regulated in vitro by phosphorylation/dephosphorylation. Phosphorylation of these PDE1 isozymes decreases the affinity of the enzyme for calmodulin, decreases PDE activity, and increases steady state levels of cAMP (Kakkar et al., supra). PDE1s may provide useful therapeutic targets for disorders of the central nervous system and the cardiovascular and immune systems, due to the involvement of PDE1s in both cyclic nucleotide and calcium signaling (Perry and Higgs, supra).

PDE2s are cGMP-stimulated PDEs that have been found in the cerebellum, neocortex, heart, kidney, lung, pulmonary artery, and skeletal muscle (Sadhu, K. et al. (1999) J. Histochem. Cytochem. 47:895-906). PDE2s are thought to mediate the effects of cAMP on catecholamine secretion,

participate in the regulation of aldosterone (Beavo, *supra*), and play a role in olfactory signal transduction (Juilfs, D.M. et al. (1997) Proc. Natl. Acad. Sci. USA 94:3388-3395).

PDE3s have high affinity for both cGMP and cAMP, and so these cyclic nucleotides act as competitive substrates for PDE3s. PDE3s play roles in stimulating myocardial contractility, inhibiting 5 platelet aggregation, relaxing vascular and airway smooth muscle, inhibiting proliferation of T-lymphocytes and cultured vascular smooth muscle cells, and regulating catecholamine-induced release of free fatty acids from adipose tissue. The PDE3 family of phosphodiesterases are sensitive to specific inhibitors such as cilostamide, enoximone, and lixazinone. Isozymes of PDE3 can be regulated by cAMP-dependent protein kinase, or by insulin-dependent kinases (Degerman, E. et al. 10 (1997) J. Biol. Chem. 272:6823-6826).

PDE4s are specific for cAMP; are localized to airway smooth muscle, the vascular endothelium, and all inflammatory cells; and can be activated by cAMP-dependent phosphorylation. Since elevation of cAMP levels can lead to suppression of inflammatory cell activation and to relaxation of bronchial smooth muscle, PDE4s have been studied extensively as possible targets for 15 novel anti-inflammatory agents, with special emphasis placed on the discovery of asthma treatments. PDE4 inhibitors are currently undergoing clinical trials as treatments for asthma, chronic obstructive pulmonary disease, and atopic eczema. All four known isozymes of PDE4 are susceptible to the inhibitor rolipram, a compound which has been shown to improve behavioral memory in mice (Barad, M. et al. (1998) Proc. Natl. Acad. Sci. USA 95:15020-15025). PDE4 inhibitors have also been studied as possible therapeutic agents against acute lung injury, endotoxemia, rheumatoid arthritis, multiple sclerosis, and various neurological and gastrointestinal indications (Doherty, A.M. (1999) Curr. Opin. Chem. Biol. 3:466-473).

PDE5 is highly selective for cGMP as a substrate (Turko, I.V. et al. (1998) Biochemistry 37:4200-4205), and has two allosteric cGMP-specific binding sites (McAllister-Lucas, L.M. et al. (1995) J. Biol. Chem. 270:30671-30679). Binding of cGMP to these allosteric binding sites seems to be important for phosphorylation of PDE5 by cGMP-dependent protein kinase rather than for direct regulation of catalytic activity. High levels of PDE5 are found in vascular smooth muscle, platelets, lung, and kidney. The inhibitor zaprinast is effective against PDE5 and PDE1s. Modification of zaprinast to provide specificity against PDE5 has resulted in sildenafil (VIAGRA; Pfizer, Inc., New York NY), a treatment for male erectile dysfunction (Terrett, N. et al. (1996) Bioorg. Med. Chem. Lett. 6:1819-1824). Inhibitors of PDE5 are currently being studied as agents for cardiovascular therapy (Perry and Higgs, supra).

PDE6s, the photoreceptor cyclic nucleotide phosphodiesterases, are crucial components of the

phototransduction cascade. In association with the G-protein transducin, PDE6s hydrolyze cGMP to regulate cGMP-gated cation channels in photoreceptor membranes. In addition to the cGMP-binding active site, PDE6s also have two high-affinity cGMP-binding sites which are thought to play a regulatory role in PDE6 function (Artemyev, N.O. et al. (1998) Methods 14:93-104). Defects in 5 PDE6s have been associated with retinal disease. Retinal degeneration in the rd mouse (Yan, W. et al. (1998) Invest. Opthalmol. Vis. Sci. 39:2529-2536), autosomal recessive retinitis pigmentosa in humans (Danciger, M. et al. (1995) Genomics 30:1-7), and rod/cone dysplasia 1 in Irish Setter dogs (Suber, M.L. et al. (1993) Proc. Natl. Acad. Sci. USA 90:3968-3972) have been attributed to mutations in the PDE68 gene.

The PDE7 family of PDEs consists of only one known member having multiple splice variants (Bloom, T.J. and J.A. Beavo (1996) Proc. Natl. Acad. Sci. USA 93:14188-14192). PDE7s are cAMP specific, but little else is known about their physiological function. Although mRNAs encoding PDE7s are found in skeletal muscle, heart, brain, lung, kidney, and pancreas, expression of PDE7 proteins is restricted to specific tissue types (Han, P. et al. (1997) J. Biol. Chem. 272:16152-16157;

15 Perry and Higgs, supra). PDE7s are very closely related to the PDE4 family; however, PDE7s are not inhibited by rolivram, a specific inhibitor of PDE4s (Beavo, supra).

PDE8s are cAMP specific, and are closely related to the PDE4 family. PDE8s are expressed in thyroid gland, testis, eye, liver, skeletal muscle, heart, kidney, ovary, and brain. The cAMP-hydrolyzing activity of PDE8s is not inhibited by the PDE inhibitors rolipram, vinpocetine, milrinone, IBMX (3-isobutyl-1-methylxanthine), or zaprinast, but PDE8s are inhibited by dipyridamole (Fisher, D.A. et al. (1998) Biochem. Biophys. Res. Commun. 246:570-577; Hayashi, M. et al. (1998) Biochem. Biophys. Res. Commun. 250:751-756; Soderling, S.H. et al. (1998) Proc. Natl. Acad. Sci. USA 95:8991-8996).

PDE9s are cGMP specific and most closely resemble the PDE8 family of PDEs. PDE9s are

expressed in kidney, liver, lung, brain, spleen, and small intestine. PDE9s are not inhibited by sildenafil

(VIAGRA; Pfizer, Inc., New York NY), rolipram, vinpocetine, dipyridamole, or IBMX (3-isobutyl-1
methylxanthine), but they are sensitive to the PDE5 inhibitor zaprinast (Fisher, D.A. et al. (1998) J.

Biol. Chem. 273:15559-15564; Soderling, S.H. et al. (1998) J. Biol. Chem. 273:15553-15558).

PDE10s are dual-substrate PDEs, hydrolyzing both cAMP and cGMP. PDE10s are

or expressed in brain, thyroid, and testis. (Soderling, S.H. et al. (1999) Proc. Natl. Acad. Sci. USA

96:7071-7076; Fujishige, K. et al. (1999) J. Biol. Chem. 274:18438-18445; Loughney, K. et al (1999)

Gene 234:109-117).

PDEs are composed of a catalytic domain of about 270-300 amino acids, an N-terminal

regulatory domain responsible for binding cofactors, and, in some cases, a hydrophilic C-terminal domain of unknown function (Conti and Jin, supra). A conserved, putative zinc-binding motif has been identified in the catalytic domain of all PDEs. N-terminal regulatory domains include noncatalytic cGMP-binding domains in PDE2s, PDE5s, and PDE6s; calmodulin-binding domains in 5 PDE1s; and domains containing phosphorylation sites in PDE3s and PDE4s. In PDE5, the N-terminal cGMP-binding domain spans about 380 amino acid residues and comprises tandem repeats of a conserved sequence motif (McAllister-Lucas, L.M. et al. (1993) J. Biol. Chem. 268:22863-22873). The NKXnD motif has been shown by mutagenesis to be important for cGMP binding (Turko, I.V. et al. (1996) J. Biol. Chem. 271:22240-22244). PDE families display approximately 30% amino acid 10 identity within the catalytic domain; however, isozymes within the same family typically display about 85-95% identity in this region (e.g. PDE4A vs PDE4B). Furthermore, within a family there is extensive similarity (>60%) outside the catalytic domain; while across families, there is little or no sequence similarity outside this domain.

Many of the constituent functions of immune and inflammatory responses are inhibited by 15 agents that increase intracellular levels of cAMP (Verghese, M.W. et al. (1995) Mol. Pharmacol. 47:1164-1171). A variety of diseases have been attributed to increased PDE activity and associated with decreased levels of cyclic nucleotides. For example, a form of diabetes insipidus in mice has been associated with increased PDE4 activity, an increase in low-Km cAMP PDE activity has been reported in leukocytes of atopic patients, and PDE3 has been associated with cardiac disease.

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Many inhibitors of PDEs have undergone clinical evaluation (Perry and Higgs, supra; Torphy, T.J. (1998) Am. J. Respir, Crit. Care Med. 157:351-370). PDE3 inhibitors are being developed as antithrombotic agents, antihypertensive agents, and as cardiotonic agents useful in the treatment of congestive heart failure. Rolipram, a PDE4 inhibitor, has been used in the treatment of depression, and other PDE4 inhibitors have an anti-inflammatory effect. Rolipram may inhibit HIV-1 replication 25 (Angel, J.B. et al. (1995) AIDS 9:1137-1144). Additionally, rolipram suppresses the production of cytokines such as TNF-a and b and interferon g, and thus is effective against encephalomyelitis. Rolipram may also be effective in treating tardive dyskinesia and multiple sclerosis (Sommer, N. et al. (1995) Nat. Med. 1:244-248; Sasaki, H. et al. (1995) Eur. J. Pharmacol. 282:71-76). Theophylline is a nonspecific PDE inhibitor used in treatment of bronchial asthma and other respiratory diseases. 30 Theophylline is believed to act on airway smooth muscle function and in an anti-inflammatory or immunomodulatory capacity (Banner, K.H. and C.P. Page (1995) Eur. Respir. J. 8:996-1000). Pentoxifylline is another nonspecific PDE inhibitor used in the treatment of intermittent claudication and diabetes-induced peripheral vascular disease. Pentoxifylline is also known to block TNF-a

production and may inhibit HIV-1 replication (Angel et al., supra).

PDEs have been reported to affect cellular proliferation of a variety of cell types (Conti et al. (1995) Endocrine Rev. 16:370-389) and have been implicated in various cancers. Growth of prostate carcinoma cell lines DU145 and LNCaP was inhibited by delivery of cAMP derivatives and PDE 5 inhibitors (Bang, Y.J. et al. (1994) Proc. Natl. Acad. Sci. USA 91:5330-5334). These cells also showed a permanent conversion in phenotype from epithelial to neuronal morphology. It has also been suggested that PDE inhibitors can regulate mesangial cell proliferation (Matousovic, K. et al. (1995) J. Clin. Invest. 96:401-410) and lymphocyte proliferation (Joulain, C. et al. (1995) J. Lipid Mediat. Cell Signal, 11:63-79). One cancer treatment involves intracellular delivery of PDEs to particular cellular 10 compartments of tumors, resulting in cell death (Deonarain, M.P. and A.A. Epenetos (1994) Br. J. Cancer 70:786-794).

Members of the UDP glucuronyltransferase family (UGTs) catalyze the transfer of a glucuronic acid group from the cofactor uridine diphosphate-glucuronic acid (UDP-glucuronic acid) to a substrate. The transfer is generally to a nucleophilic heteroatom (O, N, or S). Substrates include 15 xenobiotics which have been functionalized by Phase I reactions, as well as endogenous compounds such as bilirubin, steroid hormones, and thyroid hormones. Products of glucuronidation are excreted in urine if the molecular weight of the substrate is less than about 250 g/mol, whereas larger glucuronidated substrates are excreted in bile.

UGTs are located in the microsomes of liver, kidney, intestine, skin, brain, spleen, and nasal 20 mucosa, where they are on the same side of the endoplasmic reticulum membrane as cytochrome P450 enzymes and flavin-containing monooxygenases. UGTs have a C-terminal membrane-spanning domain which anchors them in the endoplasmic reticulum membrane, and a conserved signature domain of about 50 amino acid residues in their C terminal section (PROSITE PDOC00359 UDPglycosyltransferase signature).

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UGTs involved in drug metabolism are encoded by two gene families, UGT1 and UGT2. Members of the UGT1 family result from alternative splicing of a single gene locus, which has a variable substrate binding domain and constant region involved in cofactor binding and membrane insertion. Members of the UGT2 family are encoded by separate gene loci, and are divided into two families, UGT2A and UGT2B. The 2A subfamily is expressed in olfactory epithelium, and the 2B 30 subfamily is expressed in liver microsomes. Mutations in UGT genes are associated with hyperbilirubinemia (OMIM #143500 Hyperbilirubinemia I); Crigler-Najjar syndrome, characterized by intense hyperbilirubinemia from birth (OMIM #218800 Crigler-Najjar syndrome); and a milder form of hyperbilirubinemia termed Gilbert's disease (OMIM #191740 UGT1).

#### Thioesterases

Two soluble thioesterases involved in fatty acid biosynthesis have been isolated from mammalian tissues, one which is active only toward long-chain fatty-acyl thioesters and one which is active toward thioesters with a wide range of fatty-acyl chain-lengths. These thioesterases catalyze the chain-terminating step in the *de novo* biosynthesis of fatty acids. Chain termination involves the hydrolysis of the thioester bond which links the fatty acyl chain to the 4-phosphopantetheine prosthetic group of the acyl carrier protein (ACP) subunit of the fatty acid synthase (Smith, S. (1981a) Methods Enzymol. 71:181-188; Smith, S. (1981b) Methods Enzymol. 71:188-200).

E. coli contains two soluble thioesterases, thioesterase I which is active only toward long-chain acyl thioesters, and thioesterase II (TEII) which has a broad chain-length specificity (Naggert, J. et al. (1991) J. Biol. Chem. 266:11044-11050). E. coli TEII does not exhibit sequence similarity with either of the two types of mammalian thioesterases which function as chain-terminating enzymes in de novo fatty acid biosynthesis. Unlike the mammalian thioesterases, E. coli TEII lacks the characteristic serine active site gly-X-ser-X-gly sequence motif and is not inactivated by the serine modifying agent diisopropyl fluorophosphate. However, modification of histidine 58 by iodoacetamide and diethylpyrocarbonate abolished TEII activity. Overexpression of TEII did not alter fatty acid content in E. coli, which suggests that it does not function as a chain-terminating enzyme in fatty acid biosynthesis (Naggert et al., supra). For that reason, Naggert et al. (supra) proposed that the physiological substrates for E. coli TEII may be coenzyme A (CoA)-fatty acid esters instead of ACP-phosphopanthetheine-fatty acid esters.

#### Carboxylesterases

Mammalian carboxylesterases are a multigene family expressed in a variety of tissues and cell types. Acetylcholinesterase, butyrylcholinesterase, and carboxylesterase are grouped into the serine superfamily of esterases (B-esterases). Other carboxylesterases include thyroglobulin, thrombin,

Factor IX, gliotactin, and plasminogen. Carboxylesterases catalyze the hydrolysis of ester- and amide-groups from molecules and are involved in detoxification of drugs, environmental toxins, and carcinogens. Substrates for carboxylesterases include short- and long-chain acyl-glycerols, acylcarnitine, carbonates, dipivefrin hydrochloride, cocaine, salicylates, capsaicin, palmitoyl-coenzyme A, imidapril, haloperidol, pyrrolizidine alkaloids, steroids, p-nitrophenyl acetate, malathion, butanilicaine, and isocarboxazide. Carboxylesterases are also important for the conversion of prodrugs to free acids, which may be the active form of the drug (e.g., lovastatin, used to lower blood cholesterol) (reviewed in Satoh, T. and Hosokawa, M. (1998) Annu. Rev. Pharmacol. Toxicol.38:257-288). Neuroligins are a class of molecules that (i) have N-terminal signal sequences. (ii) resemble cell-

surface receptors, (iii) contain carboxylesterase domains, (iv) are highly expressed in the brain, and (v) bind to neurexins in a calcium-dependent manner. Despite the homology to carboxylesterases, neuroligins lack the active site serine residue, implying a role in substrate binding rather than eatalysis (Ichtchenko, K. et al. (1996) J. Biol. Chem. 271:2676-2682).

#### 5 Squalene epoxidase

Squalene epoxidase (squalene monooxygenase, SE) is a microsomal membrane-bound, FADdependent oxidoreductase that catalyzes the first oxygenation step in the sterol biosynthetic pathway
of eukaryotic cells. Cholesterol is an essential structural component of cytoplasmic membranes
acquired via the LDL receptor-mediated pathway or the biosynthetic pathway. SE converts squalene
to 2.3(5)-oxidosqualene, which is then converted to lanosterol and then cholesterol.

High serum cholesterol levels result in the formation of atherosclerotic plaques in the arteries of higher organisms. This deposition of highly insoluble lipid material onto the walls of essential blood vessels results in decreased blood flow and potential necrosis. HMG-CoA reductase is responsible for the first committed step in cholesterol biosynthesis, conversion of 3-hydroxyl-3-methyl-glutaryl CoA (HMG-CoA) to mevalonate. HMG-CoA is the target of a number of pharmaceutical compounds designed to lower plasma cholesterol levels, but inhibition of MHG-CoA also results in the reduced synthesis of non-sterol intermediates required for other biochemical pathways. Since SE catalyzes a rate-limiting reaction that occurs later in the sterol synthesis pathway with cholesterol as the only end product, SE is a better ideal target for the design of anti-hyperlipidemic drugs (Nakamura, Y. et al. 20 (1996) 271:8053-8056).

#### Epoxide hydrolases

Epoxide hydrolases catalyze the addition of water to epoxide-containing compounds, thereby hydrolyzing epoxides to their corresponding 1,2-diols. They are related to bacterial haloalkane dehalogenases and show sequence similarity to other members of the α/β hydrolase fold family of enzymes. This family of enzymes is important for the detoxification of xenobiotic epoxide compounds which are often highly electrophilic and destructive when introduced. Examples of epoxide hydrolase reactions include the hydrolysis of some leukotoxin to leukotoxin diol, and isoleukotoxin to isoleukotoxin diol. Leukotoxins alter membrane permeability and ion transport and cause inflammatory responses. In addition, epoxide carcinogens are produced by cytochrome P450 as intermediates in the detoxification of drugs and environmental toxins. Epoxide hydrolases possess a catalytic triad composed of Asp, Asp, and His (Arand, M. et al. (1996) J. Biol. Chem. 271:4223-4229; Rink, R. et al. (1997) J. Biol. Chem. 272:14650-14657; Argiriadi, M.A. et al. (2000) J. Biol. Chem. 275:15265-15270).

### Enzymes involved in tyrosine catalysis

The degradation of the amino acid tyrosine, to either succinate and pyruvate or fumarate and acetoacetate, requires a large number of enzymes and generates a large number of intermediate compounds. In addition, many xenobiotic compounds may be metabolized using one or more reactions 5 that are part of the tyrosine catabolic pathway. Enzymes involved in the degradation of tyrosine to succinate and pyruvate (e.g., in Arthrobacter species) include 4-hydroxyphenylpyruvate oxidase, 4-hydroxyphenylacetate 3-hydroxylase, 3,4-dihydroxyphenylacetate 2,3-dioxygenase, 5-carboxymethyl-2-hydroxymuconic semialdehyde dehydrogenase, trans, cis-5-carboxymethyl-2-hydroxymuconate isomerase, homoprotocatechuate isomerase/decarboxylase, 10 cis-2-oxohept-3-ene-1,7-dioate hydratase, 2.4-dihydroxyhept-trans-2-ene-1,7-dioate aldolase, and succinic semialdehyde dehydrogenase. Enzymes involved in the degradation of tyrosine to fumarate and acetoacetate (e.g., in Pseudomonas species) include 4-hydroxyphenylpyruvate dioxygenase, homogentisate 1,2-dioxygenase, maleylacetoacetate isomerase, fumarylacetoacetase and 4-hydroxyphenylacetate. Additional enzymes associated with tyrosine metabolism in different 15 organisms include 4-chlorophenylacetate-3,4-dioxygenase, aromatic aminotransferase, 5-oxopent-3-ene-1,2,5-tricarboxylate decarboxylase, 2-oxo-hept-3-ene-1,7-dioate hydratase, and 5-carboxymethyl-2-hydroxymuconate isomerase (Ellis, L.B.M. et al. (1999) Nucleic Acids Res. 27:373-376; Wackett, L.P. and Ellis, L.B.M. (1996) J. Microbiol. Meth. 25:91-93; and Schmidt, M. (1996) Amer. Soc. Microbiol. News 62:102). 20

In humans, acquired or inherited genetic defects in enzymes of the tyrosine degradation pathway may result in hereditary tyrosinemia. One form of this disease, hereditary tyrosinemia 1 (HT1) is caused by a deficiency in the enzyme fumarylacetoacetate hydrolase, the last enzyme in the pathway in organisms that metabolize tyrosine to fumarate and acetoacetate. HT1 is characterized by progressive liver damage beginning at infancy, and increased risk for liver cancer (Endo, F. et al. 25 (1997) J. Biol. Chem. 272:24426-24432).

#### Expression profiling

Microarrays are analytical tools used in bioanalysis. A microarray has a plurality of molecules spatially distributed over, and stably associated with, the surface of a solid support. Microarrays of polypeptides, polynucleotides, and/or antibodies have been developed and find use in a variety of applications, such as gene sequencing, monitoring gene expression, gene mapping, bacterial identification, drug discovery, and combinatorial chemistry.

One area in particular in which microarrays find use is in gene expression analysis. Array technology can provide a simple way to explore the expression of a single polymorphic gene or the

expression profile of a large number of related or unrelated genes. When the expression of a single gene is examined, arrays are employed to detect the expression of a specific gene or its variants.

When an expression profile is examined, arrays provide a platform for identifying genes that are tissue specific, are affected by a substance being tested in a toxicology assay, are part of a signaling

cascade, carry out housekeeping functions, or are specifically related to a particular genetic predisposition, condition, disease, or disorder.

### Cancer

#### Breast cancer

Breast cancer is the most frequently diagnosed type of cancer in American women and the second most frequent cause of cancer death. The lifetime risk of an American woman developing breast cancer is 1 in 8, and one-third of women diagnosed with breast cancer die of the disease. A number of risk factors have been identified, including hormonal and genetic factors. One genetic defect associated with breast cancer results in a loss of heterozygosity (LOH) at multiple loci such as p53, Rb, BRCA1, and BRCA2. Another genetic defect is gene amplification involving genes such as c-myc and c-erbB2 (Her2-neu gene). Steroid and growth factor pathways are also altered in breast cancer, notably the estrogen, progesterone, and epidermal growth factor (EGF) pathways. Breast cancer evolves through a multi-step process whereby premalignant mammary epithelial cells undergo a relatively defined sequence of events leading to tumor formation. An early event in tumor development is ductal hyperplasia. Cells undergoing rapid neoplastic growth gradually progress to invasive carcinoma and become metastatic to the lung, bone, and potentially other organs. Variables that may influence the process of tumor progression and malignant transformation include genetic factors, environmental factors, growth factors, and hormones.

Tumor cells stimulate the formation of stroma that secretes various mediators, such as growth factors, cytokines, and proteases, which are critical for tumor growth. For instance, serum tumor necrosis factor alpha (TNF-α) is increased in the circulation of patients with malignancy. Interferongamma (IFN-γ), also known as Type II interferon or immune interferon, is produced primarily by T-lymphocytesand natural killer cells. IFN-γ was originally characterized based on its antiviral characteristics. The protein exhibits antiproliferative, immunoregulatory and proinflammatory activities and is thus important in host defense mechanisms. IFN-γ induces the production of cytokines, upregulates the expression of class I and II MHC antigens, Fe receptor, and leukocyte adhesion molecules. It modulates macrophage effector functions, influences isotype switching and potentiates the exerction of immunoglobulins by B cells. IFN-γ also augments TH1 cell expansion and may be required for TH1 cell differentiation. The IFN-γ receptor has been cloned and characterized, and is

structurally related to the recently cloned IL-10 receptor. It is present on almost all cell types except mature crythrocytes. T-47D is a breast carcinoma cell line isolated from a pleural effusion obtained from a donor with an infiltrating ductal carcinoma of the breast.

### Colon cancer

While soft tissue sarcomas are relatively rare, more than 50% of new patients diagnosed with the disease will die from it. The molecular pathways leading to the development of sarcomas are relatively unknown, due to the rarity of the disease and variation in pathology. Colon cancer evolves through a multi-step process whereby pre-malignant colonocytes undergo a relatively defined sequence of events leading to tumor formation. Several factors participate in the process of tumor progression and malignant transformation including genetic factors, mutations, and selection.

To understand the nature of gene alterations in colorectal cancer, a number of studies have focused on the inherited syndromes. Familial adenomatous polyposis (FAP), is caused by mutations in the adenomatous polyposis coli gene (APC), resulting in truncated or inactive forms of the protein. This tumor suppressor gene has been mapped to chromosome 5q. Hereditary nonpolyposis colorectal cancer (HNPCC) is caused by mutations in mis-match repair genes. Although hereditary colon cancer syndromes occur in a small percentage of the population and most colorectal cancers are considered sporadic, knowledge from studies of the hereditary syndromes can be generally applied. For instance, somatic mutations in APC occur in at least 80% of sporadic colon tumors. APC mutations are thought to be the initiating event in the disease. Other mutations occur subsequently.

20 Approximately 50% of colorectal cancers contain activating mutations in ras, while 85% contain inactivating mutations in p53. Changes in all of these genes lead to gene expression changes in colon cancer.

#### Lung cancer

Lung cancer is the leading cause of cancer death for men and the second leading cause of

cancer death for women in the U.S. Lung cancers are divided into four histopathologically distinct
groups. Three groups (squamous cell carcinoma, adenocarcinoma, and large cell carcinoma) are
classified as non-small cell lung cancers (NSCLCs). The fourth group of cancers is referred to as
small cell lung cancer (SCLC). Deletions on chromosome 3 are common in this disease and are
thought to indicate the presence of a tumor suppressor gene in this region. Activating mutations in K
or ras are commonly found in lung cancer and are the basis of one of the mouse models for the disease.

Osteosarcoma

Osteosarcoma is the most common malignant bone tumor in children. Approximately 80% of patients present with non-metastatic disease. After the diagnosis is made by an initial biopsy,

treatment involves the use of 3-4 courses of neoadjuvant chemotherapy before definitive surgery, followed by post-operative chemotherapy. With currently available treatment regimens, approximately 30-40% of patients with non-metastatic disease relapse after therapy. Currently, there is no prognostic factor that can be used at the time of initial diagnosis to predict which patients will have a high risk of relapse. The only significant prognostic factor predicting the outcome in a patient with non-metastatic osteosarcoma is the histopathologic response of the primary tumor resected at the time of definitive surgery. The degree of necrosis in the primary tumor is a reflection of the tumor response to neoadjuvant chemotherapy. A higher degree of necrosis (good or favorable response) is associated with a lower risk of relapse and a better outcome. Patients with a lower degree of 10 necrosis (poor or unfavorable response) have a much higher risk of relapse and poor outcome even after complete resection of the primary tumor. Unfortunately, poor outcome cannot be altered despite modification of post-operative chemotherapy to account for the resistance of the primary tumor to neoadjuvant chemotherapy. Thus, there is an urgent need to identify prognostic factors that can be used at the time of diagnosis to recognize the subtypes of osteosarcomas that have various risks of 15 relapse, so that more appropriate chemotherapy can be used at the outset to improve the outcome. Ovarian cancer

Ovarian cancer is the leading cause of death from a gynecologic cancer. The majority of ovarian cancers are derived from epithelial cells, and 70% of patients with epithelial ovarian cancers present with late-stage disease. As a result, the long-term survival rate for this disease is very low. Identification of early-stage markers for ovarian cancer would significantly increase the survival rate. Genetic variations involved in ovarian cancer development include mutation of p53 and microsatellite instability. Gene expression patterns likely vary when normal ovary is compared to ovarian tumors. Prostate cancer

As with most tumors, prostate cancer develops through a multistage progression ultimately
resulting in an aggressive tumor phenotype. The initial step in tumor progression involves the
hyperproliferation of normal luminal and/or basal epithelial cells. Androgen responsive cells become
hyperplastic and evolve into early-stage tumors. Although early-stage tumors are often androgen
sensitive and respond to androgen ablation, a population of androgen independent cells evolve from the
hyperplastic population. These cells represent a more advanced form of prostate tumor that may
become invasive and potentially become metastatic to the bone, brain, or lung. A variety of genes
may be differentially expressed during tumor progression. For example, loss of heterozygosity (LOH)
is frequently observed on chromosome 8p in prostate cancer. Fluorescence in situ hybridization
(FISII) revealed a deletion for at least 1 locus on 8p in 29 (69%) tumors, with a significantly higher

frequency of the deletion on 8p21.2-p21.1 in advanced prostate cancer than in localized prostate cancer, implying that deletions on 8p22-p21.3 play an important role in tumor differentiation, while 8p21.2-p21.1 deletion plays a role in progression of prostate cancer (Oba, K. et al. (2001) Cancer Genet. Cytogenet. 124: 20-26).

### 5 Tangier Disease

Tangier disease (TD) is a genetic disorder characterized by the near absence of circulating high density lipoprotein (HDL) and the accumulation of cholesterol esters in many tissues, including tonsils, lymph nodes, liver, spleen, thymus, and intestine. Low levels of HDL represent a clear predictor of premature coronary artery disease and homozygous TD correlates with a four- to six-fold increase in cardiovascular disease compared to controls. HDL plays a cardio-protective role in reverse cholesterol transport, the flux of cholesterol from peripheral cells such as tissue macrophages through plasma lipoproteins to the liver. The HDL protein, apolipoprotein A-I, plays a major role in this process, interacting with the cell surface to remove excess cholesterol and phospholipids. This pathway is severely impaired in TD and the defect lies in a specific gene, the ABC1 transporter. This gene is a member of the family of ATP-binding cassette transporters, which utilize ATP hydrolysis to transport a variety of substrates across membranes.

# Alzheimer's Disease

Alzheimer's disease (AD) is a progressive, neurodestructive process of the human neocortex, characterized by the deterioration of memory and higher cognitive function. A progressive and 20 irreversible brain disorder, AD is characterized by three major pathogenic episodes involving (a) an aberrant processing and deposition of beta-amyloid precursor protein (betaAPP) to form neurotoxic beta-amyloid (betaA) peptides and an aggregated insoluble polymer of betaA that forms the senile plaque, (b) the establishment of intraneuronal neuritic tau pathology yielding widespread deposits of agyrophilic neurofibrillary tangles (NFT) and (c) the initiation and proliferation of a brain-specific 25 inflammatory response. These three seemingly disperse attributes of AD etiopathogenesis are linked by the fact that proinflammatory microglia, reactive astrocytes and their associated cytokines and chemokines are associated with the biology of the microtubule associated protein tau, betaA speciation and aggregation. Missense mutations in the presentlin genes PS1 and PS2, implicated in early onset familial AD, cause abnormal betaAPP processing with resultant overproduction of betaA42 and 30 related neurotoxic peptides. Specific betaA fragments such as betaA42 can further potentiate proinflammatory mechanisms. Expression of the inducible oxidoreductase cyclooxygenase-2 and cytosolic phospholipase A2 (cPLA2) is strongly activated during cerebral ischemia and trauma, epilepsy and AD, indicating the induction of proinflammatory gene pathways as a response to brain

injury. Neurotoxic metals such as aluminum and zinc, both implicated in AD etiopathogenesis, and arachidonic acid, a major metabolite of brain cPLA2 activity, each polymerize hyperphosphorylated tau to form NFT-like bundles. Studies have identified a reduced risk for AD in patients aged over 70 years who were previously treated with non-steroidal anti-inflammatory drugs for non-CNS afflictions 5 that include arthritis. (For a review of the interrelationships between the mechanisms of PS1, PS2 and betaAPP gene expression, tau and betaA deposition and the induction, regulation and proliferation in AD of the neuroinflammatory response, see Lukiw, W.J, and Bazan, N.G. (2000) Neurochem. Res. 2000 25:1173-1184).

#### C3A Cells

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The human C3A cell line is a clonal derivative of HepG2/C3 (hepatoma cell line, isolated from a 15-year-old male with liver tumor), which was selected for strong contact inhibition of growth. The use of a clonal population enhances the reproducibility of the cells. C3A cells have many characteristics of primary human hepatocytes in culture: i) expression of insulin receptor and insulinlike growth factor II receptor; ii) secretion of a high ratio of serum albumin compared with α-15 fetoprotein; iii) conversion of ammonia to urea and glutamine; iv) metabolism of aromatic amino acids; and v) proliferation in glucose-free and insulin-free medium. The C3A cell line is now well established as an in vitro model of the mature human liver (Mickelson et al. (1995) Hepatology 22:866-875; Nagendra et al. (1997) Am. J. Physiol. 272:G408-G416).

Gemfibrozil is a fibric acid antilipemic agent that lowers serum triglycerides and produces 20 favorable changes in lipoproteins. Gemfibrozil is effective in reducing the risk of coronary heart disease in men (Frick, M.H., et al. (1987) New Engl. J. Med; 317:1237-1245). The compound can inhibit peripheral lipolysis and decrease hepatic extraction of free fatty acids, which decreases hepatic triglyceride production. Gemfibrozil also inhibits the synthesis and increases the clearance of apolipoprotein B, a carrier molecule for VLDL. Gemfibrozil has variable effects on LDL cholesterol. 25 Although it causes moderate reductions in patients with type IIa hyperlipoproteinemia, changes in patients with either type IIb or type IV hyperlipoproteinemia are unpredictable. In general, the HMG-CoA reductase inhibitors are more effective than gemfibrozil in reducing LDL cholesterol. At the molecular level gemfibozil may function as a peroxisome proliferator-activated receptor (PPAR) agonist. Gemfibrozil is rapidly and completely absorbed from the GI tract and undergoes 30 enterohepatic recirculation. Gemfibrozil is metabolized by the liver and excreted by the kidneys, mainly as metabolites, one of which possesses pharmacologic activity. Gemfibozil causes peroxisome proliferation and hepatocarcinogenesis in rats, which is a cause for concern generally for fibric acid derivative drugs. In humans, fibric acid derivatives are known to increase the risk of gall bladder

disease although gemfibrozil is better tolerated than other fibrates. The relative safety of gemfibrozil in humans compared to rodent species including rats may be attributed to differences in metabolism and clearance of the compound in different species (Dix, K.J., et al. (1999) Drug Metab. Distrib. 27:138-146; Thomas, B.F., et al. (1999) Drug Metab. Distrib. 27:147-157).

### 5 Dendritic cells

Dendritic cells (DCs) are antigen presenting cells that play a crucial role in the initiation of the immune response. DCs can be derived in vitro either from CD34+ bone marrow precursors (IDCs) or from peripheral blood monocytic cells (mDCs). In vivo, DCs reside in two distinct compartments: the peripheral tissues such as lung, skin, kidney, heart, and intestine; and in secondary lymphoid organs such as lymph node, spleen, and Peyer's patches. In the periphery, DCs are efficient antigen processing cells but are limited in their capacity to activate naive T cells. Upon activation (injury, inflammation, infection), DCs enter their final stage of maturation during which they downregulate the capacity to process new antigens, migrate out of the periphery into the secondary lymphoid organs, and acquire an extremely potent capacity to activate naive T cells. Several factors, such as cross-linking the CD40 surface molecules or the presence of TNF-α, can induce this final stage of maturation. CD40 is a type I integral membrane glycoprotein belonging to the TNF-receptor family. It is expressed on all mature B lymphocytes, dendritic cells, and some epithelial cells. Antibodies specific for CD40 molecules can induce proliferation of B cells when presented with interleukin-4 (IL-4) or antibodies specific for CD20 molecules. Also, stimulation of B cells with anti-CD40 antibodies

#### Steroid Hormones

Steroids are a class of lipid-soluble molecules, including cholesterol, bile acids, vitamin D, and hormones, that share a common four-ring structure based on cyclopentanoperhydrophenanthrene and that carrry out a wide variety of functions. Cholesterol, for example, is a component of cell

25 membranes that controls membrane fluidity. It is also a precursor for bile acids which solubilize lipids and facilitate absorption in the small intestine during digestion. Vitamin D regulates the absorption of calcium in the small intestine and controls the concentration of calcium in plasma. Steroid hormones, produced by the adrenal cortex, ovaries, and testes, include glucocorticoids, mineralocorticoids, androgens, and estrogens. They control various biological processes by binding to intracellular receptors that regulate transcription of specific genes in the nucleus. Glucocorticoids, for example, increase blood glucose concentrations by regulation of gluconeogenesis in the liver, increase blood concentrations of fatty acids by promoting lipolysis in adipose tissues, modulate sensitivity to catcholamines in the central nervous system, and reduce inflammation. The principal

mineralocorticoid, aldosterone, is produced by the adrenal cortex and acts on cells of the distal tubules of the kidney to enhance sodium ion reabsorption. Androgens, produced by the interstitial cells of Leydig in the testis, include the male sex hormone testosterone, which triggers changes at puberty, the production of sperm and maintenance of secondary sexual characteristics. Female sex hormones, estrogen and progesterone, are produced by the ovaries and also by the placenta and adrenal cortex of the fetus during pregnancy. Estrogen regulates female reproductive processes and secondary sexual characteristics. Progesterone regulates changes in the endometrium during the menstrual cycle and pregnancy.

Steroid hormones are widely used for fertility control and in anti-inflammatory treatments for physical injuries and diseases such as arthritis, asthma, and autoimmune disorders. Progesterone, a naturally occurring progestin, is primarily used to treat amenorrhea, abnormal uterine bleeding, or as a contraceptive. Endogenous progesterone is responsible for inducing secretory activity in the endometrium of the estrogen-primed uterus in preparation for the implantation of a fertilized egg and for the maintenance of pregnancy. It is secreted from the corpus luteum in response to luteinizing 15 hormone (LH). The primary contraceptive effect of exogenous progestins involves the suppression of the midcycle surge of LH. At the cellular level, progestins diffuse freely into target cells and bind to the progesterone receptor. Target cells include the female reproductive tract, the mammary gland, the hypothalamus, and the pituitary. Once bound to the receptor, progestins slow the frequency of release of gonadotropin releasing hormone from the hypothalamus and blunt the pre-ovulatory LH surge, thereby preventing follicular maturation and ovulation. Progesterone has minimal estrogenic and androgenic activity. Progesterone is metabolized hepatically to pregnanediol and conjugated with elucuronic acid.

Medroxyprogesterone (MAH), also known as 60-methyl-17-hydroxyprogesterone, is a
synthetic progestin with a pharmacological activity about 15 times greater than progesterone. MAH is
used for the treatment of renal and endometrial carcinomas, amenorrhea, abnormal uterine bleeding,
and endometriosis associated with hormonal imbalance. MAH has a stimulatory effect on respiratory
centers and has been used in cases of low blood oxygenation caused by sleep apnea, chronic
obstructive pulmonary disease, or hypercapnia.

Mifepristone, also known as RU-486, is an antiprogesterone drug that blocks receptors of
progesterone. It counteracts the effects of progesterone, which is needed to sustain pregnancy.
Mifepristone induces spontaneous abortion when administered in early pregnancy followed by
treatment with the prostaglandin misoprostol. Further studies show that mifepristone at a substantially
lower dose can be highly effective as a postcoital contraceptive when administered within five days

after unprotected intercourse, thus providing women with a "morning-after pill" in case of contraceptive failure or sexual assault. Mifepristone also has potential uses in the treatment of breast and ovarian cancers in cases in which tumors are progesterone-dependent. It interferes with steroiddependent growth of brain meningiomas, and may be useful in treatment of endometriosis where it 5 blocks the estrogen-dependent growth of endometrial tissues. It may also be useful in treatment of uterine fibroid tumors and Cushing's Syndrome. Mifepristone binds to glucocorticoid receptors and interferes with cortisol binding. Mifepristone also may act as an anti-glucocorticoid and be effective for treating conditions where cortisol levels are elevated such as AIDS, anorexia nervosa, ulcers, diabetes. Parkinson's disease, multiple sclerosis, and Alzheimer's disease.

Danazol is a synthetic steroid derived from ethinyl testosterone. Danazol indirectly reduces estrogen production by lowering pituitary synthesis of follicle-stimulating hormone and LH. Danazol also binds to sex hormone receptors in target tissues, thereby exhibiting anabolic, antiestrognic, and weakly androgenic activity. Danazol does not possess any progestogenic activity, and does not suppress normal pituitary release of corticotropin or release of cortisol by the adrenal glands. Danazol 15 is used in the treatment of endometriosis to relieve pain and inhibit endometrial cell growth. It is also used to treat fibrocystic breast disease and hereditary angioedema.

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Corticosteroids are used to relieve inflammation and to suppress the immune response. They inhibit eosinophil, basophil, and airway epithelial cell function by regulation of cytokines that mediate the inflammatory response. They inhibit leukocyte infiltration at the site of inflammation, interfere in the function of mediators of the inflammatory response, and suppress the humoral immune response. Corticosteroids are used to treat allergies, asthma, arthritis, and skin conditions. Beclomethasone is a synthetic glucocorticoid that is used to treat steroid-dependent asthma, to relieve symptoms associated with allergic or nonallergic (vasomotor) rhinitis, or to prevent recurrent nasal polyps following surgical removal. The anti-inflammatory and vasoconstrictive effects of intranasal beclomethasone are 5000 times greater than those produced by hydrocortisone. Budesonide is a corticosteroid used to control symptoms associated with allergic rhinitis or asthma. Budesonide has high topical anti-inflammatory activity but low systemic activity. Dexamethasone is a synthetic glucocorticoid used in antiinflammatory or immunosuppressive compositions. It is also used in inhalants to prevent symptoms of asthma. Due to its greater ability to reach the central nervous system, dexamethasone is usually the treatment of choice to control cerebral edema. Dexamethasone is approximately 20-30 times more potent than hydrocortisone and 5-7 times more potent than prednisone. Prednisone is metabolized in the liver to its active form, prednisolone, a glucocorticoid with anti-inflammatory properties. Prednisone is approximately 4 times more potent than hydrocortisone and the duration of action of

prednisone is intermediate between hydrocortisone and dexamethasone. Prednisone is used to treat allograft rejection, asthma, systemic lupus erythematosus, arthritis, ulcerative colitis, and other inflammatory conditions. Betamethasone is a synthetic glucocorticoid with antiinflammatory and immunosuppressive activity and is used to treat psoriasis and fungal infections, such as athlete's foot and ringworm.

The anti-inflammatory actions of corticosteroids are thought to involve phospholipase A<sub>2</sub>
inhibitory proteins, collectively called lipocortins. Lipocortins, in turn, control the biosynthesis of potent
mediators of inflammation such as prostaglandins and leukotrienes by inhibiting the release of the
precursor molecule arachidonic acid. Proposed mechanisms of action include decreased IgE
synthesis, increased number of β-adrenergic receptors on leukocytes, and decreased arachidonic acid
metabolism. During an immediate allergic reaction, such as in chronic bronchial asthma, allergens
bridge the IgE antibodies on the surface of mast cells, which triggers these cells to release
chemotactic substances. Mast cell influx and activation, therefore, is partially responsible for the
inflammation and hyperirritability of the oral mucosa in asthmatic patients. This inflammation can be
retarded by administration of corticosteroids.

There is a need in the art for new compositions, including nucleic acids and proteins, for the diagnosis, prevention, and treatment of autoimmune/inflammatory disorders, infectious disorders, immune deficiencies, disorders of metabolism, reproductive disorders, neurological disorders, cardiovascular disorders, eye disorders, and cell proliferative disorders, including cancer.

# SUMMARY OF THE INVENTION

Various embodiments of the invention provide purified polypeptides, enzymes, referred to collectively as 'ENZM' and individually as 'ENZM-1,' 'ENZM-2,' 'ENZM-3,' 'ENZM-4,' 'ENZM-25 5,' 'ENZM-6,' 'ENZM-7,' 'ENZM-8,' 'ENZM-9,' 'ENZM-10,' 'ENZM-11,' 'ENZM-12,' 'ENZM-13,' 'ENZM-14,' 'ENZM-16,' 'ENZM-16,' 'ENZM-17,' 'ENZM-18,' 'ENZM-19,' 'ENZM-20,' 'ENZM-21,' 'ENZM-22,' 'ENZM-22,' 'ENZM-23,' 'ENZM-26,' 'ENZM-27,' 'ENZM-28,' 'ENZM-30,' 'ENZM-31,' 'ENZM-32,' 'ENZM-33,' 'ENZM-34,' 'ENZM-35,' 'ENZM-36,' 'ENZM-37,' and 'ENZM-38,' and methods for using these proteins and their encoding polynucleotides for the detection, diagnosis, and treatment of diseases and medical conditions. Embodiments also provide methods for utilizing the purified enzymes and/or their encoding polynucleotides for facilitating the drug discovery process, including determination of efficacy, dosage, toxicity, and pharmacology. Related embodiments provide methods for utilizing the purified enzymes

and/or their encoding polynucleotides for investigating the pathogenesis of diseases and medical conditions.

An embodiment provides an isolated polypeptide selected from the group consisting of a) a
polypeptide comprising an amino acid sequence selected from the group consisting of SEQ ID NO:138, b) a polypeptide comprising a naturally occurring amino acid sequence at least 90% identical or at
least about 90% identical to an amino acid sequence selected from the group consisting of SEQ ID
NO:1-38, c) a biologically active fragment of a polypeptide having an amino acid sequence selected
from the group consisting of SEQ ID NO:1-38, and d) an immunogenic fragment of a polypeptide
having an amino acid sequence selected from the group consisting of SEQ ID NO:1-38. Another
embodiment provides an isolated polypeptide comprising an amino acid sequence of SEQ ID NO:1-38.

Still another embodiment provides an isolated polynucleotide encoding a polypeptide selected from the group consisting of a) a polypeptide comprising an amino acid sequence selected from the group consisting of SEQ ID NO:1-38, b) a polypeptide comprising a naturally occurring amino acid sequence at least 90% identical or at least about 90% identical to an amino acid sequence selected from the group consisting of SEQ ID NO:1-38, c) a biologically active fragment of a polypeptide having an amino acid sequence selected from the group consisting of SEQ ID NO:1-38, and d) an immunogenic fragment of a polypeptide having an amino acid sequence selected from the group consisting of SEQ ID NO:1-38. In another embodiment, the polynucleotide encodes a polypeptide selected from the group consisting of SEQ ID NO:1-38. In an alternative embodiment, the polynucleotide is selected from the group consisting of SEQ ID NO:39-76.

Still another embodiment provides a recombinant polynucleotide comprising a promoter sequence operably linked to a polynucleotide encoding a polypeptide selected from the group consisting of a) a polypeptide comprising an amino acid sequence selected from the group consisting of SEQ ID NO:1-38, b) a polypeptide comprising a naturally occurring amino acid sequence at least 90% identical or at least about 90% identical to an amino acid sequence selected from the group consisting of SEQ ID NO:1-38, c) a biologically active fragment of a polypeptide having an amino acid sequence selected from the group consisting of SEQ ID NO:1-38, and d) an immunogenic fragment of a polypeptide having an amino acid sequence selected from the group consisting of SEQ ID NO:1-38. Another embodiment provides a cell transformed with the recombinant polynucleotide. Yet another embodiment provides a transgenic organism comprising the recombinant polynucleotide.

Another embodiment provides a method for producing a polypeptide selected from the group consisting of a) a polypeptide comprising an amino acid sequence selected from the group consisting of SEQ ID NO:1-38, b) a polypeptide comprising a naturally occurring amino acid sequence at least

90% identical or at least about 90% identical to an amino acid sequence selected from the group consisting of SEQ ID NO:1-38, c) a biologically active fragment of a polypeptide having an amino acid sequence selected from the group consisting of SEQ ID NO:1-38, and d) an immunogenic fragment of a polypeptide having an amino acid sequence selected from the group consisting of SEQ ID NO:1-38.

The method comprises a) culturing a cell under conditions suitable for expression of the polypeptide, wherein said cell is transformed with a recombinant polynucleotide comprising a promoter sequence operably linked to a polynucleotide encoding the polypeptide, and b) recovering the polypeptide so expressed.

Yet another embodiment provides an isolated antibody which specifically binds to a

10 polypeptide selected from the group consisting of a) a polypeptide comprising an amino acid sequence
selected from the group consisting of SEQ ID NO:1-38, b) a polypeptide comprising a naturally
occurring amino acid sequence at least 90% identical or at least about 90% identical to an amino acid
sequence selected from the group consisting of SEQ ID NO:1-38, c) a biologically active fragment of
a polypeptide having an amino acid sequence selected from the group consisting of SEQ ID NO:1-38,

15 and d) an immunogenic fragment of a polypeptide having an amino acid sequence selected from the
group consisting of SEQ ID NO:1-38.

Still yet another embodiment provides an isolated polynucleotide selected from the group consisting of a) a polynucleotide comprising a polynucleotide sequence selected from the group consisting of SEQ ID NO:39-76, b) a polynucleotide comprising a naturally occurring polynucleotide sequence at least 90% identical or at least about 90% identical to a polynucleotide sequence selected from the group consisting of SEQ ID NO:39-76, c) a polynucleotide complementary to the polynucleotide of a), d) a polynucleotide complementary to the polynucleotide of b), and e) an RNA equivalent of a)-d). In other embodiments, the polynucleotide can comprise at least about 20, 30, 40, 60, 80, or 100 continuous nucleotides.

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Yet another embodiment provides a method for detecting a target polynucleotide in a sample, said target polynucleotide being selected from the group consisting of a) a polynucleotide comprising a polynucleotide sequence selected from the group consisting of SEQ ID NO:39-76, b) a polynucleotide comprising a naturally occurring polynucleotide sequence at least 90% identical or at least about 90% identical to a polynucleotide sequence selected from the group consisting of SEQ ID NO:39-76, c) a polynucleotide complementary to the polynucleotide of a), d) a polynucleotide complementary to the polynucleotide of b), and e) an RNA equivalent of a)-d). The method comprises a) hybridizing the sample with a probe comprising at least 20 contiguous nucleotides comprising a sequence complementary to said target polynucleotide in the sample, and which probe specifically hybridizes to

said target polynucleotide, under conditions whereby a hybridization complex is formed between said probe and said target polynucleotide or fragments thereof, and b) detecting the presence or absence of said hybridization complex. In a related embodiment, the method can include detecting the amount of the hybridization complex. In still other embodiments, the probe can comprise at least about 20, 30, 40, 60, 80, or 100 contiguous nucleotides.

Still yet another embodiment provides a method for detecting a target polynucleotide in a sample, said target polynucleotide being selected from the group consisting of a) a polynucleotide comprising a polynucleotide sequence selected from the group consisting of SEQ ID NO:39-76, b) a polynucleotide comprising a naturally occurring polynucleotide sequence at least 90% identical or at least about 90% identical to a polynucleotide sequence selected from the group consisting of SEQ ID NO:39-76, c) a polynucleotide complementary to the polynucleotide of a), d) a polynucleotide complementary to the polynucleotide of a), d) a polynucleotide complementary to the polynucleotide of a). The method comprises a) amplifying said target polynucleotide or fragment thereof using polymerase chain reaction amplification, and b) detecting the presence or absence of said amplified target polynucleotide or fragment thereof. In a related embodiment, the method can include detecting the amount of the amplified target polynucleotide or fragment thereof.

Another embodiment provides a composition comprising an effective amount of a polypeptide selected from the group consisting of a) a polypeptide comprising an amino acid sequence selected from the group consisting of SEQ ID NO:1-38, b) a polypeptide comprising a naturally occurring amino acid sequence at least 90% identical or at least about 90% identical to an amino acid sequence selected from the group consisting of SEQ ID NO:1-38, c) a biologically active fragment of a polypeptide having an amino acid sequence selected from the group consisting of SEQ ID NO:1-38, and d) an immunogenic fragment of a polypeptide having an amino acid sequence selected from the group consisting of SEQ ID NO:1-38, and a pharmaceutically acceptable excipient. In one embodiment, the composition can comprise an amino acid sequence selected from the group consisting of SEQ ID NO:1-38. Other embodiments provide a method of treating a disease or condition associated with decreased or abnormal expression of functional ENZM, comprising administering to a patient in need of such treatment the composition.

Yet another embodiment provides a method for screening a compound for effectiveness as an

30 agonist of a polypeptide selected from the group consisting of a) a polypeptide comprising an amino

acid sequence selected from the group consisting of SEQ ID NO:1-38, b) a polypeptide comprising a

naturally occurring amino acid sequence at least 90% identical or at least about 90% identical to an

amino acid sequence selected from the group consisting of SEQ ID NO:1-38, c) a biologically active

fragment of a polypeptide having an amino acid sequence selected from the group consisting of SEQ ID NO:1-38, and d) an immunogenic fragment of a polypeptide having an amino acid sequence selected from the group consisting of SEQ ID NO:1-38. The method comprises a) contacting a sample comprising the polypeptide with a compound, and b) detecting agonist activity in the sample.

Another embodiment provides a composition comprising an agonist compound identified by the method and a pharmaceutically acceptable excipient. Yet another embodiment provides a method of treating a disease or condition associated with decreased expression of functional ENZM, comprising administering to a patient in need of such treatment the composition.

Still yet another embodiment provides a method for screening a compound for effectiveness

as an antagonist of a polypeptide selected from the group consisting of a) a polypeptide comprising an
amino acid sequence selected from the group consisting of SEQ ID NO:1-38, b) a polypeptide
comprising a naturally occurring amino acid sequence at least 90% identical or at least about 90%
identical to an amino acid sequence selected from the group consisting of SEQ ID NO:1-38, c) a
biologically active fragment of a polypeptide having an amino acid sequence selected from the group

consisting of SEQ ID NO:1-38, and d) an immunogenic fragment of a polypeptide having an amino
acid sequence selected from the group consisting of SEQ ID NO:1-38. The method comprises a)
contacting a sample comprising the polypeptide with a compound, and b) detecting antagonist activity
in the sample. Another embodiment provides a composition comprising an antagonist compound
identified by the method and a pharmaceutically acceptable excipient. Yet another embodiment
provides a method of treating a disease or condition associated with overexpression of functional
ENZM, comprising administering to a patient in need of such treatment the composition.

Another embodiment provides a method of screening for a compound that specifically binds to a polypeptide selected from the group consisting of a) a polypeptide comprising an amino acid sequence selected from the group consisting of SEQ ID NO:1-38, b) a polypeptide comprising a 25 naturally occurring amino acid sequence at least 90% identical or at least about 90% identical to an amino acid sequence selected from the group consisting of SEQ ID NO:1-38, c) a biologically active fragment of a polypeptide having an amino acid sequence selected from the group consisting of SEQ ID NO:1-38, and d) an immunogenic fragment of a polypeptide having an amino acid sequence selected from the group consisting of SEQ ID NO:1-38. The method comprises a) combining the polypeptide with at least one test compound under suitable conditions, and b) detecting binding of the polypeptide to the test compound, thereby identifying a compound that specifically binds to the polypeptide.

Yet another embodiment provides a method of screening for a compound that modulates the

activity of a polypeptide selected from the group consisting of a) a polypeptide comprising an amino acid sequence selected from the group consisting of SEQ ID NO:1-38, b) a polypeptide comprising a naturally occurring amino acid sequence at least 90% identical or at least about 90% identical to an amino acid sequence selected from the group consisting of SEQ ID NO:1-38, c) a biologically active fragment of a polypeptide having an amino acid sequence selected from the group consisting of SEQ ID NO:1-38, and d) an immunogenic fragment of a polypeptide having an amino acid sequence selected from the group consisting of SEQ ID NO:1-38. The method comprises a) combining the polypeptide with at least one test compound under conditions permissive for the activity of the polypeptide in the polypeptide in the presence of the test compound, and c) comparing the activity of the polypeptide in the presence of the test compound with the activity of the polypeptide in the presence of the test compound is indicative of a compound that modulates the activity of the polypeptide.

Still yet another embodiment provides a method for screening a compound for effectiveness in
altering expression of a target polynucleotide, wherein said target polynucleotide comprises a
polynucleotide sequence selected from the group consisting of SEQ ID NO:39-76, the method
comprising a) contacting a sample comprising the target polynucleotide with a compound, b) detecting
altered expression of the target polynucleotide, and c) comparing the expression of the target
polynucleotide in the presence of varying amounts of the compound and in the absence of the
compound.

Another embodiment provides a method for assessing toxicity of a test compound, said method comprising a) treating a biological sample containing nucleic acids with the test compound; b) hybridizing the nucleic acids of the treated biological sample with a probe comprising at least 20 contiguous nucleotides of a polynucleotide selected from the group consisting of i) a polynucleotide comprising a polynucleotide sequence selected from the group consisting of SEQ ID NO:39-76, ii) a polynucleotide comprising a naturally occurring polynucleotide sequence at least 90% identical or at least about 90% identical to a polynucleotide sequence selected from the group consisting of SEQ ID NO:39-76, iii) a polynucleotide having a sequence complementary to i), iv) a polynucleotide complementary to the polynucleotide of ii), and v) an RNA equivalent of i)-iv). Hybridization occurs under conditions whereby a specific hybridization complex is formed between said probe and a target polynucleotide in the biological sample, said target polynucleotide selected from the group consisting of i) a polynucleotide comprising a polynucleotide sequence selected from the group consisting of SEQ ID NO:39-76, ii) a polynucleotide comprising a naturally occurring polynucleotide sequence at least

90% identical or at least about 90% identical to a polynucleotide sequence selected from the group consisting of SEQ ID NO:39-76, iii) a polynucleotide complementary to the polynucleotide of i), iv) a polynucleotide complementary to the polynucleotide of ii), and v) an RNA equivalent of i)-iv).

Alternatively, the target polynucleotide can comprise a fragment of a polynucleotide selected from the group consisting of i)-v) above; c) quantifying the amount of hybridization complex; and d) comparing the amount of hybridization complex in an untreated biological sample, wherein a difference in the amount of hybridization complex in the treated biological sample is indicative of toxicity of the test compound.

### BRIEF DESCRIPTION OF THE TABLES

Table 1 summarizes the nomenclature for full length polynucleotide and polypeptide embodiments of the invention.

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Table 2 shows the GenBank identification number and annotation of the nearest GenBank homolog, and the PROTEOME database identification numbers and annotations of PROTEOME 15 database homologs, for polypeptide embodiments of the invention. The probability scores for the matches between each polypeptide and its homolog(s) are also shown.

Table 3 shows structural features of polypeptide embodiments, including predicted motifs and domains, along with the methods, algorithms, and searchable databases used for analysis of the polypeptides.

Table 4 lists the cDNA and/or genomic DNA fragments which were used to assemble polynucleotide embodiments, along with selected fragments of the polynucleotides.

Table 5 shows representative cDNA libraries for polynucleotide embodiments.

Table 6 provides an appendix which describes the tissues and vectors used for construction of the cDNA libraries shown in Table 5.

Table 7 shows the tools, programs, and algorithms used to analyze polynucleotides and polypeptides, along with applicable descriptions, references, and threshold parameters.

Table 8 shows single nucleotide polymorphisms found in polynucleotide sequences of the invention, along with allele frequencies in different human populations.

# DESCRIPTION OF THE INVENTION

Before the present proteins, nucleic acids, and methods are described, it is understood that embodiments of the invention are not limited to the particular machines, instruments, materials, and methods described, as these may vary. It is also to be understood that the terminology used herein is

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for the purpose of describing particular embodiments only, and is not intended to limit the scope of the invention

As used herein and in the appended claims, the singular forms "a," "an," and "the" include
plural reference unless the context clearly dictates otherwise. Thus, for example, a reference to "a

5 host cell" includes a plurality of such host cells, and a reference to "an antibody" is a reference to one
or more antibodies and equivalents thereof known to those skilled in the art, and so forth.

Unless defined otherwise, all technical and scientific terms used herein have the same meanings as commonly understood by one of ordinary skill in the art to which this invention belongs. Although any machines, materials, and methods similar or equivalent to those described herein can be used to practice or test the present invention, the preferred machines, materials and methods are now described. All publications mentioned herein are cited for the purpose of describing and disclosing the cell lines, protocols, reagents and vectors which are reported in the publications and which might be used in connection with various embodiments of the invention. Nothing herein is to be construed as an admission that the invention is not entitled to antedate such disclosure by virtue of prior invention.

## 15 DEFINITIONS

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"ENZM" refers to the amino acid sequences of substantially purified ENZM obtained from any species, particularly a mammalian species, including bovine, ovine, porcine, murine, equine, and human, and from any source, whether natural, synthetic, semi-synthetic, or recombinant.

The term "agonist" refers to a molecule which intensifies or mimics the biological activity of

ENZM. Agonists may include proteins, nucleic acids, carbohydrates, small molecules, or any other

compound or composition which modulates the activity of ENZM either by directly interacting with

ENZM or by acting on components of the biological pathway in which ENZM participates.

An "allelic variant" is an alternative form of the gene encoding ENZM. Allelic variants may result from at least one mutation in the nucleic acid sequence and may result in altered mRNAs or in

25 polypeptides whose structure or function may or may not be altered. A gene may have none, one, or many allelic variants of its naturally occurring form. Common mutational changes which give rise to allelic variants are generally ascribed to natural deletions, additions, or substitutions of nucleotides.

Each of these types of changes may occur alone, or in combination with the others, one or more times in a given sequence.

"Altered" nucleic acid sequences encoding ENZM include those sequences with deletions, insertions, or substitutions of different nucleotides, resulting in a polypeptide the same as ENZM or a polypeptide with at least one functional characteristic of ENZM. Included within this definition are polymorphisms which may or may not be readily detectable using a particular oligonucleotide probe of

the polynucleotide encoding ENZM, and improper or unexpected hybridization to allelic variants, with a locus other than the normal chromosomal locus for the polynucleotide encoding ENZM. The encoded protein may also be "altered," and may contain deletions, insertions, or substitutions of amino acid residues which produce a silent change and result in a functionally equivalent ENZM. Deliberate amino acid substitutions may be made on the basis of one or more similarities in polarity, charge, solubility, hydrophobicity, hydrophilicity, and/or the amphipathic nature of the residues, as long as the biological or immunological activity of ENZM is retained. For example, negatively charged amino acids may include aspartic acid and glutamic acid, and positively charged amino acids may include aspartic acid swith uncharged polar side chains having similar hydrophilicity values may include: asparagine and glutamine; and serine and threonine. Amino acids with uncharged side chains having similar hydrophilicity values may include: leucine, isoleucine, and valine; glycine and alanine; and phenylalanine and tyrosine.

The terms "amino acid" and "amino acid sequence" can refer to an oligopeptide, a peptide, a
polypeptide, or a protein sequence, or a fragment of any of these, and to naturally occurring or

synthetic molecules. Where "amino acid sequence" is recited to refer to a sequence of a naturally
occurring protein molecule, "amino acid sequence" and like terms are not meant to limit the amino acid
sequence to the complete native amino acid sequence associated with the recited protein molecule.

"Amplification" relates to the production of additional copies of a nucleic acid. Amplification
may be carried out using polymerase chain reaction (PCR) technologies or other nucleic acid
amplification technologies well known in the art.

The term "antagonist" refers to a molecule which inhibits or attenuates the biological activity of ENZM. Antagonists may include proteins such as antibodies, anticalins, nucleic acids, carbohydrates, small molecules, or any other compound or composition which modulates the activity of ENZM either by directly interacting with ENZM or by acting on components of the biological pathway in which ENZM participates.

The term "antibody" refers to intact immunoglobulin molecules as well as to fragments thereof, such as Fab, F(ab')<sub>2</sub>, and Fv fragments, which are capable of binding an epitopic determinant. Antibodies that bind ENZM polypeptides can be prepared using intact polypeptides or using fragments containing small peptides of interest as the immunizing antigen. The polypeptide or oligopeptide used to immunize an animal (e.g., a mouse, a rat, or a rabbit) can be derived from the translation of RNA, or synthesized chemically, and can be conjugated to a carrier protein if desired. Commonly used carriers that are chemically coupled to peptides include bovine serum albumin, thyroglobulin, and keyhole limpet hemocyanin (KLIh). The coupled peptide is then used to immunize the animal.

The term "antigenic determinant" refers to that region of a molecule (i.e., an epitope) that makes contact with a particular antibody. When a protein or a fragment of a protein is used to immunize a host animal, numerous regions of the protein may induce the production of antibodies which bind specifically to antigenic determinants (particular regions or three-dimensional structures on the protein). An antigenic determinant may compete with the intact antigen (i.e., the immunogen used to elicit the immune response) for binding to an antibody.

The term "aptamer" refers to a nucleic acid or oligonucleotide molecule that binds to a specific molecular target. Aptamers are derived from an *in vitro* evolutionary process (e.g., SELEX (Systematic Evolution of Ligands by EXponential Enrichment), described in U.S. Patent No.

10 5,270,163), which selects for target-specific aptamer sequences from large combinatorial libraries. Aptamer compositions may be double-stranded or single-stranded, and may include deoxyribonucleotides, ribonucleotides, nucleotide derivatives, or other nucleotide-like molecules. The nucleotide components of an aptamer may have modified sugar groups (e.g., the 2'-OH group of a ribonucleotide may be replaced by 2'-F or 2'-NH<sub>2</sub>), which may improve a desired property, e.g., resistance to nucleases or longer lifetime in blood. Aptamers may be conjugated to other molecules, e.g., a high molecular weight carrier to slow clearance of the aptamer from the circulatory system. Aptamers may be specifically cross-linked to their cognate ligands, e.g., by photo-activation of a cross-linker (Brody, E.N. and L. Gold (2000) J. Biotechnol. 74:5-13).

The term "intramer" refers to an aptamer which is expressed in vivo. For example, a

vaccinia virus-based RNA expression system has been used to express specific RNA aptamers at
high levels in the cytoplasm of leukocytes (Blind, M. et al. (1999) Proc. Natl. Acad. Sci. USA

96:3606-3610).

The term "spiegelmer" refers to an aptamer which includes L-DNA, L-RNA, or other lefthanded nucleotide derivatives or nucleotide-like molecules. Aptamers containing left-handed 25 nucleotides are resistant to degradation by naturally occurring enzymes, which normally act on substrates containing right-handed nucleotides.

The term "antisense" refers to any composition capable of base-pairing with the "sense" 
(coding) strand of a polynucleotide having a specific nucleic acid sequence. Antisense compositions 
may include DNA; RNA; peptide nucleic acid (PNA); oligonucleotides having modified backbone 
linkages such as phosphorothicates, methylphosphonates, or benzylphosphonates; oligonucleotides 
having modified sugar groups such as 2'-methoxyethyl sugars or 2'-methoxyethoxy sugars; or 
oligonucleotides having modified bases such as 5-methyl cytosine, 2'-deoxyuracil, or 7-deaza-2'deoxyguanosine. Antisense molecules may be produced by any method including chemical synthesis

or transcription. Once introduced into a cell, the complementary antisense molecule base-pairs with a naturally occurring nucleic acid sequence produced by the cell to form duplexes which block either transcription or translation. The designation "negative" or "minus" can refer to the antisense strand, and the designation "positive" or "plus" can refer to the sense strand of a reference DNA molecule.

The term "biologically active" refers to a protein having structural, regulatory, or biochemical functions of a naturally occurring molecule. Likewise, "immunologically active" or "immunogenio" refers to the capability of the natural, recombinant, or synthetic ENZM, or of any oligopeptide thereof, to induce a specific immune response in appropriate animals or cells and to bind with specific antibodies.

"Complementary" describes the relationship between two single-stranded nucleic acid sequences that anneal by base-pairing. For example, 5'-AGT-3' pairs with its complement, 3'-TCA-5'.

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A "composition comprising a given polynucleotide" and a "composition comprising a given polypeptide" can refer to any composition containing the given polynucleotide or polypeptide. The composition may comprise a dry formulation or an aqueous solution. Compositions comprising polynucleotides encoding ENZM or fragments of ENZM may be employed as hybridization probes. The probes may be stored in freeze-dried form and may be associated with a stabilizing agent such as a carbohydrate. In hybridizations, the probe may be deployed in an aqueous solution containing salts (e.g., NaCl), detergents (e.g., sodium dodecyl sulfate; SDS), and other components (e.g., Denhardt's solution, dry milk, salmon sperm DNA, etc.).

"Consensus sequence" refers to a nucleic acid sequence which has been subjected to repeated DNA sequence analysis to resolve uncalled bases, extended using the XL-PCR kit (Applied Biosystems, Foster City CA) in the 5' and/or the 3' direction, and resequenced, or which has been assembled from one or more overlapping cDNA, EST, or genomic DNA fragments using a computer program for fragment assembly, such as the GELVIEW fragment assembly system (Accelrys, Burlington MA) or Phrap (University of Washington, Seattle WA). Some sequences have been both extended and assembled to produce the consensus sequence.

"Conservative amino acid substitutions" are those substitutions that are predicted to least
interfere with the properties of the original protein, i.e., the structure and especially the function of the
protein is conserved and not significantly changed by such substitutions. The table below shows amino
acids which may be substituted for an original amino acid in a protein and which are regarded as
conservative amino acid substitutions.

	Original Residue	Conservative Substitution
	Ala	Gly, Ser
	Arg	His, Lys
	Asn	Asp, Gln, His
5	Asp	Asn, Glu
	Cys	Ala, Ser
	Gln	Asn, Glu, His
	Glu	Asp, Gln, His
	Gly	Ala
10	His	Asn, Arg, Gln, Glu
	Ile	Leu, Val
	Leu	Ile, Val
	Lys	Arg, Gln, Glu
	Met	Leu, Ile
15	Phe	His, Met, Leu, Trp, Tyr
	Ser	Cys, Thr
	Thr	Ser, Val
	Trp	Phe, Tyr
	Tyr	His, Phe, Trp
20	Val	Ile, Leu, Thr

Conservative amino acid substitutions generally maintain (a) the structure of the polypeptide backbone in the area of the substitution, for example, as a beta sheet or alpha helical conformation, (b) the charge or hydrophobicity of the molecule at the site of the substitution, and/or (c) the bulk of the side chain.

A "deletion" refers to a change in the amino acid or nucleotide sequence that results in the absence of one or more amino acid residues or nucleotides.

The term "derivative" refers to a chemically modified polynucleotide or polypeptide.

Chemical modifications of a polynucleotide can include, for example, replacement of hydrogen by an alkyl, acyl, hydroxyl, or amino group. A derivative polynucleotide encodes a polypeptide which retains at least one biological or immunological function of the natural molecule. A derivative polypeptide is one modified by glycosylation, pegylation, or any similar process that retains at least one biological or immunological function of the polypeptide from which it was derived.

A "detectable label" refers to a reporter molecule or enzyme that is capable of generating a measurable signal and is covalently or noncovalently joined to a polynucleotide or polypeptide.

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"Differential expression" refers to increased or upregulated; or decreased, downregulated, or absent gene or protein expression, determined by comparing at least two different samples. Such comparisons may be carried out between, for example, a treated and an untreated sample, or a diseased and a normal sample.

"Exon shuffling" refers to the recombination of different coding regions (exons). Since an

exon may represent a structural or functional domain of the encoded protein, new proteins may be assembled through the novel reassortment of stable substructures, thus allowing acceleration of the evolution of new protein functions.

A "fragment" is a unique portion of ENZM or a polynucleotide encoding ENZM which can be identical in sequence to, but shorter in length than, the parent sequence. A fragment may comprise up to the entire length of the defined sequence, minus one nucleotide/amino acid residue. For example, a fragment may comprise from about 5 to about 1000 contiguous nucleotides or amino acid residues. A fragment used as a probe, primer, antigen, therapeutic molecule, or for other purposes, may be at least 5, 10, 15, 16, 20, 25, 30, 40, 50, 60, 75, 100, 150, 250 or at least 500 contiguous nucleotides or amino acid residues in length. Fragments may be preferentially selected from certain regions of a molecule. For example, a polypeptide fragment may comprise a certain length of contiguous amino acids selected from the first 250 or 500 amino acids (or first 25% or 50%) of a polypeptide as shown in a certain defined sequence. Clearly these lengths are exemplary, and any length that is supported by the specification, including the Sequence Listing, tables, and figures, may be encompassed by the present 15 embodiments.

A fragment of SEQ ID NO:39-76 can comprise a region of unique polynucleotide sequence that specifically identifies SEQ ID NO:39-76, for example, as distinct from any other sequence in the genome from which the fragment was obtained. A fragment of SEQ ID NO:39-76 can be employed in one or more embodiments of methods of the invention, for example, in hybridization and amplification technologies and in analogous methods that distinguish SEQ ID NO:39-76 from related polynucleotides. The precise length of a fragment of SEQ ID NO:39-76 and the region of SEQ ID NO:39-76 to which the fragment corresponds are routinely determinable by one of ordinary skill in the art based on the intended purpose for the fragment.

A fragment of SEQ ID NO:1-38 is encoded by a fragment of SEQ ID NO:39-76. A

fragment of SEQ ID NO:1-38 can comprise a region of unique amino acid sequence that specifically identifies SEQ ID NO:1-38. For example, a fragment of SEQ ID NO:1-38 can be used as an immunogenic peptide for the development of antibodies that specifically recognize SEQ ID NO:1-38. The precise length of a fragment of SEQ ID NO:1-38 and the region of SEQ ID NO:1-38 to which the fragment corresponds can be determined based on the intended purpose for the fragment using

one or more analytical methods described herein or otherwise known in the art.

A "full length" polynucleotide is one containing at least a translation initiation codon (e.g., methionine) followed by an open reading frame and a translation termination codon. A "full length" polynucleotide sequence encodes a "full length" polynucleotide sequence.

"Homology" refers to sequence similarity or, alternatively, sequence identity, between two or more polynucleotide sequences or two or more polypeptide sequences.

The terms "percent identity" and "% identity," as applied to polynucleotide sequences, refer to the percentage of identical nucleotide matches between at least two polynucleotide sequences aligned using a standardized algorithm. Such an algorithm may insert, in a standardized and reproducible way, gaps in the sequences being compared in order to optimize alignment between two sequences, and therefore achieve a more meaningful comparison of the two sequences.

Percent identity between polynucleotide sequences may be determined using one or more computer algorithms or programs known in the art or described herein. For example, percent identity 10 can be determined using the default parameters of the CLUSTAL V algorithm as incorporated into the MEGALIGN version 3.12e sequence alignment program. This program is part of the LASERGENE software package, a suite of molecular biological analysis programs (DNASTAR, Madison WI). CLUSTAL V is described in Higgins, D.G. and P.M. Sharp (1989; CABIOS 5:151-153) and in Higgins, D.G. et al. (1992; CABIOS 8:189-191). For pairwise alignments of 15 polynucleotide sequences, the default parameters are set as follows: Ktuple=2, gap penalty=5, window=4, and "diagonals saved"=4. The "weighted" residue weight table is selected as the default. Alternatively, a suite of commonly used and freely available sequence comparison algorithms which can be used is provided by the National Center for Biotechnology Information (NCBI) Basic Local Alignment Search Tool (BLAST) (Altschul, S.F. et al. (1990) J. Mol. Biol. 215:403-410), which 20 is available from several sources, including the NCBI, Bethesda, MD, and on the Internet at http://www.ncbi.nlm.nih.gov/BLAST/. The BLAST software suite includes various sequence analysis programs including "blastn," that is used to align a known polynucleotide sequence with other polynucleotide sequences from a variety of databases. Also available is a tool called "BLAST 2 Sequences" that is used for direct pairwise comparison of two nucleotide sequences. "BLAST 2 25 Sequences" can be accessed and used interactively at http://www.ncbi.nlm.nih.gov/gorf/bl2.html. The "BLAST 2 Sequences" tool can be used for both blastn and blastp (discussed below). BLAST programs are commonly used with gap and other parameters set to default settings. For example, to compare two nucleotide sequences, one may use blastn with the "BLAST 2 Sequences" tool Version 2.0.12 (April-21-2000) set at default parameters. Such default parameters may be, for example:

Matrix: BLOSUM62

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Reward for match: 1

Penalty for mismatch: -2

Open Gap: 5 and Extension Gap: 2 penalties

Gap x drop-off: 50
Expect: 10
Word Size: 11
Filter: on

Percent identity may be measured over the length of an entire defined sequence, for example, as defined by a particular SEQ ID number, or may be measured over a shorter length, for example, over the length of a fragment taken from a larger, defined sequence, for instance, a fragment of at least 20, at least 30, at least 40, at least 50, at least 70, at least 100, or at least 200 contiguous nucleotides. Such lengths are exemplary only, and it is understood that any fragment length supported by the sequences shown herein, in the tables, figures, or Sequence Listing, may be used to describe a length over which percentage identity may be measured.

Nucleic acid sequences that do not show a high degree of identity may nevertheless encode
similar amino acid sequences due to the degeneracy of the genetic code. It is understood that changes
in a nucleic acid sequence can be made using this degeneracy to produce multiple nucleic acid
sequences that all encode substantially the same protein.

The phrases "percent identity" and "% identity," as applied to polypeptide sequences, refer to the percentage of identical residue matches between at least two polypeptide sequences aligned using a standardized algorithm. Methods of polypeptide sequence alignment are well-known. Some alignment methods take into account conservative amino acid substitutions. Such conservative 20 substitutions, explained in more detail above, generally preserve the charge and hydrophobicity at the site of substitution, thus preserving the structure (and therefore function) of the polypeptide. The phrases "percent similarity" and "% similarity," as applied to polypeptide sequences, refer to the percentage of residue matches, including identical residue matches and conservative substitutions, between at least two polypeptide sequences aligned using a standardized algorithm. In contrast, conservative substitutions are not included in the calculation of percent identity between polypeptide sequences.

Percent identity between polypeptide sequences may be determined using the default parameters of the CLUSTAL V algorithm as incorporated into the MEGALIGN version 3.12e sequence alignment program (described and referenced above). For pairwise alignments of polypeptide sequences using CLUSTAL V, the default parameters are set as follows: Ktuple=1, gap penalty=3, window=5, and "diagonals saved"=5. The PAM250 matrix is selected as the default residue weight table.

Alternatively the NCBI BLAST software suite may be used. For example, for a pairwise

comparison of two polypeptide sequences, one may use the "BLAST 2 Sequences" tool Version 2.0.12 (April-21-2000) with blastp set at default parameters. Such default parameters may be, for example:

Matrix: BLOSUM62

Open Gap: 11 and Extension Gap: 1 penalties

Gap x drop-off: 50 Expect: 10 Word Size: 3

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Filter: on

Percent identity may be measured over the length of an entire defined polypeptide sequence, for example, as defined by a particular SEQ ID number, or may be measured over a shorter length, for example, over the length of a fragment taken from a larger, defined polypeptide sequence, for instance, a fragment of at least 15, at least 20, at least 30, at least 40, at least 50, at least 70 or at least 150 contiguous residues. Such lengths are exemplary only, and it is understood that any fragment 15 length supported by the sequences shown herein, in the tables, figures or Sequence Listing, may be used to describe a length over which percentage identity may be measured.

"Human artificial chromosomes" (HACs) are linear microchromosomes which may contain DNA sequences of about 6 kb to 10 Mb in size and which contain all of the elements required for chromosome replication, segregation and maintenance.

The term "humanized antibody" refers to an antibody molecule in which the amino acid sequence in the non-antigen binding regions has been altered so that the antibody more closely resembles a human antibody, and still retains its original binding ability.

"Hybridization" refers to the process by which a polynucleotide strand anneals with a complementary strand through base pairing under defined hybridization conditions. Specific 25 hybridization is an indication that two nucleic acid sequences share a high degree of complementarity. Specific hybridization complexes form under permissive annealing conditions and remain hybridized after the "washing" step(s). The washing step(s) is particularly important in determining the stringency of the hybridization process, with more stringent conditions allowing less non-specific binding, i.e., binding between pairs of nucleic acid strands that are not perfectly matched. Permissive 30 conditions for annealing of nucleic acid sequences are routinely determinable by one of ordinary skill in the art and may be consistent among hybridization experiments, whereas wash conditions may be varied among experiments to achieve the desired stringency, and therefore hybridization specificity. Permissive annealing conditions occur, for example, at 68°C in the presence of about 6 x SSC, about

1% (w/v) SDS, and about 100 μg/ml sheared, denatured salmon sperm DNA.

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Generally, stringency of hybridization is expressed, in part, with reference to the temperature under which the wash step is carried out. Such wash temperatures are typically selected to be about 5°C to 20°C lower than the thermal melting point (Tm) for the specific sequence at a defined ionic 5 strength and pH. The T<sub>m</sub> is the temperature (under defined ionic strength and pH) at which 50% of the target sequence hybridizes to a perfectly matched probe. An equation for calculating T<sub>m</sub> and conditions for nucleic acid hybridization are well known and can be found in Sambrook, J. and D.W. Russell (2001; Molecular Cloning: A Laboratory Manual, 3rd ed., vol. 1-3, Cold Spring Harbor Press, Cold Spring Harbor NY, ch. 9).

High stringency conditions for hybridization between polynucleotides of the present invention include wash conditions of 68°C in the presence of about 0.2 x SSC and about 0.1% SDS, for 1 hour. Alternatively, temperatures of about 65°C, 60°C, 55°C, or 42°C may be used. SSC concentration may be varied from about 0.1 to 2 x SSC, with SDS being present at about 0.1%. Typically, blocking reagents are used to block non-specific hybridization. Such blocking reagents include, for instance, 15 sheared and denatured salmon sperm DNA at about 100-200 μg/ml. Organic solvent, such as formamide at a concentration of about 35-50% v/v, may also be used under particular circumstances, such as for RNA:DNA hybridizations. Useful variations on these wash conditions will be readily apparent to those of ordinary skill in the art. Hybridization, particularly under high stringency conditions, may be suggestive of evolutionary similarity between the nucleotides. Such similarity is 20 strongly indicative of a similar role for the nucleotides and their encoded polypeptides.

The term "hybridization complex" refers to a complex formed between two nucleic acids by virtue of the formation of hydrogen bonds between complementary bases. A hybridization complex may be formed in solution (e.g., Cot or Rot analysis) or formed between one nucleic acid present in solution and another nucleic acid immobilized on a solid support (e.g., paper, membranes, filters, chips, 25 pins or glass slides, or any other appropriate substrate to which cells or their nucleic acids have been fixed).

The words "insertion" and "addition" refer to changes in an amino acid or polynucleotide sequence resulting in the addition of one or more amino acid residues or nucleotides, respectively.

"Immune response" can refer to conditions associated with inflammation, trauma, immune 30 disorders, or infectious or genetic disease, etc. These conditions can be characterized by expression of various factors, e.g., cytokines, chemokines, and other signaling molecules, which may affect cellular and systemic defense systems.

An "immunogenic fragment" is a polypeptide or oligopeptide fragment of ENZM which is

capable of eliciting an immune response when introduced into a living organism, for example, a mammal. The term "immunogenic fragment" also includes any polypeptide or oligopeptide fragment of ENZM which is useful in any of the antibody production methods disclosed herein or known in the art.

5 The term "microarray" refers to an arrangement of a plurality of polynucleotides, polypeptides, antibodies, or other chemical compounds on a substrate.

The terms "element" and "array element" refer to a polynucleotide, polypeptide, antibody, or other chemical compound having a unique and defined position on a microarray.

The term "modulate" refers to a change in the activity of ENZM. For example, modulation

may cause an increase or a decrease in protein activity, binding characteristics, or any other biological, functional, or immunological properties of ENZM.

The phrases "nucleic acid" and "nucleic acid sequence" refer to a nucleotide, oligonucleotide, polynucleotide, or any fragment thereof. These phrases also refer to DNA or RNA of genomic or synthetic origin which may be single-stranded or double-stranded and may represent the sense or the

antisense strand, to peptide nucleic acid (PNA), or to any DNA-like or RNA-like material.

"Operably linked" refers to the situation in which a first nucleic acid sequence is placed in a
functional relationship with a second nucleic acid sequence. For instance, a promoter is operably
linked to a coding sequence if the promoter affects the transcription or expression of the coding
sequence. Operably linked DNA sequences may be in close proximity or contiguous and, where
necessary to join two protein coding regions, in the same reading frame.

"Peptide nucleic acid" (PNA) refers to an antisense molecule or anti-gene agent which comprises an oligonucleotide of at least about 5 nucleotides in length linked to a peptide backbone of amino acid residues ending in lysine. The terminal lysine confers solubility to the composition. PNAs preferentially bind complementary single stranded DNA or RNA and stop transcript elongation, and may be pegylated to extend their lifespan in the cell.

"Post-translational modification" of an ENZM may involve lipidation, glycosylation, phosphorylation, acetylation, racemization, proteolytic cleavage, and other modifications known in the art. These processes may occur synthetically or biochemically. Biochemical modifications will vary by cell type depending on the enzymatic milieu of ENZM.

"Probe" refers to nucleic acids encoding ENZM, their complements, or fragments thereof, which are used to detect identical, allelic or related nucleic acids. Probes are isolated oligonucleotides or polynucleotides attached to a detectable label or reporter molecule. Typical labels include radioactive isotopes, ligands, chemiluminescent agents, and enzymes. "Primers" are short nucleic

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acids, usually DNA oligonucleotides, which may be annealed to a target polynucleotide by complementary base-pairing. The primer may then be extended along the target DNA strand by a DNA polymerase enzyme. Primer pairs can be used for amplification (and identification) of a nucleic acid, e.g., by the polymerase chain reaction (PCR).

Probes and primers as used in the present invention typically comprise at least 15 contiguous nucleotides of a known sequence. In order to enhance specificity, longer probes and primers may also be employed, such as probes and primers that comprise at least 20, 25, 30, 40, 50, 60, 70, 80, 90, 100, or at least 150 consecutive nucleotides of the disclosed nucleic acid sequences. Probes and primers may be considerably longer than these examples, and it is understood that any length supported by the 10 specification, including the tables, figures, and Sequence Listing, may be used.

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Methods for preparing and using probes and primers are described in, for example, Sambrook, J. and D.W. Russell (2001; Molecular Cloning: A Laboratory Manual, 3rd ed., vol. 1-3, Cold Spring Harbor Press, Cold Spring Harbor NY), Ausubel, F.M. et al. (1999; Short Protocols in Molecular Biology, 4th ed., John Wiley & Sons, New York NY), and Innis, M. et al. (1990; PCR Protocols, A 15 Guide to Methods and Applications, Academic Press, San Diego CA). PCR primer pairs can be derived from a known sequence, for example, by using computer programs intended for that purpose such as Primer (Version 0.5, 1991, Whitehead Institute for Biomedical Research, Cambridge MA).

Oligonucleotides for use as primers are selected using software known in the art for such purpose. For example, OLIGO 4.06 software is useful for the selection of PCR primer pairs of up to 20 100 nucleotides each, and for the analysis of oligonucleotides and larger polynucleotides of up to 5,000 nucleotides from an input polynucleotide sequence of up to 32 kilobases. Similar primer selection programs have incorporated additional features for expanded capabilities. For example, the PrimOU primer selection program (available to the public from the Genome Center at University of Texas South West Medical Center, Dallas TX) is capable of choosing specific primers from megabase 25 sequences and is thus useful for designing primers on a genome-wide scope. The Primer3 primer selection program (available to the public from the Whitehead Institute/MIT Center for Genome Research, Cambridge MA) allows the user to input a "mispriming library," in which sequences to avoid as primer binding sites are user-specified. Primer3 is useful, in particular, for the selection of oligonucleotides for microarrays. (The source code for the latter two primer selection programs may also be obtained from their respective sources and modified to meet the user's specific needs.) The PrimeGen program (available to the public from the UK Human Genome Mapping Project Resource Centre, Cambridge UK) designs primers based on multiple sequence alignments, thereby allowing selection of primers that hybridize to either the most conserved or least conserved regions of aligned

nucleic acid sequences. Hence, this program is useful for identification of both unique and conserved oligonucleotides and polynucleotide fragments. The oligonucleotides and polynucleotide fragments identified by any of the above selection methods are useful in hybridization technologies, for example, as PCR or sequencing primers, microarray elements, or specific probes to identify fully or partially complementary polynucleotides in a sample of nucleic acids. Methods of oligonucleotide selection are not limited to those described above.

A "recombinant nucleic acid" is a nucleic acid that is not naturally occurring or has a sequence that is made by an artificial combination of two or more otherwise separated segments of sequence. This artificial combination is often accomplished by chemical synthesis or, more commonly, by the artificial manipulation of isolated segments of nucleic acids, e.g., by genetic engineering techniques such as those described in Sambrook and Russell (supra). The term recombinant includes nucleic acids that have been altered solely by addition, substitution, or deletion of a portion of the nucleic acid. Frequently, a recombinant nucleic acid may include a nucleic acid sequence operably linked to a promoter sequence. Such a recombinant nucleic acid may be part of a vector that is used, 15 for example, to transform a cell.

Alternatively, such recombinant nucleic acids may be part of a viral vector, e.g., based on a vaccinia virus, that could be use to vaccinate a mammal wherein the recombinant nucleic acid is expressed, inducing a protective immunological response in the mammal.

A "regulatory element" refers to a nucleic acid sequence usually derived from untranslated regions of a gene and includes enhancers, promoters, introns, and 5' and 3' untranslated regions (UTRs). Regulatory elements interact with host or viral proteins which control transcription, translation, or RNA stability.

"Reporter molecules" are chemical or biochemical moieties used for labeling a nucleic acid, amino acid, or antibody. Reporter molecules include radionuclides; enzymes; fluorescent,

chemiluminescent, or chromogenic agents; substrates; cofactors; inhibitors; magnetic particles; and other moieties known in the art.

An "RNA equivalent," in reference to a DNA molecule, is composed of the same linear
sequence of nucleotides as the reference DNA molecule with the exception that all occurrences of
the nitrogenous base thymine are replaced with uracil, and the sugar backbone is composed of ribose
instead of deoxyribose.

The term "sample" is used in its broadest sense. A sample suspected of containing ENZM, nucleic acids encoding ENZM, or fragments thereof may comprise a bodily fluid; an extract from a cell, chromosome, organelle, or membrane isolated from a cell; a cell; genomic DNA, RNA, or cDNA,

in solution or bound to a substrate; a tissue; a tissue print; etc.

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The terms "specific binding" and "specifically binding" refer to that interaction between a protein or peptide and an agonist, an antibody, an antagonist, a small molecule, or any natural or synthetic binding composition. The interaction is dependent upon the presence of a particular structure of the protein, e.g., the antigenic determinant or epitope, recognized by the binding molecule. For example, if an antibody is specific for epitope "A," the presence of a polypeptide comprising the epitope A, or the presence of free unlabeled A, in a reaction containing free labeled A and the antibody will reduce the amount of labeled A that binds to the antibody.

The term "substantially purified" refers to nucleic acid or amino acid sequences that are

removed from their natural environment and are isolated or separated, and are at least about 60%

free, preferably at least about 75% free, and most preferably at least about 90% free from other

components with which they are naturally associated.

A "substitution" refers to the replacement of one or more amino acid residues or nucleotides by different amino acid residues or nucleotides, respectively.

"Substrate" refers to any suitable rigid or semi-rigid support including membranes, filters, chips, slides, wafers, fibers, magnetic or nonmagnetic beads, gels, tubing, plates, polymers, microparticles and capillaries. The substrate can have a variety of surface forms, such as wells, trenches, pins, channels and pores, to which polynucleotides or polypeptides are bound.

A "transcript image" or "expression profile" refers to the collective pattern of gene expression

by a particular cell type or tissue under given conditions at a given time.

"Transformation" describes a process by which exogenous DNA is introduced into a recipient cell. Transformation may occur under natural or artificial conditions according to various methods well known in the art, and may rely on any known method for the insertion of foreign nucleic acid sequences into a prokaryotic or eukaryotic host cell. The method for transformation is selected based on the type of host cell being transformed and may include, but is not limited to, bacteriophage or viral infection, electroporation, heat shock, lipofection, and particle bombardment. The term "transformed cells" includes stably transformed cells in which the inserted DNA is capable of replication either as an autonomously replicating plasmid or as part of the host chromosome, as well as transiently transformed cells which express the inserted DNA or RNA for limited periods of time.

A "transgenic organism," as used herein, is any organism, including but not limited to animals and plants, in which one or more of the cells of the organism contains heterologous nucleic acid introduced by way of human intervention, such as by transgenic techniques well known in the art. The nucleic acid is introduced into the cell, directly or indirectly by introduction into a precursor of the cell,

by way of deliberate genetic manipulation, such as by microinjection or by infection with a recombinant virus. In another embodiment, the nucleic acid can be introduced by infection with a recombinant viral vector, such as a lentiviral vector (Lois, C. et al. (2002) Science 295:868-872). The term genetic manipulation does not include classical cross-breeding, or in vitro fertilization, but rather 5 is directed to the introduction of a recombinant DNA molecule. The transgenic organisms contemplated in accordance with the present invention include bacteria, cyanobacteria, fungi, plants and animals. The isolated DNA of the present invention can be introduced into the host by methods known in the art, for example infection, transfection, transformation or transconjugation. Techniques for transferring the DNA of the present invention into such organisms are widely known and provided in references such as Sambrook and Russell (supra).

A "variant" of a particular nucleic acid sequence is defined as a nucleic acid sequence having at least 40% sequence identity to the particular nucleic acid sequence over a certain length of one of the nucleic acid sequences using blastn with the "BLAST 2 Sequences" tool Version 2.0.9 (May-07-1999) set at default parameters. Such a pair of nucleic acids may show, for example, at least 50%, at 15 least 60%, at least 70%, at least 80%, at least 85%, at least 90%, at least 91%, at least 92%, at least 93%, at least 94%, at least 95%, at least 96%, at least 97%, at least 98%, or at least 99% or greater sequence identity over a certain defined length. A variant may be described as, for example, an "allelic" (as defined above), "splice," "species," or "polymorphic" variant. A splice variant may have significant identity to a reference molecule, but will generally have a greater or lesser number of 20 polynucleotides due to alternate splicing during mRNA processing. The corresponding polypeptide may possess additional functional domains or lack domains that are present in the reference molecule. Species variants are polynucleotides that vary from one species to another. The resulting polypeptides will generally have significant amino acid identity relative to each other. A polymorphic variant is a variation in the polynucleotide sequence of a particular gene between individuals of a given species. Polymorphic variants also may encompass "single nucleotide polymorphisms" (SNPs) in which the polynucleotide sequence varies by one nucleotide base. The presence of SNPs may be indicative of, for example, a certain population, a disease state, or a propensity for a disease state.

A "variant" of a particular polypeptide sequence is defined as a polypeptide sequence having at least 40% sequence identity or sequence similarity to the particular polypeptide sequence over a certain length of one of the polypeptide sequences using blastp with the "BLAST 2 Sequences" tool Version 2.0.9 (May-07-1999) set at default parameters. Such a pair of polypeptides may show, for example, at least 50%, at least 60%, at least 70%, at least 80%, at least 85%, at least 90%, at least 91%, at least 92%, at least 94%, at least 95%, at least 96%, at least 97%, at least 98%,

or at least 99% or greater sequence identity or sequence similarity over a certain defined length of one of the polypeptides.

## THE INVENTION

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Various embodiments of the invention include new human enzymes (ENZM), the polynucleotides encoding ENZM, and the use of these compositions for the diagnosis, treatment, or prevention of autoimmune/inflammatory disorders, infectious disorders, immune deficiencies, disorders of metabolism, reproductive disorders, neurological disorders, cardiovascular disorders, eye disorders, and cell proliferative disorders, including cancer.

Table 1 summarizes the nomenclature for the full length polynucleotide and polypeptide embodiments of the invention. Each polynucleotide and its corresponding polypeptide are correlated to a single Incyte project identification number (Incyte Project ID). Each polypeptide sequence is denoted by both a polypeptide sequence identification number (Polypeptide SEQ ID NO:) and an Incyte polypeptide sequence number (Incyte Polypeptide ID) as shown. Each polynucleotide 15 sequence is denoted by both a polynucleotide sequence identification number (Polynucleotide SEQ ID NO:) and an Incyte polynucleotide consensus sequence number (Incyte Polynucleotide ID) as shown.

Table 2 shows sequences with homology to polypeptide embodiments of the invention as identified by BLAST analysis against the GenBank protein (genpept) database and the PROTEOME database. Columns 1 and 2 show the polypeptide sequence identification number (Polypeptide SEQ 20 ID NO:) and the corresponding Incyte polypeptide sequence number (Incyte Polypeptide ID) for polypeptides of the invention. Column 3 shows the GenBank identification number (GenBank ID NO:) of the nearest GenBank homolog and the PROTEOME database identification numbers (PROTEOME ID NO:) of the nearest PROTEOME database homologs. Column 4 shows the probability scores for the matches between each polypeptide and its homolog(s). Column 5 shows the 25 annotation of the GenBank and PROTEOME database homolog(s) along with relevant citations where applicable, all of which are expressly incorporated by reference herein.

Table 3 shows various structural features of the polypeptides of the invention. Columns 1 and 2 show the polypeptide sequence identification number (SEQ ID NO:) and the corresponding Incyte polypeptide sequence number (Incyte Polypeptide ID) for each polypeptide of the invention. Column 30 3 shows the number of amino acid residues in each polypeptide. Column 4 shows potential phosphorylation sites, and column 5 shows potential glycosylation sites, as determined by the MOTIFS program of the GCG sequence analysis software package (Accelrys, Burlington MA). Column 6 shows amino acid residues comprising signature sequences, domains, and motifs. Column 7 shows

analytical methods for protein structure/function analysis and in some cases, searchable databases to which the analytical methods were applied.

Together, Tables 2 and 3 summarize the properties of polypeptides of the invention, and these properties establish that the claimed polypeptides are enzymes. For example, SEQ ID NO:3 is 100% 5 identical, from residue M1 to residue I539, to human acetolactate synthase (GenBank ID g2725625) as determined by the Basic Local Alignment Search Tool (BLAST). (See Table 2.) The BLAST probability score is 2.0E-287, which indicates the probability of obtaining the observed polypeptide sequence alignment by chance. SEQ ID NO:3 also has homology to proteins that are localized to the cytoplasm and are lyases, as determined by BLAST analysis using the PROTEOME database. SEQ 10 ID NO:3 also contains a thiamine pyrophosphate enzyme domain and a thiamine pyrophosphate enzyme N-terminus domain as determined by searching for statistically significant matches in the hidden Markov model (HMM)-based PFAM database of conserved protein families/domains, and a 2-succinyl-6-hydroxy-2,4-cyclohexadiene-1-carboxylic acid synthase/2-oxoglutarate decarboxylase domain as determined by searching for statistically significant matches in the hidden Markov model 15 (HMM)-based TIGRFAM database. (See Table 3.) Data from BLIMPS, MOTIFS, and PROFILESCAN analyses, and BLAST analyses against the PRODOM and DOMO databases, provide further corroborative evidence that SEQ ID NO:3 is an acetolactate synthase. In another example, SEQ ID NO:20 is 90% identical, from residue W142 to residue Y293, to human aldose reductase-like peptide (GenBank ID g3150035) as determined by the Basic Local Alignment Search Tool (BLAST). (See Table 2.) The BLAST probability score is 1.9E-146, which indicates the probability of obtaining the observed polypeptide sequence alignment by chance. SEQ ID NO:20 also has homology to human small intestine reductase, as determined by BLAST analysis using the PROTEOME database. SEQ ID NO:20 also contains an aldo/keto reductase family domain as determined by searching for statistically significant matches in the hidden Markov model (HMM)-25 based PFAM database of conserved protein families/domains. (See Table 3.) Data from BLIMPS, MOTIFS, and PROFILESCAN analyses, and BLAST analyses against the PRODOM and DOMO databases, provide further corroborative evidence that SEQ ID NO:20 is an aldo/keto reductase. SEQ ID NO:1-2, SEQ ID NO:4-19, and SEQ ID NO:21-38 were analyzed and annotated in a similar manner. The algorithms and parameters for the analysis of SEQ ID NO:1-38 are described in Table 7. 30

As shown in Table 4, the full length polynucleotide embodiments were assembled using cDNA sequences or coding (exon) sequences derived from genomic DNA, or any combination of these two types of sequences. Column 1 lists the polynucleotide sequence identification number (Polynucleotide)

SEQ ID NO:), the corresponding Incyte polynucleotide consensus sequence number (Incyte ID) for each polynucleotide of the invention, and the length of each polynucleotide sequence in basepairs.

Column 2 shows the nucleotide start (5') and stop (3') positions of the cDNA and/or genomic sequences used to assemble the full length polynucleotide embodiments, and of fragments of the polynucleotides which are useful, for example, in hybridization or amplification technologies that identify SEQ ID NO:39-76 or that distinguish between SEQ ID NO:39-76 and related polynucleotides.

The polynucleotide fragments described in Column 2 of Table 4 may refer specifically, for example, to Incyte cDNAs derived from tissue-specific cDNA libraries or from pooled cDNA libraries. Alternatively, the polynucleotide fragments described in column 2 may refer to GenBank 10 cDNAs or ESTs which contributed to the assembly of the full length polynucleotides. In addition, the polynucleotide fragments described in column 2 may identify sequences derived from the ENSEMBL (The Sanger Centre, Cambridge, UK) database (i.e., those sequences including the designation "ENST"). Alternatively, the polynucleotide fragments described in column 2 may be derived from the NCBI RefSeq Nucleotide Sequence Records Database (i.e., those sequences including the 15 designation "NM" or "NT") or the NCBI RefSeq Protein Sequence Records (i.e., those sequences including the designation "NP"). Alternatively, the polynucleotide fragments described in column 2 may refer to assemblages of both cDNA and Genscan-predicted exons brought together by an "exon stitching" algorithm. For example, a polynucleotide sequence identified as FL\_XXXXXX\_N<sub>1</sub>\_N<sub>2</sub>\_YYYYY\_N<sub>3</sub>\_N<sub>4</sub> represents a "stitched" sequence in which XXXXXX is the 20 identification number of the cluster of sequences to which the algorithm was applied, and YYYYYY is the number of the prediction generated by the algorithm, and  $N_{123...}$ , if present, represent specific exons that may have been manually edited during analysis (See Example V). Alternatively, the polynucleotide fragments in column 2 may refer to assemblages of exons brought together by an "exon-stretching" algorithm. For example, a polynucleotide sequence identified as 25 FLXXXXXX\_gAAAAA\_gBBBBB\_1\_N is a "stretched" sequence, with XXXXXX being the Incyte project identification number, gAAAAA being the GenBank identification number of the human genomic sequence to which the "exon-stretching" algorithm was applied, gBBBBB being the GenBank identification number or NCBI RefSeq identification number of the nearest GenBank protein homolog, and N referring to specific exons (See Example V). In instances where a RefSeq sequence was used 30 as a protein homolog for the "exon-stretching" algorithm, a RefSeq identifier (denoted by "NM," "NP," or "NT") may be used in place of the GenBank identifier (i.e., gBBBBB).

Alternatively, a prefix identifies component sequences that were hand-edited, predicted from genomic DNA sequences, or derived from a combination of sequence analysis methods. The

following Table lists examples of component sequence prefixes and corresponding sequence analysis methods associated with the prefixes (see Example IV and Example V).

Prefix	Type of analysis and/or examples of programs	
GNN, GFG,	Exon prediction from genomic sequences using, for example,	
ENST	GENSCAN (Stanford University, CA, USA) or FGENES	
	(Computer Genomics Group, The Sanger Centre, Cambridge, UK).	
GBI	Hand-edited analysis of genomic sequences.	
FL	Stitched or stretched genomic sequences (see Example V).	
INCY	Full length transcript and exon prediction from mapping of EST	
	sequences to the genome. Genomic location and EST composition	
	data are combined to predict the exons and resulting transcript.	

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In some cases, Incyte cDNA coverage redundant with the sequence coverage shown in

Table 4 was obtained to confirm the final consensus polynucleotide sequence, but the relevant Incyte
cDNA identification numbers are not shown.

Table 5 shows the representative cDNA libraries for those full length polynucleotides which
were assembled using Incyte cDNA sequences. The representative cDNA library is the Incyte

cDNA library which is most frequently represented by the Incyte cDNA sequences which were used
to assemble and confirm the above polynucleotides. The tissues and vectors which were used to
construct the cDNA libraries shown in Table 5 are described in Table 6.

Table 8 shows single nucleotide polymorphisms (SNPs) found in polynucleotide sequences of the invention, along with allele frequencies in different human populations. Columns 1 and 2 show the polynucleotide sequence identification number (SEQ ID NO:) and the corresponding Incyte project identification number (PID) for polynucleotides of the invention. Column 3 shows the Incyte identification number for the EST in which the SNP was detected (EST ID), and column 4 shows the identification number for the SNP (SNP ID). Column 5 shows the position within the EST sequence at which the SNP is located (EST SNP), and column 6 shows the position of the SNP within the full-length polynucleotide sequence (CB1 SNP). Column 7 shows the allefound in the EST sequence. Columns 8 and 9 show the two alleles found at the SNP site. Column 10 shows the amino acid encoded by the codon including the SNP site, based upon the allele found in the EST. Columns 11-14 show the frequency of allele 1 in four different human populations. An entry of n/d (not detected) indicates that the frequency of allele 1 in the population was too low to be detected, while n/a (not available) indicates that the allele frequency was not determined for the population.

The invention also encompasses ENZM variants. Various embodiments of ENZM variants can have at least about 80%, at least about 90%, or at least about 95% amino acid sequence identity to the ENZM amino acid sequence, and can contain at least one functional or structural characteristic of ENZM.

Various embodiments also encompass polynucleotides which encode ENZM. In a particular embodiment, the invention encompasses a polynucleotide sequence comprising a sequence selected from the group consisting of SEQ ID NO:39-76, which encodes ENZM. The polynucleotide sequences of SEO ID NO:39-76, as presented in the Sequence Listing, embrace the equivalent RNA sequences, wherein occurrences of the nitrogenous base thymine are replaced with uracil, and the 10 sugar backbone is composed of ribose instead of deoxyribose.

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The invention also encompasses variants of a polynucleotide encoding ENZM. In particular, such a variant polynucleotide will have at least about 70%, or alternatively at least about 85%, or even at least about 95% polynucleotide sequence identity to a polynucleotide encoding ENZM. A particular aspect of the invention encompasses a variant of a polynucleotide comprising a sequence selected 15 from the group consisting of SEQ ID NO:39-76 which has at least about 70%, or alternatively at least about 85%, or even at least about 95% polynucleotide sequence identity to a nucleic acid sequence selected from the group consisting of SEQ ID NO:39-76. Any one of the polynucleotide variants described above can encode a polypeptide which contains at least one functional or structural characteristic of ENZM.

In addition, or in the alternative, a polynucleotide variant of the invention is a splice variant of a polynucleotide encoding ENZM. A splice variant may have portions which have significant sequence identity to a polynucleotide encoding ENZM, but will generally have a greater or lesser number of nucleotides due to additions or deletions of blocks of sequence arising from alternate splicing during mRNA processing. A splice variant may have less than about 70%, or alternatively less than about 25 60%, or alternatively less than about 50% polynucleotide sequence identity to a polynucleotide encoding ENZM over its entire length; however, portions of the splice variant will have at least about 70%, or alternatively at least about 85%, or alternatively at least about 95%, or alternatively 100% polynucleotide sequence identity to portions of the polynucleotide encoding ENZM. For example, a polynucleotide comprising a sequence of SEQ ID NO:52 and a polynucleotide comprising a sequence 30 of SEQ ID NO:53 are splice variants of each other; a polynucleotide comprising a sequence of SEQ ID NO:55 and a polynucleotide comprising a sequence of SEQ ID NO:56 are splice variants of each other; a polynucleotide comprising a sequence of SEQ ID NO:63 and a polynucleotide comprising a sequence of SEQ ID NO:64 are splice variants of each other; a polynucleotide comprising a sequence

of SEQ ID NO:66, a polynucleotide comprising a sequence of SEQ ID NO:67 and a polynucleotide comprising a sequence of SEQ ID NO:68 are splice variants of each other; and a polynucleotide comprising a sequence of SEQ ID NO:72 and a polynucleotide comprising a sequence of SEQ ID NO:73 are splice variants of each other. Any one of the splice variants described above can encode a polyneptide which contains at least one functional or structural characteristic of ENZM.

It will be appreciated by those skilled in the art that as a result of the degeneracy of the genetic code, a multitude of polynucleotide sequences encoding ENZM, some bearing minimal similarity to the polynucleotide sequences of any known and naturally occurring gene, may be produced. Thus, the invention contemplates each and every possible variation of polynucleotide sequence that could be made by selecting combinations based on possible codon choices. These combinations are made in accordance with the standard triplet genetic code as applied to the polynucleotide sequence of naturally occurring ENZM, and all such variations are to be considered as being specifically disclosed.

Although polynucleotides which encode ENZM and its variants are generally capable of
hybridizing to polynucleotides encoding naturally occurring ENZM under appropriately selected
conditions of stringency, it may be advantageous to produce polynucleotides encoding ENZM or its
derivatives possessing a substantially different codon usage, e.g., inclusion of non-naturally occurring
codons. Codons may be selected to increase the rate at which expression of the peptide occurs in a
particular prokaryotic or eukaryotic host in accordance with the frequency with which particular
codons are utilized by the host. Other reasons for substantially altering the nucleotide sequence
encoding ENZM and its derivatives without altering the encoded amino acid sequences include the
production of RNA transcripts having more desirable properties, such as a greater half-life, than
transcripts produced from the naturally occurring sequence.

The invention also encompasses production of polynucleotides which encode ENZM and

ENZM derivatives, or fragments thereof, entirely by synthetic chemistry. After production, the

synthetic polynucleotide may be inserted into any of the many available expression vectors and cell

systems using reagents well known in the art. Moreover, synthetic chemistry may be used to

introduce mutations into a polynucleotide encoding ENZM or any fragment thereof.

Embodiments of the invention can also include polynucleotides that are capable of hybridizing
to the claimed polynucleotides, and, in particular, to those having the sequences shown in SEQ ID
NO:39-76 and fragments thereof, under various conditions of stringency (Wahl, G.M. and S.L. Berger
(1987) Methods Enzymol. 152:399-407; Kimmel, A.R. (1987) Methods Enzymol. 152:507-511).
Hybridization conditions, including annealing and wash conditions, are described in "Definitions."

Methods for DNA sequencing are well known in the art and may be used to practice any of the embodiments of the invention. The methods may employ such enzymes as the Klenow fragment of DNA polymerase I, SEQUENASE (US Biochemical, Cleveland OH), Taq polymerase (Applied Biosystems), thermostable T7 polymerase (Amersham Biosciences, Piscataway NJ), or combinations of polymerases and proofreading exonucleases such as those found in the ELONGASE amplification system (Invitrogen, Carlsbad CA). Preferably, sequence preparation is automated with machines such as the MICROLAB 2200 liquid transfer system (Hamilton, Reno NV), PTC200 thermal cycler (MJ Research, Watertown MA) and ABI CATALYST 800 thermal cycler (Applied Biosystems). Sequencing is then carried out using either the ABI 373 or 377 DNA sequencing system (Applied Biosystems), the MEGABACE 1000 DNA sequencing system (Amersham Biosciences), or other systems known in the art. The resulting sequences are analyzed using a variety of algorithms which are well known in the art (Ausubel et al., supra, ch. 7; Meyers, R.A. (1995) Molecular Biology and Biotechnology, Wiley VCH, New York NY, pp. 856-853).

The nucleic acids encoding ENZM may be extended utilizing a partial nucleotide sequence 15 and employing various PCR-based methods known in the art to detect upstream sequences, such as promoters and regulatory elements. For example, one method which may be employed, restriction-site PCR, uses universal and nested primers to amplify unknown sequence from genomic DNA within a cloning vector (Sarkar, G. (1993) PCR Methods Applic. 2:318-322). Another method, inverse PCR, uses primers that extend in divergent directions to amplify unknown sequence from a circularized 20 template. The template is derived from restriction fragments comprising a known genomic locus and surrounding sequences (Triglia, T. et al. (1988) Nucleic Acids Res. 16:8186). A third method, capture PCR, involves PCR amplification of DNA fragments adjacent to known sequences in human and yeast artificial chromosome DNA (Lagerstrom, M. et al. (1991) PCR Methods Applic. 1:111-119). In this method, multiple restriction enzyme digestions and ligations may be used to insert an engineered 25 double-stranded sequence into a region of unknown sequence before performing PCR. Other methods which may be used to retrieve unknown sequences are known in the art (Parker, J.D. et al. (1991) Nucleic Acids Res. 19:3055-3060). Additionally, one may use PCR, nested primers, and PROMOTERFINDER libraries (BD Clontech, Palo Alto CA) to walk genomic DNA. This procedure avoids the need to screen libraries and is useful in finding intron/exon junctions. For all 30 PCR-based methods, primers may be designed using commercially available software, such as OLIGO 4.06 primer analysis software (National Biosciences, Plymouth MN) or another appropriate program, to be about 22 to 30 nucleotides in length, to have a GC content of about 50% or more, and to anneal to the template at temperatures of about 68°C to 72°C.

When screening for full length cDNAs, it is preferable to use libraries that have been size-selected to include larger cDNAs. In addition, random-primed libraries, which often include sequences containing the 5' regions of genes, are preferable for situations in which an oligo d(T) library does not yield a full-length cDNA. Genomic libraries may be useful for extension of sequence 5 into 5' non-transcribed regulatory regions.

Capillary electrophoresis systems which are commercially available may be used to analyze the size or confirm the nucleotide sequence of sequencing or PCR products. In particular, capillary sequencing may employ flowable polymers for electrophoretic separation, four different nucleotidespecific, laser-stimulated fluorescent dyes, and a charge coupled device camera for detection of the 10 emitted wavelengths. Output/light intensity may be converted to electrical signal using appropriate software (e.g., GENOTYPER and SEQUENCE NAVIGATOR, Applied Biosystems), and the entire process from loading of samples to computer analysis and electronic data display may be computer controlled. Capillary electrophoresis is especially preferable for sequencing small DNA fragments which may be present in limited amounts in a particular sample.

In another embodiment of the invention, polynucleotides or fragments thereof which encode ENZM may be cloned in recombinant DNA molecules that direct expression of ENZM, or fragments or functional equivalents thereof, in appropriate host cells. Due to the inherent degeneracy of the genetic code, other polynucleotides which encode substantially the same or a functionally equivalent polypeptides may be produced and used to express ENZM.

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The polynucleotides of the invention can be engineered using methods generally known in the art in order to alter ENZM-encoding sequences for a variety of purposes including, but not limited to, modification of the cloning, processing, and/or expression of the gene product. DNA shuffling by random fragmentation and PCR reassembly of gene fragments and synthetic oligonucleotides may be used to engineer the nucleotide sequences. For example, oligonucleotide-mediated site-directed 25 mutagenesis may be used to introduce mutations that create new restriction sites, alter glycosylation natterns, change codon preference, produce splice variants, and so forth.

The nucleotides of the present invention may be subjected to DNA shuffling techniques such as MOLECULARBREEDING (Maxygen Inc., Santa Clara CA; described in U.S. Patent No. 5,837,458; Chang, C.-C. et al. (1999) Nat. Biotechnol. 17:793-797; Christians, F.C. et al. (1999) Nat. 30 Biotechnol. 17:259-264; and Crameri, A. et al. (1996) Nat. Biotechnol. 14:315-319) to alter or improve the biological properties of ENZM, such as its biological or enzymatic activity or its ability to bind to other molecules or compounds. DNA shuffling is a process by which a library of gene variants is produced using PCR-mediated recombination of gene fragments. The library is then subjected to

selection or screening procedures that identify those gene variants with the desired properties. These preferred variants may then be pooled and further subjected to recursive rounds of DNA shuffling and selection/screening. Thus, genetic diversity is created through "artificial" breeding and rapid molecular evolution. For example, fragments of a single gene containing random point mutations may be recombined, screened, and then reshuffled until the desired properties are optimized. Alternatively, fragments of a given gene may be recombined with fragments of homologous genes in the same gene family, either from the same or different species, thereby maximizing the genetic diversity of multiple naturally occurring genes in a directed and controllable manner.

In another embodiment, polynucleotides encoding ENZM may be synthesized, in whole or in

part, using one or more chemical methods well known in the art (Caruthers, M.H. et al. (1980)

Nucleic Acids Symp. Ser. 7:215-223; Horn, T. et al. (1980) Nucleic Acids Symp. Ser. 7:225-232).

Alternatively, ENZM itself or a fragment thereof may be synthesized using chemical methods known in the art. For example, peptide synthesis can be performed using various solution-phase or solid-phase techniques (Creighton, T. (1984) Proteins, Structures and Molecular Properties, WH

Freeman, New York NY, pp. 55-60; Roberge, J.Y. et al. (1995) Science 269:202-204). Automated synthesis may be achieved using the ABI 431A peptide synthesizer (Applied Biosystems).

Additionally, the amino acid sequence of ENZM, or any part thereof, may be altered during direct synthesis and/or combined with sequences from other proteins, or any part thereof, to produce a variant polypeptide or a polypeptide having a sequence of a naturally occurring polypeptide.

The peptide may be substantially purified by preparative high performance liquid chromatography (Chiez, R.M. and F.Z. Regnier (1990) Methods Enzymol. 182:392-421). The composition of the synthetic peptides may be confirmed by amino acid analysis or by sequencing (Creighton, supra, pp. 28-53).

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In order to express a biologically active ENZM, the polynucleotides encoding ENZM or
derivatives thereof may be inserted into an appropriate expression vector, i.e., a vector which contains
the necessary elements for transcriptional and translational control of the inserted coding sequence in
a suitable host. These elements include regulatory sequences, such as enhancers, constitutive and
inducible promoters, and 5' and 3' untranslated regions in the vector and in polynucleotides encoding
ENZM. Such elements may vary in their strength and specificity. Specific initiation signals may also
be used to achieve more efficient translation of polynucleotides encoding ENZM. Such signals include
the ATG initiation codon and adjacent sequences, e.g. the Kozak sequence. In cases where a
polynucleotide sequence encoding ENZM and its initiation codon and upstream regulatory sequences
are inserted into the appropriate expression vector, no additional transcriptional or translational control

signals may be needed. However, in cases where only coding sequence, or a fragment thereof, is inserted, exogenous translational control signals including an in-frame ATG initiation codon should be provided by the vector. Exogenous translational elements and initiation codons may be of various origins, both natural and synthetic. The efficiency of expression may be enhanced by the inclusion of enhancers appropriate for the particular host cell system used (Scharf, D. et al. (1994) Results Probl. Cell Differ. 20:125-162).

Methods which are well known to those skilled in the art may be used to construct expression vectors containing polynucleotides encoding ENZM and appropriate transcriptional and translational control elements. These methods include in vitro recombinant DNA techniques, synthetic techniques, and in vivo genetic recombination (Sambrook and Russell, supra, ch. 1-4, and 8; Ausubel et al., supra, ch. 1, 3, and 15).

A variety of expression vector/host systems may be utilized to contain and express polynucleotides encoding ENZM. These include, but are not limited to, microorganisms such as bacteria transformed with recombinant bacteriophage, plasmid, or cosmid DNA expression vectors; 15 yeast transformed with yeast expression vectors; insect cell systems infected with viral expression vectors (e.g., baculovirus); plant cell systems transformed with viral expression vectors (e.g., cauliflower mosaic virus, CaMV, or tobacco mosaic virus, TMV) or with bacterial expression vectors (e.g., Ti or pBR322 plasmids); or animal cell systems (Sambrook and Russell, supra; Ausubel et al., supra; Van Heeke, G. and S.M. Schuster (1989) J. Biol. Chem. 264:5503-5509; Engelhard, E.K. et al. 20 (1994) Proc. Natl. Acad. Sci. USA 91:3224-3227; Sandig, V. et al. (1996) Hum. Gene Ther. 7:1937-1945: Takamatsu, N. (1987) EMBO J. 6:307-311: The McGraw Hill Yearbook of Science and Technology (1992) McGraw Hill, New York NY, pp. 191-196; Logan, J. and T. Shenk (1984) Proc. Natl. Acad. Sci. USA 81:3655-3659; Harrington, J.J. et al. (1997) Nat. Genet. 15:345-355). Expression vectors derived from retroviruses, adenoviruses, or herpes or vaccinia viruses, or from 25 various bacterial plasmids, may be used for delivery of polynucleotides to the targeted organ, tissue, or cell population (Di Nicola, M. et al. (1998) Cancer Gen. Ther. 5:350-356; Yu, M. et al. (1993) Proc. Natl. Acad. Sci. USA 90:6340-6344; Buller, R.M. et al. (1985) Nature 317:813-815; McGregor, D.P. et al. (1994) Mol. Immunol. 31:219-226; Verma, I.M. and N. Somia (1997) Nature 389:239-242). The invention is not limited by the host cell employed.

In bacterial systems, a number of cloning and expression vectors may be selected depending upon the use intended for polynucleotides encoding ENZM. For example, routine cloning, subcloning, and propagation of polynucleotides encoding ENZM can be achieved using a multifunctional E. coli vector such as PBLUESCRIPT (Stratagene, La Jolla CA) or PSPORT1 plasmid (Invitrogen).

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Ligation of polynucleotides encoding ENZM into the vector's multiple cloning site disrupts the lacZ gene, allowing a colorimetric screening procedure for identification of transformed bacteria containing recombinant molecules. In addition, these vectors may be useful for in vitro transcription, dideoxy sequencing, single strand rescue with helper phage, and creation of nested deletions in the cloned sequence (Van Heeke, G. and S.M. Schuster (1989) J. Biol. Chem. 264:5503-5509). When large quantities of ENZM are needed, e.g. for the production of antibodies, vectors which direct high level expression of ENZM may be used. For example, vectors containing the strong, inducible SP6 or T7 bacteriophage promoter may be used.

Yeast expression systems may be used for production of ENZM. A number of vectors

containing constitutive or inducible promoters, such as alpha factor, alcohol oxidase, and PGH promoters, may be used in the yeast Saccharomyces cerevisiae or Pichia pastoris. In addition, such vectors direct either the secretion or intracellular retention of expressed proteins and enable integration of foreign polynucleotide sequences into the host genome for stable propagation (Ausubel et al., supra; Bitter, G.A. et al. (1987) Methods Enzymol. 153:516-544; Scorer, C.A. et al. (1994)

Bio/Technology 12:181-184).

Plant systems may also be used for expression of ENZM. Transcription of polynucleotides encoding ENZM may be driven by viral promoters, e.g., the 35S and 198 promoters of CaMV used alone or in combination with the omega leader sequence from TMV (Takamatsu, N. (1987) EMBO J. 6:307-311). Alternatively, plant promoters such as the small subunit of RUBISCO or heat shock promoters may be used (Coruzzi, G. et al. (1984) EMBO J. 3:1671-1680; Broglie, R. et al. (1984) Science 224:838-843; Winter, J. et al. (1991) Results Probl. Cell Differ. 17:85-105). These constructs can be introduced into plant cells by direct DNA transformation or pathogen-mediated transfection (The McGraw Hill Yearbook of Science and Technology (1992) McGraw Hill, New York NY, pp. 191-196).

In mammalian cells, a number of viral-based expression systems may be utilized. In cases where an adenovirus is used as an expression vector, polynucleotides encoding ENZM may be ligated into an adenovirus transcription/translation complex consisting of the late promoter and tripartite leader sequence. Insertion in a non-essential E1 or E3 region of the viral genome may be used to obtain infective virus which expresses ENZM in host cells (Logan, J. and T. Shenk (1984) Proc. Natl. Acad. Sci. USA 81:3655-3659). In addition, transcription enhancers, such as the Rous sarcoma virus (RSV) enhancer, may be used to increase expression in mammalian host cells. SV40 or EBV-based vectors may also be used for high-level protein expression.

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Human artificial chromosomes (HACs) may also be employed to deliver larger fragments of

DNA than can be contained in and expressed from a plasmid. HACs of about 6 kb to 10 Mb are constructed and delivered via conventional delivery methods (liposomes, polycationic amino polymers, or vesicles) for therapeutic purposes (Harrington, J.J. et al. (1997) Nat. Genet. 15:345-355).

For long term production of recombinant proteins in mammalian systems, stable expression of

ENZM in cell lines is preferred. For example, polynucleotides encoding ENZM can be transformed
into cell lines using expression vectors which may contain viral origins of replication and/or
endogenous expression elements and a selectable marker gene on the same or on a separate vector.
Following the introduction of the vector, cells may be allowed to grow for about 1 to 2 days in enriched
media before being switched to selective media. The purpose of the selectable marker is to confer

resistance to a selective agent, and its presence allows growth and recovery of cells which
successfully express the introduced sequences. Resistant clones of stably transformed cells may be
propagated using tissue culture techniques appropriate to the cell type.

Any number of selection systems may be used to recover transformed cell lines. These include, but are not limited to, the herpes simplex virus thymidine kinase and adenine

15 phosphoribosyltransferase genes, for use in it and apr cells, respectively (Wigler, M. et al. (1977)

Cell 11:223-232; Lowy, I. et al. (1980) Cell 22:817-823). Also, antimetabolite, antibiotic, or herbicide resistance can be used as the basis for selection. For example, dhfr confers resistance to methotrexate; neo confers resistance to the aminoglycosides neomycin and G-418; and als and pat confer resistance to chlorsulfuron and phosphinotricin acetyltransferase, respectively (Wigler, M. et al. (1980) Proc. Natl. Acad. Sci. USA 77:3567-3570; Colbere-Garapin, F. et al. (1981) J. Mol. Biol. 150:1-14). Additional selectable genes have been described, e.g., trpB and hisD, which alter cellular requirements for metabolites (Hartman, S.C. and R.C. Mulligan (1988) Proc. Natl. Acad. Sci. USA 85:8047-8051). Visible markers, e.g., anthocyanins, green fluorescent proteins (GFP; BD Clontech), β-glucuronidase and its substrate β-glucuronide, or luciferase and its substrate luciferin may be used.

These markers can be used not only to identify transformants, but also to quantify the amount of transient or stable protein expression attributable to a specific vector system (Rhodes, C.A. (1995) Methods Mol. Biol. 55:121-131).

Although the presence/absence of marker gene expression suggests that the gene of interest is also present, the presence and expression of the gene may need to be confirmed. For example, if the sequence encoding ENZM is inserted within a marker gene sequence, transformed cells containing polynucleotides encoding ENZM can be identified by the absence of marker gene function.

Alternatively, a marker gene can be placed in tandem with a sequence encoding ENZM under the control of a single promoter. Expression of the marker gene in response to induction or selection

usually indicates expression of the tandem gene as well.

In general, host cells that contain the polynucleotide encoding ENZM and that express ENZM may be identified by a variety of procedures known to those of skill in the art. These procedures include, but are not limited to, DNA-DNA or DNA-RNA hybridizations, PCR amplification, and protein bioassay or immunoassay techniques which include membrane, solution, or chip based technologies for the detection and/or quantification of nucleic acid or protein sequences.

Immunological methods for detecting and measuring the expression of ENZM using either specific polyclonal or monoclonal antibodies are known in the art. Examples of such techniques include enzyme-linked immunosorbent assays (ELISAs), radioimmunoassays (RIAs), and of fluorescence activated cell sorting (FACS). A two-site, monoclonal-based immunoassay utilizing monoclonal antibodies reactive to two non-interfering epitopes on ENZM is preferred, but a competitive binding assay may be employed. These and other assays are well known in the art (Hampton, R. et al. (1990) Serological Methods, a Laboratory Manual. APS Press, St. Paul MN, Sect. IV; Coligan, J.E. et al. (1997) Current Protocols in Immunology. Greene Pub. Associates and Wiley-Interscience, New York NY; Pound, J.D. (1998) Immunochemical Protocols. Humana Press, Totowa ND.

A wide variety of labels and conjugation techniques are known by those skilled in the art and may be used in various nucleic acid and amino acid assays. Means for producing labeled hybridization or PCR probes for detecting sequences related to polynucleotides encoding ENZM include considerable in the constant of the

Host cells transformed with polynucleotides encoding ENZM may be cultured under

conditions suitable for the expression and recovery of the protein from cell culture. The protein

produced by a transformed cell may be secreted or retained intracellularly depending on the sequence

and/or the vector used. As will be understood by those of skill in the art, expression vectors containing

polynucleotides which encode ENZM may be designed to contain signal sequences which direct

secretion of ENZM through a prokaryotic or eukaryotic cell membrane.

In addition, a host cell strain may be chosen for its ability to modulate expression of the inserted polynucleotides or to process the expressed protein in the desired fashion. Such modifications of the polypeptide include, but are not limited to, acetylation, carboxylation, glycosylation, 5 phosphorylation, lipidation, and acylation. Post-translational processing which cleaves a "prepro" or "pro" form of the protein may also be used to specify protein targeting, folding, and/or activity. Different host cells which have specific cellular machinery and characteristic mechanisms for post-translational activities (e.g., CHO, HeLa, MDCK, HEK293, and WI38) are available from the American Type Culture Collection (ATCC, Manassas VA) and may be chosen to ensure the correct modification and processing of the foreign protein.

In another embodiment of the invention, natural, modified, or recombinant polynucleotides encoding ENZM may be ligated to a heterologous sequence resulting in translation of a fusion protein in any of the aforementioned host systems. For example, a chimeric ENZM protein containing a heterologous moiety that can be recognized by a commercially available antibody may facilitate the 15 screening of peptide libraries for inhibitors of ENZM activity. Heterologous protein and peptide moieties may also facilitate purification of fusion proteins using commercially available affinity matrices. Such moieties include, but are not limited to, glutathione S-transferase (GST), maltose binding protein (MBP), thioredoxin (Trx), calmodulin binding peptide (CBP), 6-His, FLAG, c-myc, and hemagglutinin (HA). GST, MBP, Trx, CBP, and 6-His enable purification of their cognate fusion 20 proteins on immobilized glutathione, maltose, phenylarsine oxide, calmodulin, and metal-chelate resins, respectively. FLAG, c-myc, and hemagglutinin (HA) enable immunoaffinity purification of fusion proteins using commercially available monoclonal and polyclonal antibodies that specifically recognize these epitope tags. A fusion protein may also be engineered to contain a proteolytic cleavage site located between the ENZM encoding sequence and the heterologous protein sequence, so that ENZM 25 may be cleaved away from the heterologous moiety following purification. Methods for fusion protein expression and purification are discussed in Ausubel et al. (supra, ch. 10 and 16). A variety of commercially available kits may also be used to facilitate expression and purification of fusion proteins.

In another embodiment, synthesis of radiolabeled ENZM may be achieved in vitro using the TNT rabbit reticulocyte lysate or wheat germ extract system (Promega). These systems couple transcription and translation of protein-coding sequences operably associated with the T7, T3, or SP6 promoters. Translation takes place in the presence of a radiolabeled amino acid precursor, for example, <sup>35</sup>S-methionine.

ENZM, fragments of ENZM, or variants of ENZM may be used to screen for compounds

that specifically bind to ENZM. One or more test compounds may be screened for specific binding to ENZM. In various embodiments, 1, 2, 3, 4, 5, 10, 20, 50, 100, or 200 test compounds can be screened for specific binding to ENZM. Examples of test compounds can include antibodies, anticalins, oligonucleotides, proteins (e.g., ligands or receptors), or small molecules.

In related embodiments, variants of ENZM can be used to screen for binding of test compounds, such as antibodies, to ENZM, a variant of ENZM, or a combination of ENZM and/or one or more variants ENZM. In an embodiment, a variant of ENZM can be used to screen for compounds that bind to a variant of ENZM, but not to ENZM having the exact sequence of a sequence of SEQ ID NO:1-38. ENZM variants used to perform such screening can have a range of about 50% to about 99% sequence identity to ENZM, with various embodiments having 60%, 70%, 75%, 80%, 85%, 90%, and 95% sequence identity.

In an embodiment, a compound identified in a screen for specific binding to ENZM can be closely related to the natural ligand of ENZM, e.g., a ligand or fragment thereof, a natural substrate, a structural or functional mimetic, or a natural binding partner (Coligan, J.E. et al. (1991) Current

15 Protocols in Immunology 1(2):Chapter 5). In another embodiment, the compound thus identified can be a natural ligand of a receptor ENZM (Howard, A.D. et al. (2001) Trends Pharmacol. Sci.22:132-140; Wise, A. et al. (2002) Drug Discovery Today 7:235-246).

In other embodiments, a compound identified in a screen for specific binding to ENZM can be closely related to the natural receptor to which ENZM binds, at least a fragment of the receptor, or a 20 fragment of the receptor including all or a portion of the ligand binding site or binding pocket. For example, the compound may be a receptor for ENZM which is capable of propagating a signal, or a decoy receptor for ENZM which is not capable of propagating a signal (Ashkenazi, A. and V.M. Divit (1999) Curr. Opin. Cell Biol. 11:255-260; Mantovani, A. et al. (2001) Trends Immunol. 22:328-336). The compound can be rationally designed using known techniques. Examples of such techniques include those used to construct the compound etanercept (ENBREL; Amgen Inc., Thousand Oaks CA), which is efficacious for treating rheumatoid arthritis in humans. Etanercept is an engineered p75 tumor necrosis factor (TNF) receptor dimer linked to the Fe portion of human IgG<sub>1</sub> (Taylor, P.C. et al. (2001) Curr. Opin. Immunol. 13:611-616).

In one embodiment, two or more antibodies having similar or, alternatively, different
specificities can be screened for specific binding to ENZM, fragments of ENZM, or variants of
ENZM. The binding specificity of the antibodies thus screened can thereby be selected to identify
particular fragments or variants of ENZM. In one embodiment, an antibody can be selected such that
its binding specificity allows for preferential identification of specific fragments or variants of ENZM.

In another embodiment, an antibody can be selected such that its binding specificity allows for preferential diagnosis of a specific disease or condition having increased, decreased, or otherwise abnormal production of ENZM.

In an embodiment, anticalins can be screened for specific binding to ENZM, fragments of 5 ENZM, or variants of ENZM. Anticalins are ligand-binding proteins that have been constructed based on a lipocalin scaffold (Weiss, G.A. and H.B. Lowman (2000) Chem. Biol. 7:R177-R184; Skerra, A. (2001) J. Biotechnol. 74:257-275). The protein architecture of lipocalins can include a beta-barrel having eight antiparallel beta-strands, which supports four loops at its open end. These loops form the natural ligand-binding site of the lipocalins, a site which can be re-engineered in vitro by amino acid 10 substitutions to impart novel binding specificities. The amino acid substitutions can be made using methods known in the art or described herein, and can include conservative substitutions (e.g., substitutions that do not alter binding specificity) or substitutions that modestly, moderately, or significantly alter binding specificity.

In one embodiment, screening for compounds which specifically bind to, stimulate, or inhibit 15 ENZM involves producing appropriate cells which express ENZM, either as a secreted protein or on the cell membrane. Preferred cells can include cells from mammals, yeast, Drosophila, or E. coli. Cells expressing ENZM or cell membrane fractions which contain ENZM are then contacted with a test compound and binding, stimulation, or inhibition of activity of either ENZM or the compound is analyzed.

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An assay may simply test binding of a test compound to the polypeptide, wherein binding is detected by a fluorophore, radioisotope, enzyme conjugate, or other detectable label. For example, the assay may comprise the steps of combining at least one test compound with ENZM, either in solution or affixed to a solid support, and detecting the binding of ENZM to the compound. Alternatively, the assay may detect or measure binding of a test compound in the presence of a labeled competitor. 25 Additionally, the assay may be carried out using cell-free preparations, chemical libraries, or natural product mixtures, and the test compound(s) may be free in solution or affixed to a solid support.

An assay can be used to assess the ability of a compound to bind to its natural ligand and/or to inhibit the binding of its natural ligand to its natural receptors. Examples of such assays include radiolabeling assays such as those described in U.S. Patent No. 5,914,236 and U.S. Patent No. 6,372,724. 30 In a related embodiment, one or more amino acid substitutions can be introduced into a polypeptide compound (such as a receptor) to improve or alter its ability to bind to its natural ligands (Matthews, D.J. and J.A. Wells. (1994) Chem. Biol. 1:25-30). In another related embodiment, one or more amino acid substitutions can be introduced into a polypeptide compound (such as a ligand) to improve or alter

its ability to bind to its natural receptors (Cunningham, B.C. and J.A. Wells (1991) Proc. Natl. Acad. Sci. USA 88:3407-3411; Lowman, H.B. et al. (1991) J. Biol. Chem. 266:10982-10988).

ENZM, fragments of ENZM, or variants of ENZM may be used to screen for compounds that modulate the activity of ENZM. Such compounds may include agonists, antagonists, or partial or 5 inverse agonists. In one embodiment, an assay is performed under conditions permissive for ENZM activity, wherein ENZM is combined with at least one test compound, and the activity of ENZM in the presence of a test compound is compared with the activity of ENZM in the absence of the test compound. A change in the activity of ENZM in the presence of the test compound is indicative of a compound that modulates the activity of ENZM. Alternatively, a test compound is combined with an 10 in vitro or cell-free system comprising ENZM under conditions suitable for ENZM activity, and the assay is performed. In either of these assays, a test compound which modulates the activity of ENZM may do so indirectly and need not come in direct contact with the test compound. At least one and up to a plurality of test compounds may be screened.

In another embodiment, polynucleotides encoding ENZM or their mammalian homologs may

be "knocked out" in an animal model system using homologous recombination in embryonic stem (ES)

cells. Such techniques are well known in the art and are useful for the generation of animal models of
human disease (see, e.g., U.S. Patent No. 5,175,383 and U.S. Patent No. 5,767,337). For example,
mouse ES cells, such as the mouse 129/SvJ cell line, are derived from the early mouse embryo and
grown in culture. The ES cells are transformed with a vector containing the gene of interest disrupted

20 by a marker gene, e.g., the neomycin phosphotransferase gene (neo; Capecchi, M.R. (1989) Science

244:1288-1292). The vector integrates into the corresponding region of the host genome by
homologous recombination. Alternatively, homologous recombination takes place using the Cre-loxP
system to knockout a gene of interest in a tissue- or developmental stage-specific manner (Marth, J.D.
(1996) Clin. Invest. 97:1999-2002; Wagner, K.U. et al. (1997) Nucleic Acids Res. 25:4323-4330).

Transformed ES cells are identified and microinjected into mouse cell blastocysts such as those from
the C57BL/6 mouse strain. The blastocysts are surgically transferred to pseudopregnant dams, and
the resulting chimeric progeny are genotyped and bred to produce heterozygous or homozygous
strains. Transgenic animals thus generated may be tested with potential therapeutic or toxic agents.

Polynucleotides encoding ENZM may also be manipulated in vitro in ES cells derived from human blastocysts. Human ES cells have the potential to differentiate into at least eight separate cell lineages including endoderm, mesoderm, and ectodermal cell types. These cell lineages differentiate into, for example, neural cells, hematopoietic lineages, and cardiomyocytes (Thomson, J.A. et al. (1998) Science 282:1145-1147).

Polynucleotides encoding ENZM can also be used to create "knockin" humanized animals (pigs) or transgenic animals (mice or rats) to model human disease. With knockin technology, a region of a polynucleotide encoding ENZM is injected into animal ES cells, and the injected sequence integrates into the animal cell genome. Transformed cells are injected into blastulae, and the blastulae 5 are implanted as described above. Transgenic progeny or inbred lines are studied and treated with potential pharmaceutical agents to obtain information on treatment of a human disease. Alternatively, a mammal inbred to overexpress ENZM, e.g., by secreting ENZM in its milk, may also serve as a convenient source of that protein (Janne, J. et al. (1998) Biotechnol. Annu. Rev. 4:55-74).

## THERAPEUTICS

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Chemical and structural similarity, e.g., in the context of sequences and motifs, exists between regions of ENZM and enzymes. In addition, examples of tissues expressing ENZM can be found in Table 6 and can also be found in Example XI. Therefore, ENZM appears to play a role in autoimmune/inflammatory disorders, infectious disorders, immune deficiencies, disorders of metabolism, reproductive disorders, neurological disorders, cardiovascular disorders, eye disorders, and 15 cell proliferative disorders, including cancer. In the treatment of disorders associated with increased ENZM expression or activity, it is desirable to decrease the expression or activity of ENZM. In the treatment of disorders associated with decreased ENZM expression or activity, it is desirable to increase the expression or activity of ENZM.

Therefore, in one embodiment, ENZM or a fragment or derivative thereof may be administered to a subject to treat or prevent a disorder associated with decreased expression or activity of ENZM. Examples of such disorders include, but are not limited to, an autoimmune/inflammatory disorder such as acquired immunodeficiency syndrome (AIDS), Addison's disease, adult respiratory distress syndrome, allergies, ankylosing spondylitis, amyloidosis, anemia, asthma, atherosclerosis, autoimmune hemolytic anemia, autoimmune thyroiditis, autoimmune 25 polyendocrinopathy-candidiasis-ectodermal dystrophy (APECED), bronchitis, cholecystitis, contact dermatitis, Crohn's disease, atopic dermatitis, dermatomyositis, diabetes mellitus, emphysema, episodic lymphopenia with lymphocytotoxins, erythroblastosis fetalis, erythema nodosum, atrophic gastritis, glomerulonephritis, Goodpasture's syndrome, gout, Graves' disease, Hashimoto's thyroiditis, hypereosinophilia, irritable bowel syndrome, multiple sclerosis, myasthenia gravis, myocardial or 30 pericardial inflammation, osteoarthritis, osteoporosis, pancreatitis, polymyositis, psoriasis, Reiter's syndrome, rheumatoid arthritis, scleroderma, Sjögren's syndrome, systemic anaphylaxis, systemic lupus erythematosus, systemic sclerosis, thrombocytopenic purpura, ulcerative colitis, uveitis, Werner syndrome, complications of cancer, hemodialysis, and extracorporeal circulation, and trauma; an

infectious disorder such as a viral infection, e.g., caused by an adenovirus (acute respiratory disease, pneumonia), an arenavirus (lymphocytic choriomeningitis), a bunyavirus (Hantavirus), a coronavirus (pneumonia, chronic bronchitis), a hepadnavirus (hepatitis), a herpesvirus (herpes simplex virus, varicella-zoster virus, Epstein-Barr virus, cytomegalovirus), a flavivirus (yellow fever), an 5 orthomyxovirus (influenza), a papillomavirus (cancer), a paramyxovirus (measles, mumps), a picornovirus (rhinovirus, poliovirus, coxsackie-virus), a polyomavirus (BK virus, JC virus), a poxvirus (smallpox), a reovirus (Colorado tick fever), a retrovirus (human immunodeficiency virus, human T lymphotropic virus), a rhabdovirus (rabies), a rotavirus (gastroenteritis), and a togavirus (encephalitis, rubella), and a bacterial infection, a fungal infection, a parasitic infection, a protozoal infection, and a 10 helminthic infection; an immune deficiency, such as acquired immunodeficiency syndrome (AIDS), Xlinked agammaglobinemia of Bruton, common variable immunodeficiency (CVI), DiGeorge's syndrome (thymic hypoplasia), thymic dysplasia, isolated IgA deficiency, severe combined immunodeficiency disease (SCID), immunodeficiency with thrombocytopenia and eczema (Wiskott-Aldrich syndrome), Chediak-Higashi syndrome, chronic granulomatous diseases, hereditary 15 angioneurotic edema, and immunodeficiency associated with Cushing's disease; a disorder of metabolism such as Addison's disease, cerebrotendinous xanthomatosis, congenital adrenal hyperplasia, coumarin resistance, cystic fibrosis, diabetes, fatty hepatocirrhosis, fructose-1,6-diphosphatase deficiency, galactosemia, goiter, glucagonoma, glycogen storage diseases, hereditary fructose intolerance, hyperadrenalism, hypoadrenalism, hyperparathyroidism, 20 hypoparathyroidism, hypercholesterolemia, hyperthyroidism, hypoglycemia, hypothyroidism, hyperlipidemia, hyperlipemia, a lipid myopathy, a lipodystrophy, a lysosomal storage disease, mannosidosis, neuraminidase deficiency, obesity, pentosuria phenylketonuria, pseudovitamin Ddeficiency rickets; a reproductive disorder such as a disorder of prolactin production, infertility, including tubal disease, ovulatory defects, and endometriosis, a disruption of the estrous cycle, a 25 disruption of the menstrual cycle, polycystic ovary syndrome, ovarian hyperstimulation syndrome, endometrial and ovarian tumors, uterine fibroids, autoimmune disorders, ectopic pregnancies, and teratogenesis, cancer of the breast, fibrocystic breast disease, and galactorrhea, disruptions of spermatogenesis, abnormal sperm physiology, cancer of the testis, cancer of the prostate, benign prostatic hyperplasia, prostatitis, Peyronie's disease, impotence, carcinoma of the male breast, and gynecomastia; a neurological disorder such as epilepsy, ischemic cerebrovascular disease, stroke, cerebral neoplasms, Alzheimer's disease, Pick's disease, Huntington's disease, dementia, Parkinson's disease and other extrapyramidal disorders, amyotrophic lateral sclerosis and other motor neuron disorders, progressive neural muscular atrophy, retinitis pigmentosa, hereditary ataxias, multiple

sclerosis and other demyelinating diseases, bacterial and viral meningitis, brain abscess, subdural empyema, epidural abscess, suppurative intracranial thrombophlebitis, myelitis and radiculitis, viral central nervous system disease; prion diseases including kuru, Creutzfeldt-Jakob disease, and Gerstmann-Straussler-Scheinker syndrome; fatal familial insomnia, nutritional and metabolic diseases 5 of the nervous system, neurofibromatosis, tuberous sclerosis, cerebelloretinal hemangioblastomatosis, encephalotrigeminal syndrome, mental retardation and other developmental disorders of the central nervous system, cerebral palsy, neuroskeletal disorders, autonomic nervous system disorders, cranial nerve disorders, spinal cord diseases, muscular dystrophy and other neuromuscular disorders, peripheral nervous system disorders, dermatomyositis and polymyositis; inherited, metabolic, endocrine, and toxic myopathies; myasthenia gravis, periodic paralysis; mental disorders including mood, anxiety, and schizophrenic disorders; seasonal affective disorder (SAD); akathesia, amnesia, catatonia, diabetic neuropathy, tardive dyskinesia, dystonias, paranoid psychoses, postherpetic neuralgia, and Tourette's disorder; a cardiovascular disorder, such as arteriovenous fistula, atherosclerosis, hypertension, vasculitis, Raynaud's disease, aneurysms, arterial dissections, varicose veins, thrombophlebitis and 15 phlebothrombosis, vascular tumors, and complications of thrombolysis, balloon angioplasty, vascular replacement, and coronary artery bypass graft surgery, congestive heart failure, ischemic heart disease, angina pectoris, myocardial infarction, hypertensive heart disease, degenerative valvular heart disease, calcific aortic valve stenosis, congenitally bicuspid aortic valve, mitral annular calcification, mitral valve prolapse, rheumatic fever and rheumatic heart disease, infective endocarditis, nonbacterial 20 thrombotic endocarditis, endocarditis of systemic lupus erythematosus, carcinoid heart disease, cardiomyopathy, myocarditis, pericarditis, neoplastic heart disease, congenital heart disease, and complications of cardiac transplantation, congenital lung anomalies, atelectasis, pulmonary congestion and edema, pulmonary embolism, pulmonary hemorrhage, pulmonary infarction, pulmonary hypertension, vascular sclerosis, obstructive pulmonary disease, restrictive pulmonary disease, chronic 25 obstructive pulmonary disease, emphysema, chronic bronchitis, bronchial asthma, bronchiectasis, bacterial pneumonia, viral and mycoplasmal pneumonia, lung abscess, pulmonary tuberculosis, diffuse interstitial diseases, pneumoconioses, sarcoidosis, idiopathic pulmonary fibrosis, desquamative interstitial pneumonitis, hypersensitivity pneumonitis, pulmonary eosinophilia bronchiolitis obliterans-organizing pneumonia, diffuse pulmonary hemorrhage syndromes, Goodpasture's syndromes, 30 idiopathic pulmonary hemosiderosis, pulmonary involvement in collagen-vascular disorders, pulmonary alveolar proteinosis, lung tumors, inflammatory and noninflammatory pleural effusions, pneumothorax, pleural tumors, drug-induced lung disease, radiation-induced lung disease, and complications of lung transplantation; an eye disorder such as ocular hypertension and glaucoma; a disorder of cell

proliferation such as actinic keratosis, arteriosclerosis, atherosclerosis, bursitis, cirrhosis, hepatitis, mixed connective tissue disease (MCTD), myelofibrosis, paroxysmal nocturnal hemoglobinuria, polycythemia vera, psoriasis, primary thrombocythemia; and a cancer, including adenocarcinoma, leukemia, lymphoma, melanoma, myeloma, sarcoma, teratocarcinoma, and, in particular, cancers of the adrenal gland, bladder, bone, bone marrow, brain, breast, cervix, colon, gall bladder, ganglia, gastrointestinal tract, heart, kidney, liver, lung, muscle, ovary, pancreas, parathyroid, penis, prostate, salivary glands, skin, spleen, testis, thymus, thyroid, and uterus.

In another embodiment, a vector capable of expressing ENZM or a fragment or derivative thereof may be administered to a subject to treat or prevent a disorder associated with decreased expression or activity of ENZM including, but not limited to, those described above.

In a further embodiment, a composition comprising a substantially purified ENZM in conjunction with a suitable pharmaceutical carrier may be administered to a subject to treat or prevent a disorder associated with decreased expression or activity of ENZM including, but not limited to, those provided above.

In still another embodiment, an agonist which modulates the activity of ENZM may be administered to a subject to treat or prevent a disorder associated with decreased expression or activity of ENZM including, but not limited to, those listed above.

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In a further embodiment, an antagonist of ENZM may be administered to a subject to treat or prevent a disorder associated with increased expression or activity of ENZM. Examples of such disorders include, but are not limited to, those autoimmune/inflammatory disorders, infectious disorders, immune deficiencies, disorders of metabolism, reproductive disorders, neurological disorders, cardiovascular disorders, eye disorders, and cell proliferative disorders, including cancer described above. In one aspect, an antibody which specifically binds ENZM may be used directly as an antagonist or indirectly as a targeting or delivery mechanism for bringing a pharmaceutical agent to cells or tissues which express ENZM.

In an additional embodiment, a vector expressing the complement of the polynucleotide encoding ENZM may be administered to a subject to treat or prevent a disorder associated with increased expression or activity of ENZM including, but not limited to, those described above.

In other embodiments, any protein, agonist, antagonist, antibody, complementary sequence, or
vector embodiments may be administered in combination with other appropriate therapeutic agents.

Selection of the appropriate agents for use in combination therapy may be made by one of ordinary
skill in the art, according to conventional pharmaceutical principles. The combination of therapeutic
agents may act synergistically to effect the treatment or prevention of the various disorders described

above. Using this approach, one may be able to achieve therapeutic efficacy with lower dosages of each agent, thus reducing the potential for adverse side effects.

An antagonist of ENZM may be produced using methods which are generally known in the art. In particular, purified ENZM may be used to produce antibodies or to screen libraries of

5 pharmaceutical agents to identify those which specifically bind ENZM. Antibodies to ENZM may also be generated using methods that are well known in the art. Such antibodies may include, but are not limited to, polyclonal, monoclonal, chimeric, and single chain antibodies, Fab fragments, and fragments produced by a Fab expression library. In an embodiment, neutralizing antibodies (i.e., those which inhibit dimer formation) can be used therapeutically. Single chain antibodies (e.g., from camels or llamas) may be potent enzyme inhibitors and may have application in the design of peptide mimetics, and in the development of immuno-adsorbents and biosensors (Muyldermans, S. (2001) J. Biotechnol. 74:277-302).

For the production of antibodies, various hosts including goats, rabbits, rats, mice, camels, dromedaries, llamas, humans, and others may be immunized by injection with ENZM or with any

15 fragment or oligopeptide thereof which has immunogenic properties. Depending on the host species, various adjuvants may be used to increase immunological response. Such adjuvants include, but are not limited to, Freund's, mineral gels such as aluminum hydroxide, and surface active substances such as lysolecithin, pluronic polyols, polyanions, peptides, oil emulsions, KLH, and dinitrophenol. Among adjuvants used in humans, BCG (bacilli Calmette-Guerin) and Corynebacterium parvum are

It is preferred that the oligopeptides, peptides, or fragments used to induce antibodies to ENZM have an amino acid sequence consisting of at least about 5 amino acids, and generally will consist of at least about 10 amino acids. It is also preferable that these oligopeptides, peptides, or fragments are substantially identical to a portion of the amino acid sequence of the natural protein.

25 Short stretches of ENZM amino acids may be fused with those of another protein, such as KLH, and antibodies to the chimeric molecule may be produced.

Monoclonal antibodies to ENZM may be prepared using any technique which provides for the production of antibody molecules by continuous cell lines in culture. These include, but are not limited to, the hybridoma technique, the human B-cell hybridoma technique, and the EBV-hybridoma technique (Kohler, G. et al. (1975) Nature 256:495-497; Kozbor, D. et al. (1985) J. Immunol. Methods 81:31-42; Cote, R.J. et al. (1983) Proc. Natl. Acad. Sci. USA 80:2026-2030; Cole, S.P. et al. (1984) Mol. Cell Biol. 62:109-120).

In addition, techniques developed for the production of "chimeric antibodies," such as the

splicing of mouse antibody genes to human antibody genes to obtain a molecule with appropriate antigen specificity and biological activity, can be used (Morrison, S.L. et al. (1984) Proc. Natl. Acad. Sci. USA 81:6851-6855; Neuberger, M.S. et al. (1984) Nature 312:604-608; Takeda, S. et al. (1985) Nature 314:452-454). Alternatively, techniques described for the production of single chain antibodies may be adapted, using methods known in the art, to produce ENZM-specific single chain antibodies. Antibodies with related specificity, but of distinct idiotypic composition, may be generated by chain shuffling from random combinatorial immunoglobulin libraries (Burton, D.R. (1991) Proc. Natl. Acad. Sci. USA 88:10134-10137).

Antibodies may also be produced by inducing *in vivo* production in the lymphocyte population or by screening immunoglobulin libraries or panels of highly specific binding reagents as disclosed in the literature (Orlandi, R. et al. (1989) Proc. Natl. Acad. Sci. USA 86:3833-3837; Winter, G. et al. (1991) Nature 349:293-299).

Antibody fragments which contain specific binding sites for ENZM may also be generated.

For example, such fragments include, but are not limited to, F(ab)<sub>2</sub> fragments produced by pepsin

digestion of the antibody molecule and Fab fragments generated by reducing the disulfide bridges of
the F(ab)<sub>2</sub> fragments. Alternatively, Fab expression libraries may be constructed to allow rapid and
easy identification of monoclonal Fab fragments with the desired specificity (Huse, W.D. et al. (1989)
Science 246:1275-1281).

Various immunoassays may be used for screening to identify antibodies having the desired

specificity. Numerous protocols for competitive binding or immunoradiometric assays using either
polyclonal or monoclonal antibodies with established specificities are well known in the art. Such
immunoassays typically involve the measurement of complex formation between ENZM and its
specific antibody. A two-site, monoclonal-based immunoassay utilizing monoclonal antibodies reactive
to two non-interfering ENZM epitopes is generally used, but a competitive binding assay may also be

25 employed (Pound, supra).

Various methods such as Scatchard analysis in conjunction with radioimmunoassay techniques may be used to assess the affinity of antibodies for ENZM. Affinity is expressed as an association constant, K<sub>n</sub>, which is defined as the molar concentration of ENZM-antibody complex divided by the molar concentrations of free antigen and free antibody under equilibrium conditions. The K<sub>n</sub> determined for a preparation of polyclonal antibodies, which are heterogeneous in their affinities for multiple ENZM epitopes, represents the average affinity, or avidity, of the antibodies for ENZM. The K<sub>n</sub> determined for a preparation of monoclonal antibodies, which are monospecific for a particular ENZM epitope, represents a true measure of affinity. High-affinity antibody preparations with K<sub>n</sub>

ranging from about 10° to 1012 L/mole are preferred for use in immunoassays in which the ENZMantibody complex must withstand rigorous manipulations. Low-affinity antibody preparations with K. ranging from about 106 to 107 L/mole are preferred for use in immunopurification and similar procedures which ultimately require dissociation of ENZM, preferably in active form, from the 5 antibody (Catty, D. (1988) Antibodies, Volume I: A Practical Approach, IRL Press, Washington DC; Liddell, J.E. and A. Cryer (1991) A Practical Guide to Monoclonal Antibodies, John Wiley & Sons, New York NY).

The titer and avidity of polyclonal antibody preparations may be further evaluated to determine the quality and suitability of such preparations for certain downstream applications. For example, a 10 polyclonal antibody preparation containing at least 1-2 mg specific antibody/ml, preferably 5-10 mg specific antibody/ml, is generally employed in procedures requiring precipitation of ENZM-antibody complexes. Procedures for evaluating antibody specificity, titer, and avidity, and guidelines for antibody quality and usage in various applications, are generally available (Catty, supra; Coligan et al., supra).

In another embodiment of the invention, polynucleotides encoding ENZM, or any fragment or complement thereof, may be used for therapeutic purposes. In one aspect, modifications of gene expression can be achieved by designing complementary sequences or antisense molecules (DNA, RNA, PNA, or modified oligonucleotides) to the coding or regulatory regions of the gene encoding ENZM. Such technology is well known in the art, and antisense oligonucleotides or larger fragments 20 can be designed from various locations along the coding or control regions of sequences encoding ENZM (Agrawal, S., ed. (1996) Antisense Therapeutics, Humana Press, Totawa NJ).

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In therapeutic use, any gene delivery system suitable for introduction of the antisense sequences into appropriate target cells can be used. Antisense sequences can be delivered intracellularly in the form of an expression plasmid which, upon transcription, produces a sequence 25 complementary to at least a portion of the cellular sequence encoding the target protein (Slater, J.E. et al. (1998) J. Allergy Clin. Immunol. 102:469-475; Scanlon, K.J. et al. (1995) FASEB J. 9:1288-1296). Antisense sequences can also be introduced intracellularly through the use of viral vectors, such as retrovirus and adeno-associated virus vectors (Miller, A.D. (1990) Blood 76:271-278; Ausubel et al., supra; Uckert, W. and W. Walther (1994) Pharmacol. Ther. 63:323-347). Other gene delivery 30 mechanisms include liposome-derived systems, artificial viral envelopes, and other systems known in the art (Rossi, J.J. (1995) Br. Med. Bull. 51:217-225; Boado, R.J. et al. (1998) J. Pharm. Sci. 87:1308-1315; Morris, M.C. et al. (1997) Nucleic Acids Res. 25:2730-2736).

In another embodiment of the invention, polynucleotides encoding ENZM may be used for

somatic or germline gene therapy. Gene therapy may be performed to (i) correct a genetic deficiency (e.g., in the cases of severe combined immunodeficiency (SCID)-X1 disease characterized by Xlinked inheritance (Cavazzana-Calvo, M. et al. (2000) Science 288:669-672), severe combined immunodeficiency syndrome associated with an inherited adenosine deaminase (ADA) deficiency 5 (Blaese, R.M. et al. (1995) Science 270:475-480; Bordignon, C. et al. (1995) Science 270:470-475), cystic fibrosis (Zabner, J. et al. (1993) Cell 75:207-216; Crystal, R.G. et al. (1995) Hum. Gene Therapy 6:643-666; Crystal, R.G. et al. (1995) Hum. Gene Therapy 6:667-703), thalassamias, familial hypercholesterolemia, and hemophilia resulting from Factor VIII or Factor IX deficiencies (Crystal, R.G. (1995) Science 270:404-410; Verma, I.M. and N. Somia (1997) Nature 389:239-242)), (ii) 10 express a conditionally lethal gene product (e.g., in the case of cancers which result from unregulated cell proliferation), or (iii) express a protein which affords protection against intracellular parasites (e.g., against human retroviruses, such as human immunodeficiency virus (HIV) (Baltimore, D. (1988) Nature 335:395-396; Poeschla, E. et al. (1996) Proc. Natl. Acad. Sci. USA 93:11395-11399), hepatitis B or C virus (HBV, HCV); fungal parasites, such as Candida albicans and Paracoccidioides 15 brasiliensis; and protozoan parasites such as Plasmodium falciparum and Trypanosoma cruzi). In the case where a genetic deficiency in ENZM expression or regulation causes disease, the expression of ENZM from an appropriate population of transduced cells may alleviate the clinical manifestations caused by the genetic deficiency.

In a further embodiment of the invention, diseases or disorders caused by deficiencies in

20 ENZM are treated by constructing mammalian expression vectors encoding ENZM and introducing these vectors by mechanical means into ENZM-deficient cells. Mechanical transfer technologies for use with cells in vivo or ex vitro include (i) direct DNA microinjection into individual cells, (ii) ballistic gold particle delivery, (iii) liposome-mediated transfection, (iv) receptor-mediated gene transfer, and (v) the use of DNA transposons (Morgan, R.A. and W.F. Anderson (1993) Annu. Rev. Biochem.

25 62:191-217; Ivics, Z. (1997) Cell 91:501-510; Boulay, J.-L. and H. Récipon (1998) Curr. Opin. Biotechnol. 9:445-450).

Expression vectors that may be effective for the expression of ENZM include, but are not limited to, the PCDNA 3.1, EPITAG, PRCCMV2, PREP, PVAX, PCR2-TOPOTA vectors (Invitrogen, Carlsbad CA), PCMV-SCRIPT, PCMV-TAG, PEGSH/PERV (Stratagene, La Jolla CA), and PTET-OFF, PTET-ON, PTRE2, PTRE2-LUC, PTK-HYG (BD Clontech, Palo Alto CA). ENZM may be expressed using (i) a constitutively active promoter, (e.g., from cytomegalovirus (CMV), Rous sarcoma virus (RSV), SV40 virus, thymidine kinase (TK), or β-actin genes), (ii) an inducible promoter (e.g., the tetracycline-regulated promoter (Gossen, M. and H. Bujard (1992) Proc.

Natl. Acad. Sci. USA 89:5547-5551; Gossen, M. et al. (1995) Science 268:1766-1769; Rossi, F.M.V. and H.M. Blau (1998) Curr. Opin. Biotechnol. 9:451-456), commercially available in the T-REX plasmid (Invitrogen); the ecdysone-inducible promoter (available in the plasmids PVGRXR and PIND; Invitrogen); the FK506/rapamycin inducible promoter; or the RU486/mifepristone inducible promoter (Rossi, F.M.V. and H.M. Blau, supra), or (iii) a tissue-specific promoter or the native promoter of the endogenous gene encoding ENZM from a normal individual.

Commercially available liposome transformation kits (e.g., the PERFECT LIPID

TRANSFECTION KIT, available from Invitrogen) allow one with ordinary skill in the art to deliver
polynucleotides to target cells in culture and require minimal effort to optimize experimental

parameters. In the alternative, transformation is performed using the calcium phosphate method
(Graham, F.L. and A.J. Eb (1973) Virology 52:456-467), or by electroporation (Neumann, E. et al.
(1982) EMBO J. 1:841-845). The introduction of DNA to primary cells requires modification of these
standardized mammalian transfection protocols.

In another embodiment of the invention, diseases or disorders caused by genetic defects with 15 respect to ENZM expression are treated by constructing a retrovirus vector consisting of (i) the polynucleotide encoding ENZM under the control of an independent promoter or the retrovirus long terminal repeat (LTR) promoter, (ii) appropriate RNA packaging signals, and (iii) a Rev-responsive element (RRE) along with additional retrovirus cis-acting RNA sequences and coding sequences required for efficient vector propagation. Retrovirus vectors (e.g., PFB and PFBNEO) are 20 commercially available (Stratagene) and are based on published data (Riviere, I. et al. (1995) Proc. Natl. Acad. Sci. USA 92:6733-6737), incorporated by reference herein. The vector is propagated in an appropriate vector producing cell line (VPCL) that expresses an envelope gene with a tropism for receptors on the target cells or a promiscuous envelope protein such as VSVg (Armentano, D. et al. (1987) J. Virol. 61:1647-1650; Bender, M.A. et al. (1987) J. Virol. 61:1639-1646; Adam, M.A. and 25 A.D. Miller (1988) J. Virol. 62:3802-3806; Dull, T. et al. (1998) J. Virol. 72:8463-8471; Zufferey, R. et al. (1998) J. Virol. 72:9873-9880). U.S. Patent No. 5,910,434 to Rigg ("Method for obtaining retrovirus packaging cell lines producing high transducing efficiency retroviral supernatant") discloses a method for obtaining retrovirus packaging cell lines and is hereby incorporated by reference. Propagation of retrovirus vectors, transduction of a population of cells (e.g., CD4+ T-cells), and the 30 return of transduced cells to a patient are procedures well known to persons skilled in the art of gene therapy and have been well documented (Ranga, U. et al. (1997) J. Virol. 71:7020-7029; Bauer, G. et al. (1997) Blood 89:2259-2267; Bonyhadi, M.L. (1997) J. Virol. 71:4707-4716; Ranga, U. et al. (1998) Proc. Natl. Acad. Sci. USA 95:1201-1206; Su, L. (1997) Blood 89:2283-2290).

In an embodiment, an adenovirus-based gene therapy delivery system is used to deliver polynucleotides encoding ENZM to cells which have one or more genetic abnormalities with respect to the expression of ENZM. The construction and packaging of adenovirus-based vectors are well known to those with ordinary skill in the art. Replication defective adenovirus vectors have proven to 5 be versatile for importing genes encoding immunoregulatory proteins into intact islets in the pancreas (Csete, M.E. et al. (1995) Transplantation 27:263-268). Potentially useful adenoviral vectors are described in U.S. Patent No. 5,707,618 to Armentano ("Adenovirus vectors for gene therapy"), hereby incorporated by reference. For adenoviral vectors, see also Antinozzi, P.A. et al. (1999; Annu. Rev. Nutr. 19:511-544) and Verma, I.M. and N. Somia (1997; Nature 18:389:239-242).

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In another embodiment, a herpes-based, gene therapy delivery system is used to deliver polynucleotides encoding ENZM to target cells which have one or more genetic abnormalities with respect to the expression of ENZM. The use of herpes simplex virus (HSV)-based vectors may be especially valuable for introducing ENZM to cells of the central nervous system, for which HSV has a tropism. The construction and packaging of herpes-based vectors are well known to those with 15 ordinary skill in the art. A replication-competent herpes simplex virus (HSV) type 1-based vector has been used to deliver a reporter gene to the eyes of primates (Liu, X. et al. (1999) Exp. Eye Res. 169:385-395). The construction of a HSV-1 virus vector has also been disclosed in detail in U.S. Patent No. 5,804,413 to DeLuca ("Herpes simplex virus strains for gene transfer"), which is hereby incorporated by reference. U.S. Patent No. 5,804,413 teaches the use of recombinant HSV d92 20 which consists of a genome containing at least one exogenous gene to be transferred to a cell under the control of the appropriate promoter for purposes including human gene therapy. Also taught by this patent are the construction and use of recombinant HSV strains deleted for ICP4, ICP27 and ICP22. For HSV vectors, see also Goins, W.F. et al. (1999; J. Virol. 73:519-532) and Xu, H. et al. (1994; Dev. Biol. 163:152-161). The manipulation of cloned herpesvirus sequences, the generation of 25 recombinant virus following the transfection of multiple plasmids containing different segments of the large herpesvirus genomes, the growth and propagation of herpesvirus, and the infection of cells with herpesvirus are techniques well known to those of ordinary skill in the art.

In another embodiment, an alphavirus (positive, single-stranded RNA virus) vector is used to deliver polynucleotides encoding ENZM to target cells. The biology of the prototypic alphavirus, 30 Semliki Forest Virus (SFV), has been studied extensively and gene transfer vectors have been based on the SFV genome (Garoff, H. and K.-J. Li (1998) Curr. Opin. Biotechnol. 9:464-469). During alphavirus RNA replication, a subgenomic RNA is generated that normally encodes the viral capsid proteins. This subgenomic RNA replicates to higher levels than the full length genomic RNA,

resulting in the overproduction of capsid proteins relative to the viral proteins with enzymatic activity (e.g., protease and polymerase). Similarly, inserting the coding sequence for ENZM into the alphavirus genome in place of the capsid-coding region results in the production of a large number of ENZM-coding RNAs and the synthesis of high levels of ENZM in vector transduced cells. While slaphavirus infection is typically associated with cell lysis within a few days, the ability to establish a persistent infection in hamster normal kidney cells (BHK-21) with a variant of Sindbis virus (SIN) indicates that the lytic replication of alphaviruses can be altered to suit the needs of the gene therapy application (Dryga, S.A. et al. (1997) Virology 228:74-83). The wide host range of alphaviruses will allow the introduction of ENZM into a variety of cell types. The specific transduction of a subset of cells in a population may require the sorting of cells prior to transduction. The methods of manipulating infectious cDNA clones of alphaviruses, performing alphavirus cDNA and RNA transfections, and performing alphavirus infections, are well known to those with ordinary skill in the art.

Oligonucleotides derived from the transcription initiation site, e.g., between about positions -10

and +10 from the start site, may also be employed to inhibit gene expression. Similarly, inhibition can
be achieved using triple helix base-pairing methodology. Triple helix pairing is useful because it causes
inhibition of the ability of the double helix to open sufficiently for the binding of polymerases,
transcription factors, or regulatory molecules. Recent therapeutic advances using triplex DNA have
been described in the literature (Gee, J.E. et al. (1994) in Huber, B.E. and B.I. Carr, Molecular and

Immunologic Approaches, Futura Publishing, Mt. Kisco NY, pp. 163-177). A complementary
sequence or antisense molecule may also be designed to block translation of mRNA by preventing the
transcript from binding to ribosomes.

Ribozymes, enzymatic RNA molecules, may also be used to catalyze the specific cleavage of RNA. The mechanism of ribozyme action involves sequence-specific hybridization of the ribozyme molecule to complementary target RNA, followed by endonucleolytic cleavage. For example, engineered hammerhead motif ribozyme molecules may specifically and efficiently catalyze endonucleolytic cleavage of RNA molecules encoding ENZM.

Specific ribozyme cleavage sites within any potential RNA target are initially identified by scanning the target molecule for ribozyme cleavage sites, including the following sequences: GUA, GUU, and GUC. Once identified, short RNA sequences of between 15 and 20 ribonucleotides, corresponding to the region of the target gene containing the cleavage site, may be evaluated for secondary structural features which may render the oligonucleotide inoperable. The suitability of candidate targets may also be evaluated by testing accessibility to hybridization with complementary

oligonucleotides using ribonuclease protection assays.

Complementary ribonucleic acid molecules and ribozymes may be prepared by any method known in the art for the synthesis of nucleic acid molecules. These include techniques for chemically synthesizing oligonucleotides such as solid phase phosphoramidite chemical synthesis. Alternatively, 5 RNA molecules may be generated by in vitro and in vivo transcription of DNA molecules encoding ENZM. Such DNA sequences may be incorporated into a wide variety of vectors with suitable RNA polymerase promoters such as T7 or SP6. Alternatively, these cDNA constructs that synthesize complementary RNA, constitutively or inducibly, can be introduced into cell lines, cells, or tissues.

RNA molecules may be modified to increase intracellular stability and half-life. Possible

modifications include, but are not limited to, the addition of flanking sequences at the 5' and/or 3' ends
of the molecule, or the use of phosphorothioate or 2' O-methyl rather than phosphodiesterase linkages
within the backbone of the molecule. This concept is inherent in the production of PNAs and can be
extended in all of these molecules by the inclusion of nontraditional bases such as inosine, queosine,
and wybutosine, as well as acetyl-, methyl-, thio-, and similarly modified forms of adenine, cytidine,
guanine, thymine, and uridine which are not as easily recognized by endogenous endonucleases.

In other embodiments of the invention, the expression of one or more selected polynucleotides of the present invention can be altered, inhibited, decreased, or silenced using RNA interference (RNAi) or post-transcriptional gene silencing (PTGS) methods known in the art. RNAi is a post-transcriptional mode of gene silencing in which double-stranded RNA (dsRNA) introduced into a targeted cell specifically suppresses the expression of the homologous gene (i.e., the gene bearing the sequence complementary to the dsRNA). This effectively knocks out or substantially reduces the expression of the targeted gene. PTGS can also be accomplished by use of DNA or DNA fragments as well. RNAi methods are described by Fire, A. et al. (1998; Nature 391:806-811) and Gura, T. (2000; Nature 404:804-808). PTGS can also be initiated by introduction of a complementary segment

RNAi can be induced in mammalian cells by the use of small interfering RNA also known as siRNA. siRNA are shorter segments of dsRNA (typically about 21 to 23 nucleotides in length) that result in vivo from cleavage of introduced dsRNA by the action of an endogenous ribonuclease.

30 siRNA appear to be the mediators of the RNAi effect in mammals. The most effective siRNAs appear to be 21 nucleotide dsRNAs with 2 nucleotide 3' overhangs. The use of siRNA for inducing RNAi in mammalian cells is described by Elbashir, S.M. et al. (2001; Nature 411:494-498).

siRNA can be generated indirectly by introduction of dsRNA into the targeted cell.

Alternatively, siRNA can be synthesized directly and introduced into a cell by transfection methods and agents described herein or known in the art (such as liposome-mediated transfection, viral vector methods, or other polynucleotide delivery/introductory methods). Suitable siRNAs can be selected by examining a transcript of the target polynucleotide (e.g., mRNA) for nucleotide sequences 5 downstream from the AUG start codon and recording the occurrence of each nucleotide and the 3' adjacent 19 to 23 nucleotides as potential siRNA target sites, with sequences having a 21 nucleotide length being preferred. Regions to be avoided for target siRNA sites include the 5' and 3' untranslated regions (UTRs) and regions near the start codon (within 75 bases), as these may be richer in regulatory protein binding sites. UTR-binding proteins and/or translation initiation complexes may 10 interfere with binding of the siRNP endonuclease complex. The selected target sites for siRNA can then be compared to the appropriate genome database (e.g., human, etc.) using BLAST or other sequence comparison algorithms known in the art. Target sequences with significant homology to other coding sequences can be eliminated from consideration. The selected siRNAs can be produced by chemical synthesis methods known in the art or by in vitro transcription using commercially 15 available methods and kits such as the SILENCER siRNA construction kit (Ambion, Austin TX).

In alternative embodiments, long-term gene silencing and/or RNAi effects can be induced in selected tissue using expression vectors that continuously express siRNA. This can be accomplished using expression vectors that are engineered to express hairpin RNAs (shRNAs) using methods known in the art (see, e.g., Brummelkamp, T.R. et al. (2002) Science 296:550-553; and Paddison, P.J. 20 et al. (2002) Genes Dev. 16:948-958). In these and related embodiments, shRNAs can be delivered to target cells using expression vectors known in the art. An example of a suitable expression vector for delivery of siRNA is the PSILENCER1.0-U6 (circular) plasmid (Ambion). Once delivered to the target tissue, shRNAs are processed in vivo into siRNA-like molecules capable of carrying out genespecific silencing.

In various embodiments, the expression levels of genes targeted by RNAi or PTGS methods can be determined by assays for mRNA and/or protein analysis. Expression levels of the mRNA of a targeted gene can be determined, for example, by northern analysis methods using the NORTHERNMAX-GLY kit (Ambion); by microarray methods; by PCR methods; by real time PCR methods; and by other RNA/polynucleotide assays known in the art or described herein. Expression 30 levels of the protein encoded by the targeted gene can be determined, for example, by microarray methods; by polyacrylamide gel electrophoresis; and by Western analysis using standard techniques known in the art.

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An additional embodiment of the invention encompasses a method for screening for a

compound which is effective in altering expression of a polynucleotide encoding ENZM. Compounds which may be effective in altering expression of a specific polynucleotide may include, but are not limited to, oligonucleotides, antisense oligonucleotides, triple helix-forming oligonucleotides, transcription factors and other polypeptide transcriptional regulators, and non-macromolecular chemical entities which are capable of interacting with specific polynucleotide sequences. Effective compounds may alter polynucleotide expression by acting as either inhibitors or promoters of polynucleotide expression. Thus, in the treatment of disorders associated with increased ENZM expression or activity, a compound which specifically inhibits expression of the polynucleotide encoding ENZM may be therapeutically useful, and in the treatment of disorders associated with decreased ENZM expression or activity, a compound which specifically promotes expression of the polynucleotide encoding ENZM may be therapeutically useful.

In various embodiments, one or more test compounds may be screened for effectiveness in altering expression of a specific polynucleotide. A test compound may be obtained by any method commonly known in the art, including chemical modification of a compound known to be effective in 15 altering polynucleotide expression; selection from an existing, commercially-available or proprietary library of naturally-occurring or non-natural chemical compounds; rational design of a compound based on chemical and/or structural properties of the target polynucleotide; and selection from a library of chemical compounds created combinatorially or randomly. A sample comprising a polynucleotide encoding ENZM is exposed to at least one test compound thus obtained. The sample may comprise, for example, an intact or permeabilized cell, or an in vitro cell-free or reconstituted biochemical system. Alterations in the expression of a polynucleotide encoding ENZM are assayed by any method commonly known in the art. Typically, the expression of a specific nucleotide is detected by hybridization with a probe having a nucleotide sequence complementary to the sequence of the polynucleotide encoding ENZM. The amount of hybridization may be quantified, thus forming the 25 basis for a comparison of the expression of the polynucleotide both with and without exposure to one or more test compounds. Detection of a change in the expression of a polynucleotide exposed to a test compound indicates that the test compound is effective in altering the expression of the polynucleotide. A screen for a compound effective in altering expression of a specific polynucleotide can be carried out, for example, using a Schizosaccharomyces pombe gene expression system 30 (Atkins, D. et al. (1999) U.S. Patent No. 5,932,435; Arndt, G.M. et al. (2000) Nucleic Acids Res. 28:E15) or a human cell line such as HeLa cell (Clarke, M.L. et al. (2000) Biochem. Biophys. Res. Commun. 268:8-13). A particular embodiment of the present invention involves screening a combinatorial library of oligonucleotides (such as deoxyribonucleotides, ribonucleotides, peptide nucleic

acids, and modified oligonucleotides) for antisense activity against a specific polynucleotide sequence (Bruice, T.W. et al. (1997) U.S. Patent No. 5,686,242; Bruice, T.W. et al. (2000) U.S. Patent No. 6,022,691).

Many methods for introducing vectors into cells or tissues are available and equally suitable
for use in vivo, in vitro, and ex vivo. For ex vivo therapy, vectors may be introduced into stem cells
taken from the patient and clonally propagated for autologous transplant back into that same patient.
Delivery by transfection, by liposome injections, or by polycationic amino polymers may be achieved
using methods which are well known in the art (Goldman, C.K. et al. (1997) Nat. Biotechnol. 15:462466).

Any of the therapeutic methods described above may be applied to any subject in need of such therapy, including, for example, mammals such as humans, dogs, cats, cows, horses, rabbits, and monkeys.

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An additional embodiment of the invention relates to the administration of a composition which generally comprises an active ingredient formulated with a pharmaceutically acceptable excipient.

15 Excipients may include, for example, sugars, starches, celluloses, gums, and proteins. Various formulations are commonly known and are thoroughly discussed in the latest edition of Remington's Pharmaceutical Sciences (Maack Publishing, Easton PA). Such compositions may consist of ENZM, antibodies to ENZM, and mimetics, agonists, antagonists, or inhibitors of ENZM.

In various embodiments, the compositions described herein, such as pharmaceutical compositions, may be administered by any number of routes including, but not limited to, oral, intravenous, intramuscular, intra-arterial, intramedullary, intrathecal, intraventricular, pulmonary, transdermal, subcutaneous, intraperitoneal, intranasal, enteral, topical, sublingual, or rectal means.

Compositions for pulmonary administration may be prepared in liquid or dry powder form.

These compositions are generally aerosolized immediately prior to inhalation by the patient. In the

case of small molecules (e.g. traditional low molecular weight organic drugs), aerosol delivery of fastacting formulations is well-known in the art. In the case of macromolecules (e.g. larger peptides and
proteins), recent developments in the field of pulmonary delivery via the alveolar region of the lung
have enabled the practical delivery of drugs such as insulin to blood circulation (see, e.g., Patton, J.S.
et al., U.S. Patent No. 5,997,848). Pulmonary delivery allows administration without needle injection,
and obviates the need for potentially toxic penetration enhancers.

Compositions suitable for use in the invention include compositions wherein the active ingredients are contained in an effective amount to achieve the intended purpose. The determination of an effective dose is well within the capability of those skilled in the art.

Specialized forms of compositions may be prepared for direct intracellular delivery of macromolecules comprising ENZM or fragments thereof. For example, liposome preparations containing a cell-impermeable macromolecule may promote cell fusion and intracellular delivery of the macromolecule. Alternatively, ENZM or a fragment thereof may be joined to a short cationic N-5 terminal portion from the HIV Tat-1 protein. Fusion proteins thus generated have been found to transduce into the cells of all tissues, including the brain, in a mouse model system (Schwarze, S.R. et al. (1999) Science 285:1569-1572).

For any compound, the therapeutically effective dose can be estimated initially either in cell culture assays, e.g., of neoplastic cells, or in animal models such as mice, rats, rabbits, dogs, monkeys, 10 or pigs. An animal model may also be used to determine the appropriate concentration range and route of administration. Such information can then be used to determine useful doses and routes for administration in humans.

A therapeutically effective dose refers to that amount of active ingredient, for example ENZM or fragments thereof, antibodies of ENZM, and agonists, antagonists or inhibitors of ENZM, 15 which ameliorates the symptoms or condition. Therapeutic efficacy and toxicity may be determined by standard pharmaceutical procedures in cell cultures or with experimental animals, such as by calculating the ED50 (the dose therapeutically effective in 50% of the population) or LD50 (the dose lethal to 50% of the population) statistics. The dose ratio of toxic to therapeutic effects is the therapeutic index, which can be expressed as the LD<sub>50</sub>/ED<sub>50</sub> ratio. Compositions which exhibit large 20 therapeutic indices are preferred. The data obtained from cell culture assays and animal studies are used to formulate a range of dosage for human use. The dosage contained in such compositions is preferably within a range of circulating concentrations that includes the ED50 with little or no toxicity. The dosage varies within this range depending upon the dosage form employed, the sensitivity of the patient, and the route of administration.

The exact dosage will be determined by the practitioner, in light of factors related to the subject requiring treatment. Dosage and administration are adjusted to provide sufficient levels of the active moiety or to maintain the desired effect. Factors which may be taken into account include the severity of the disease state, the general health of the subject, the age, weight, and gender of the subject, time and frequency of administration, drug combination(s), reaction sensitivities, and response 30 to therapy. Long-acting compositions may be administered every 3 to 4 days, every week, or biweekly depending on the half-life and clearance rate of the particular formulation.

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Normal dosage amounts may vary from about  $0.1~\mu g$  to  $100,000~\mu g$ , up to a total dose of about 1 gram, depending upon the route of administration. Guidance as to particular dosages and

methods of delivery is provided in the literature and generally available to practitioners in the art. Those skilled in the art will employ different formulations for nucleotides than for proteins or their inhibitors. Similarly, delivery of polynucleotides or polypeptides will be specific to particular cells, conditions, locations, etc.

### 5 DIAGNOSTICS

In another embodiment, antibodies which specifically bind ENZM may be used for the diagnosis of disorders characterized by expression of ENZM, or in assays to monitor patients being treated with ENZM or agonists, antagonists, or inhibitors of ENZM. Antibodies useful for diagnostic purposes may be prepared in the same manner as described above for therapeutics. Diagnostic assays for ENZM include methods which utilize the antibody and a label to detect ENZM in human body fluids or in extracts of cells or tissues. The antibodies may be used with or without modification, and may be labeled by covalent or non-covalent attachment of a reporter molecule. A wide variety of reporter molecules, several of which are described above, are known in the art and may be used.

A variety of protocols for measuring ENZM, including ELISAs, RIAs, and FACS, are known

in the art and provide a basis for diagnosing altered or abnormal levels of ENZM expression. Normal
or standard values for ENZM expression are established by combining body fluids or cell extracts
taken from normal mammalian subjects, for example, human subjects, with antibodies to ENZM under
conditions suitable for complex formation. The amount of standard complex formation may be
quantitated by various methods, such as photometric means. Quantities of ENZM expressed in

subject, control, and disease samples from biopsied tissues are compared with the standard values.

Deviation between standard and subject values establishes the parameters for diagnosing disease.

In another embodiment of the invention, polynucleotides encoding ENZM may be used for diagnostic purposes. The polynucleotides which may be used include oligonucleotides, complementary RNA and DNA molecules, and PNAs. The polynucleotides may be used to detect and quantify gene expression in biopsied tissues in which expression of ENZM may be correlated with disease. The diagnostic assay may be used to determine absence, presence, and excess expression of ENZM, and to monitor regulation of ENZM levels during therapeutic intervention.

In one aspect, hybridization with PCR probes which are capable of detecting polynucleotides, including genomic sequences, encoding ENZM or closely related molecules may be used to identify nucleic acid sequences which encode ENZM. The specificity of the probe, whether it is made from a highly specific region, e.g., the 5' regulatory region, or from a less specific region, e.g., a conserved motif, and the stringency of the hybridization or amplification will determine whether the probe identifies only naturally occurring sequences encoding ENZM, allelic variants, or related sequences.

Probes may also be used for the detection of related sequences, and may have at least 50% sequence identity to any of the ENZM encoding sequences. The hybridization probes of the subject invention may be DNA or RNA and may be derived from the sequence of SEQ ID NO:39-76 or from genomic sequences including promoters, enhancers, and introns of the ENZM gene.

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Means for producing specific hybridization probes for polynucleotides encoding ENZM include the cloning of polynucleotides encoding ENZM or ENZM derivatives into vectors for the production of mRNA probes. Such vectors are known in the art, are commercially available, and may be used to synthesize RNA probes in vitro by means of the addition of the appropriate RNA polymerases and the appropriate labeled nucleotides. Hybridization probes may be labeled by a variety of reporter 10 groups, for example, by radionuclides such as <sup>32</sup>P or <sup>35</sup>S, or by enzymatic labels, such as alkaline phosphatase coupled to the probe via avidin/biotin coupling systems, and the like.

Polynucleotides encoding ENZM may be used for the diagnosis of disorders associated with expression of ENZM. Examples of such disorders include, but are not limited to, an autoimmune/inflammatory disorder such as acquired immunodeficiency syndrome (AIDS), Addison's 15 disease, adult respiratory distress syndrome, allergies, ankylosing spondylitis, amyloidosis, anemia, asthma, atherosclerosis, autoimmune hemolytic anemia, autoimmune thyroiditis, autoimmune polyendocrinopathy-candidiasis-ectodermal dystrophy (APECED), bronchitis, cholecystitis, contact dermatitis, Crohn's disease, atopic dermatitis, dermatomyositis, diabetes mellitus, emphysema, episodic lymphopenia with lymphocytotoxins, erythroblastosis fetalis, erythema nodosum, atrophic gastritis, glomerulonephritis, Goodpasture's syndrome, gout, Graves' disease, Hashimoto's thyroiditis, hypereosinophilia, irritable bowel syndrome, multiple sclerosis, myasthenia gravis, myocardial or pericardial inflammation, osteoarthritis, osteoporosis, pancreatitis, polymyositis, psoriasis, Reiter's syndrome, rheumatoid arthritis, scleroderma, Sjögren's syndrome, systemic anaphylaxis, systemic lupus erythematosus, systemic sclerosis, thrombocytopenic purpura, ulcerative colitis, uveitis, Werner syndrome, complications of cancer, hemodialysis, and extracorporeal circulation, and trauma; an infectious disorder such as a viral infection, e.g., caused by an adenovirus (acute respiratory disease, pneumonia), an arenavirus (lymphocytic choriomeningitis), a bunyavirus (Hantavirus), a coronavirus (pneumonia, chronic bronchitis), a hepadnavirus (hepatitis), a herpesvirus (herpes simplex virus, varicella-zoster virus, Epstein-Barr virus, cytomegalovirus), a flavivirus (yellow fever), an 30 orthomyxovirus (influenza), a papillomavirus (cancer), a paramyxovirus (measles, mumps), a picornovirus (rhinovirus, poliovirus, coxsackie-virus), a polyomavirus (BK virus, JC virus), a poxvirus (smallpox), a reovirus (Colorado tick fever), a retrovirus (human immunodeficiency virus, human T lymphotropic virus), a rhabdovirus (rabies), a rotavirus (gastroenteritis), and a togavirus (encephalitis,

rubella), and a bacterial infection, a fungal infection, a parasitic infection, a protozoal infection, and a helminthic infection; an immune deficiency, such as acquired immunodeficiency syndrome (AIDS), Xlinked agammaglobinemia of Bruton, common variable immunodeficiency (CVI), DiGeorge's syndrome (thymic hypoplasia), thymic dysplasia, isolated IgA deficiency, severe combined immunodeficiency disease (SCID), immunodeficiency with thrombocytopenia and eczema (Wiskott-Aldrich syndrome), Chediak-Higashi syndrome, chronic granulomatous diseases, hereditary angioneurotic edema, and immunodeficiency associated with Cushing's disease; a disorder of metabolism such as Addison's disease, cerebrotendinous xanthomatosis, congenital adrenal hyperplasia, coumarin resistance, cystic fibrosis, diabetes, fatty hepatocirrhosis, 10 fructose-1,6-diphosphatase deficiency, galactosemia, goiter, glucagonoma, glycogen storage diseases, hereditary fructose intolerance, hyperadrenalism, hypoadrenalism, hyperparathyroidism, hypoparathyroidism, hypercholesterolemia, hyperthyroidism, hypoglycemia, hypothyroidism, hyperlipidemia, hyperlipemia, a lipid myopathy, a lipodystrophy, a lysosomal storage disease, mannosidosis, neuraminidase deficiency, obesity, pentosuria phenylketonuria, pseudovitamin D-15 deficiency rickets; a reproductive disorder such as a disorder of prolactin production, infertility, including tubal disease, ovulatory defects, and endometriosis, a disruption of the estrous cycle, a disruption of the menstrual cycle, polycystic ovary syndrome, ovarian hyperstimulation syndrome, endometrial and ovarian tumors, uterine fibroids, autoimmune disorders, ectopic pregnancies, and teratogenesis, cancer of the breast, fibrocystic breast disease, and galactorrhea, disruptions of spermatogenesis, abnormal sperm physiology, cancer of the testis, cancer of the prostate, benign prostatic hyperplasia, prostatitis, Peyronie's disease, impotence, carcinoma of the male breast, and gynecomastia; a neurological disorder such as epilepsy, ischemic cerebrovascular disease, stroke, cerebral neoplasms, Alzheimer's disease, Pick's disease, Huntington's disease, dementia, Parkinson's disease and other extrapyramidal disorders, amyotrophic lateral sclerosis and other motor neuron 25 disorders, progressive neural muscular atrophy, retinitis pigmentosa, hereditary ataxias, multiple sclerosis and other demyelinating diseases, bacterial and viral meningitis, brain abscess, subdural empyema, epidural abscess, suppurative intracranial thrombophlebitis, myelitis and radiculitis, viral central nervous system disease; prion diseases including kuru, Creutzfeldt-Jakob disease, and Gerstmann-Straussler-Scheinker syndrome; fatal familial insomnia, nutritional and metabolic diseases of the nervous system, neurofibromatosis, tuberous sclerosis, cerebelloretinal hemangioblastomatosis, encephalotrigeminal syndrome, mental retardation and other developmental disorders of the central

nervous system, cerebral palsy, neuroskeletal disorders, autonomic nervous system disorders, cranial nerve disorders, spinal cord diseases, muscular dystrophy and other neuromuscular disorders,

peripheral nervous system disorders, dermatomyositis and polymyositis; inherited, metabolic, endocrine, and toxic myopathies; myasthenia gravis, periodic paralysis; mental disorders including mood, anxiety, and schizophrenic disorders; seasonal affective disorder (SAD); akathesia, amnesia, catatonia, diabetic neuropathy, tardive dyskinesia, dystonias, paranoid psychoses, postherpetic neuralgia, and Tourette's 5 disorder; a cardiovascular disorder, such as arteriovenous fistula, atherosclerosis, hypertension, vasculitis, Raynaud's disease, aneurysms, arterial dissections, varicose veins, thrombophlebitis and phlebothrombosis, vascular tumors, and complications of thrombolysis, balloon angioplasty, vascular replacement, and coronary artery bypass graft surgery, congestive heart failure, ischemic heart disease, angina pectoris, myocardial infarction, hypertensive heart disease, degenerative valvular heart 10 disease, calcific aortic valve stenosis, congenitally bicuspid aortic valve, mitral annular calcification, mitral valve prolapse, rheumatic fever and rheumatic heart disease, infective endocarditis, nonbacterial thrombotic endocarditis, endocarditis of systemic lupus erythematosus, carcinoid heart disease, cardiomyopathy, myocarditis, pericarditis, neoplastic heart disease, congenital heart disease, and complications of cardiac transplantation, congenital lung anomalies, atelectasis, pulmonary congestion 15 and edema, pulmonary embolism, pulmonary hemorrhage, pulmonary infarction, pulmonary hypertension, vascular sclerosis, obstructive pulmonary disease, restrictive pulmonary disease, chronic obstructive pulmonary disease, emphysema, chronic bronchitis, bronchial asthma, bronchiectasis, bacterial pneumonia, viral and mycoplasmal pneumonia, lung abscess, pulmonary tuberculosis, diffuse interstitial diseases, pneumoconioses, sarcoidosis, idiopathic pulmonary fibrosis, desquamative interstitial pneumonitis, hypersensitivity pneumonitis, pulmonary eosinophilia bronchiolitis obliterans-organizing pneumonia, diffuse pulmonary hemorrhage syndromes, Goodpasture's syndromes, idiopathic pulmonary hemosiderosis, pulmonary involvement in collagen-vascular disorders, pulmonary alveolar proteinosis, lung tumors, inflammatory and noninflammatory pleural effusions, pneumothorax, pleural tumors, drug-induced lung disease, radiation-induced lung disease, and complications of lung 25 transplantation; an eye disorder such as ocular hypertension and glaucoma; a disorder of cell proliferation such as actinic keratosis, arteriosclerosis, atherosclerosis, bursitis, cirrhosis, hepatitis, mixed connective tissue disease (MCTD), myelofibrosis, paroxysmal nocturnal hemoglobinuria, polycythemia vera, psoriasis, primary thrombocythemia; and a cancer, including adenocarcinoma, leukemia, lymphoma, melanoma, myeloma, sarcoma, teratocarcinoma, and, in particular, cancers of 30 the adrenal gland, bladder, bone, bone marrow, brain, breast, cervix, colon, gall bladder, ganglia, gastrointestinal tract, heart, kidney, liver, lung, muscle, ovary, pancreas, parathyroid, penis, prostate, salivary glands, skin, spleen, testis, thymus, thyroid, and uterus. Polynucleotides encoding ENZM may be used in Southern or northern analysis, dot blot, or other membrane-based technologies; in PCR

technologies; in dipstick, pin, and multiformat ELISA-like assays; and in microarrays utilizing fluids or tissues from patients to detect altered ENZM expression. Such qualitative or quantitative methods are well known in the art.

In a particular embodiment, polynucleotides encoding ENZM may be used in assays that

detect the presence of associated disorders, particularly those mentioned above. Polynucleotides
complementary to sequences encoding ENZM may be labeled by standard methods and added to a
fluid or tissue sample from a patient under conditions suitable for the formation of hybridization
complexes. After a suitable incubation period, the sample is washed and the signal is quantified and
compared with a standard value. If the amount of signal in the patient sample is significantly altered in

comparison to a control sample then the presence of altered levels of polynucleotides encoding ENZM
in the sample indicates the presence of the associated disorder. Such assays may also be used to
evaluate the efficacy of a particular therapeutic treatment regimen in animal studies, in clinical trials,
or to monitor the treatment of an individual patient.

In order to provide a basis for the diagnosis of a disorder associated with expression of

ENZM, a normal or standard profile for expression is established. This may be accomplished by

combining body fluids or cell extracts taken from normal subjects, either animal or human, with a

sequence, or a fragment thereof, encoding ENZM, under conditions suitable for hybridization or

amplification. Standard hybridization may be quantified by comparing the values obtained from normal

subjects with values from an experiment in which a known amount of a substantially purified

polynucleotide is used. Standard values obtained in this manner may be compared with values

obtained from samples from patients who are symptomatic for a disorder. Deviation from standard

values is used to establish the presence of a disorder.

Once the presence of a disorder is established and a treatment protocol is initiated,
hybridization assays may be repeated on a regular basis to determine if the level of expression in the

patient begins to approximate that which is observed in the normal subject. The results obtained from
successive assays may be used to show the efficacy of treatment over a period ranging from several
days to months.

With respect to cancer, the presence of an abnormal amount of transcript (either under- or overexpressed) in biopsied tissue from an individual may indicate a predisposition for the development of the disease, or may provide a means for detecting the disease prior to the appearance of actual clinical symptoms. A more definitive diagnosis of this type may allow health professionals to employ preventative measures or aggressive treatment earlier, thereby preventing the development or further progression of the cancer.

Additional diagnostic uses for oligonucleotides designed from the sequences encoding ENZM may involve the use of PCR. These oligomers may be chemically synthesized, generated enzymatically, or produced in vitro. Oligomers will preferably contain a fragment of a polynucleotide encoding ENZM, or a fragment of a polynucleotide complementary to the polynucleotide encoding ENZM, and will be employed under optimized conditions for identification of a specific gene or condition. Oligomers may also be employed under less stringent conditions for detection or quantification of closely related DNA or RNA sequences.

In a particular aspect, oligonucleotide primers derived from polynucleotides encoding ENZM may be used to detect single nucleotide polymorphisms (SNPs). SNPs are substitutions, insertions and 10 deletions that are a frequent cause of inherited or acquired genetic disease in humans. Methods of SNP detection include, but are not limited to, single-stranded conformation polymorphism (SSCP) and fluorescent SSCP (fSSCP) methods. In SSCP, oligonucleotide primers derived from polynucleotides encoding ENZM are used to amplify DNA using the polymerase chain reaction (PCR). The DNA may be derived, for example, from diseased or normal tissue, biopsy samples, bodily fluids, and the 15 like. SNPs in the DNA cause differences in the secondary and tertiary structures of PCR products in single-stranded form, and these differences are detectable using gel electrophoresis in non-denaturing gels. In fSCCP, the oligonucleotide primers are fluorescently labeled, which allows detection of the amplimers in high-throughput equipment such as DNA sequencing machines. Additionally, sequence database analysis methods, termed in silico SNP (isSNP), are capable of identifying polymorphisms by 20 comparing the sequence of individual overlapping DNA fragments which assemble into a common consensus sequence. These computer-based methods filter out sequence variations due to laboratory preparation of DNA and sequencing errors using statistical models and automated analyses of DNA sequence chromatograms. In the alternative, SNPs may be detected and characterized by mass spectrometry using, for example, the high throughput MASSARRAY system (Sequenom, Inc., San 25 Diego CA).

SNPs may be used to study the genetic basis of human disease. For example, at least 16 common SNPs have been associated with non-insulin-dependent diabetes mellitus. SNPs are also useful for examining differences in disease outcomes in monogenic disorders, such as cystic fibrosis, sickle cell anemia, or chronic granulomatous disease. For example, variants in the mannose-binding lectin, MBL2, have been shown to be correlated with deleterious pulmonary outcomes in cystic fibrosis. SNPs also have utility in pharmacogenomics, the identification of genetic variants that influence a patient's response to a drug, such as life-threatening toxicity. For example, a variation in N-acetyl transferase is associated with a high incidence of peripheral neuropathy in response to the

anti-tuberculosis drug isoniazid, while a variation in the core promoter of the ALOX5 gene results in diminished clinical response to treatment with an anti-asthma drug that targets the 5-lipoxygenase pathway. Analysis of the distribution of SNPs in different populations is useful for investigating genetic drift, mutation, recombination, and selection, as well as for tracing the origins of populations and their migrations (Taylor, J.G. et al. (2001) Trends Mol. Med. 7:507-512; Kwok, P.-Y. and Z. Gu (1999) Mol. Med. Today 5:538-543; Nowotny, P. et al. (2001) Curr. Opin. Neurobiol. 11:637-641).

Methods which may also be used to quantify the expression of ENZM include radiolabeling or biotinylating nucleotides, coamplification of a control nucleic acid, and interpolating results from standard curves (Melby, P.C. et al. (1993) J. Immunol. Methods 159:235-244; Duplaa, C. et al. (1993) 10 Anal. Biochem. 212:229-236). The speed of quantitation of multiple samples may be accelerated by running the assay in a high-throughput format where the oligomer or polynucleotide of interest is presented in various dilutions and a spectrophotometric or colorimetric response gives rapid quantitation.

In further embodiments, oligonucleotides or longer fragments derived from any of the
polynucleotides described herein may be used as elements on a microarray. The microarray can be
used in transcript imaging techniques which monitor the relative expression levels of large numbers of
genes simultaneously as described below. The microarray may also be used to identify genetic
variants, mutations, and polymorphisms. This information may be used to determine gene function, to
understand the genetic basis of a disorder, to diagnose a disorder, to monitor progression/regression of
disease as a function of gene expression, and to develop and monitor the activities of therapeutic
agents in the treatment of disease. In particular, this information may be used to develop a
pharmacogenomic profile of a patient in order to select the most appropriate and effective treatment
regimen for that patient. For example, therapeutic agents which are highly effective and display the
fewest side effects may be selected for a patient based on his/her pharmacogenomic profile.

In another embodiment, ENZM, fragments of ENZM, or antibodies specific for ENZM may be used as elements on a microarray. The microarray may be used to monitor or measure protein-protein interactions, drug-target interactions, and gene expression profiles, as described above.

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A particular embodiment relates to the use of the polynucleotides of the present invention to generate a transcript image of a tissue or cell type. A transcript image represents the global pattern of gene expression by a particular tissue or cell type. Global gene expression patterns are analyzed by quantifying the number of expressed genes and their relative abundance under given conditions and at a given time (Seilhamer et al., "Comparative Gene Transcript Analysis," U.S. Patent No. 5,840,484; hereby expressly incorporated by reference herein). Thus a transcript image may be generated by

hybridizing the polynucleotides of the present invention or their complements to the totality of transcripts or reverse transcripts of a particular tissue or cell type. In one embodiment, the hybridization takes place in high-throughput format, wherein the polynucleotides of the present invention or their complements comprise a subset of a plurality of elements on a microarray. The resultant transcript image would provide a profile of gene activity.

Transcript images may be generated using transcripts isolated from tissues, cell lines, biopsies, or other biological samples. The transcript image may thus reflect gene expression in vivo, as in the case of a tissue or biopsy sample, or in vitro, as in the case of a cell line.

Transcript images which profile the expression of the polynucleotides of the present invention 10 may also be used in conjunction with in vitro model systems and preclinical evaluation of pharmaceuticals, as well as toxicological testing of industrial and naturally-occurring environmental compounds. All compounds induce characteristic gene expression patterns, frequently termed molecular fingerprints or toxicant signatures, which are indicative of mechanisms of action and toxicity (Nuwaysir, E.F. et al. (1999) Mol. Carcinog. 24:153-159; Steiner, S. and N.L. Anderson (2000) 15 Toxicol. Lett. 112-113:467-471). If a test compound has a signature similar to that of a compound with known toxicity, it is likely to share those toxic properties. These fingerprints or signatures are most useful and refined when they contain expression information from a large number of genes and gene families. Ideally, a genome-wide measurement of expression provides the highest quality signature. Even genes whose expression is not altered by any tested compounds are important as 20 well, as the levels of expression of these genes are used to normalize the rest of the expression data. The normalization procedure is useful for comparison of expression data after treatment with different compounds. While the assignment of gene function to elements of a toxicant signature aids in interpretation of toxicity mechanisms, knowledge of gene function is not necessary for the statistical matching of signatures which leads to prediction of toxicity (see, for example, Press Release 00-02 25 from the National Institute of Environmental Health Sciences, released February 29, 2000, available at http://www.niehs.nih.gov/oc/news/toxchip.htm). Therefore, it is important and desirable in toxicological screening using toxicant signatures to include all expressed gene sequences.

In an embodiment, the toxicity of a test compound can be assessed by treating a biological sample containing nucleic acids with the test compound. Nucleic acids that are expressed in the treated biological sample are hybridized with one or more probes specific to the polynucleotides of the present invention, so that transcript levels corresponding to the polynucleotides of the present invention may be quantified. The transcript levels in the treated biological sample are compared with levels in an untreated biological sample. Differences in the transcript levels between the two samples are

indicative of a toxic response caused by the test compound in the treated sample.

Another embodiment relates to the use of the polypeptides disclosed herein to analyze the proteome of a tissue or cell type. The term proteome refers to the global pattern of protein expression in a particular tissue or cell type. Each protein component of a proteome can be subjected individually 5 to further analysis. Proteome expression patterns, or profiles, are analyzed by quantifying the number of expressed proteins and their relative abundance under given conditions and at a given time. A profile of a cell's proteome may thus be generated by separating and analyzing the polypeptides of a particular tissue or cell type. In one embodiment, the separation is achieved using two-dimensional gel electrophoresis, in which proteins from a sample are separated by isoelectric focusing in the first 10 dimension, and then according to molecular weight by sodium dodecyl sulfate slab gel electrophoresis in the second dimension (Steiner and Anderson, supra). The proteins are visualized in the gel as discrete and uniquely positioned spots, typically by staining the gel with an agent such as Coomassie Blue or silver or fluorescent stains. The optical density of each protein spot is generally proportional to the level of the protein in the sample. The optical densities of equivalently positioned protein spots 15 from different samples, for example, from biological samples either treated or untreated with a test compound or therapeutic agent, are compared to identify any changes in protein spot density related to the treatment. The proteins in the spots are partially sequenced using, for example, standard methods employing chemical or enzymatic cleavage followed by mass spectrometry. The identity of the protein in a spot may be determined by comparing its partial sequence, preferably of at least 5 contiguous 20 amino acid residues, to the polypeptide sequences of interest. In some cases, further sequence data may be obtained for definitive protein identification.

A proteomic profile may also be generated using antibodies specific for ENZM to quantify the levels of ENZM expression. In one embodiment, the antibodies are used as elements on a microarray, and protein expression levels are quantified by contacting the microarray with the sample and detecting the levels of protein bound to each array element (Lueking, A. et al. (1999) Anal. Biochem. 270:103-111; Mendoze, L.G. et al. (1999) Biotechniques 27:778-788). Detection may be performed by a variety of methods known in the art, for example, by reacting the proteins in the sample with a thiolor amino-reactive fluorescent compound and detecting the amount of fluorescence bound at each array element.

Toxicant signatures at the proteome level are also useful for toxicological screening, and should be analyzed in parallel with toxicant signatures at the transcript level. There is a poor correlation between transcript and protein abundances for some proteins in some tissues (Anderson, N.L. and J. Seilhamer (1997) Electrophoresis 18:533-537), so proteome toxicant signatures may be

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useful in the analysis of compounds which do not significantly affect the transcript image, but which alter the proteomic profile. In addition, the analysis of transcripts in body fluids is difficult, due to rapid degradation of mRNA, so proteomic profiling may be more reliable and informative in such cases.

In another embodiment, the toxicity of a test compound is assessed by treating a biological 5 sample containing proteins with the test compound. Proteins that are expressed in the treated biological sample are separated so that the amount of each protein can be quantified. The amount of each protein is compared to the amount of the corresponding protein in an untreated biological sample. A difference in the amount of protein between the two samples is indicative of a toxic response to the test compound in the treated sample. Individual proteins are identified by sequencing the amino acid 10 residues of the individual proteins and comparing these partial sequences to the polypeptides of the present invention.

In another embodiment, the toxicity of a test compound is assessed by treating a biological sample containing proteins with the test compound. Proteins from the biological sample are incubated with antibodies specific to the polypeptides of the present invention. The amount of protein recognized 15 by the antibodies is quantified. The amount of protein in the treated biological sample is compared with the amount in an untreated biological sample. A difference in the amount of protein between the two samples is indicative of a toxic response to the test compound in the treated sample.

Microarrays may be prepared, used, and analyzed using methods known in the art (Brennan, T.M. et al. (1995) U.S. Patent No. 5,474,796; Schena, M. et al. (1996) Proc. Natl. Acad. Sci. USA 20 93:10614-10619; Baldeschweiler et al. (1995) PCT application WO95/25116; Shalon, D. et al. (1995) PCT application WO95/35505; Heller, R.A. et al. (1997) Proc. Natl. Acad. Sci. USA 94:2150-2155; Heller, M.J. et al. (1997) U.S. Patent No. 5,605,662). Various types of microarrays are well known and thoroughly described in Schena, M., ed. (1999; DNA Microarrays: A Practical Approach, Oxford University Press, London).

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In another embodiment of the invention, nucleic acid sequences encoding ENZM may be used to generate hybridization probes useful in mapping the naturally occurring genomic sequence. Either coding or noncoding sequences may be used, and in some instances, noncoding sequences may be preferable over coding sequences. For example, conservation of a coding sequence among members of a multi-gene family may potentially cause undesired cross hybridization during chromosomal 30 mapping. The sequences may be mapped to a particular chromosome, to a specific region of a chromosome, or to artificial chromosome constructions, e.g., human artificial chromosomes (HACs), yeast artificial chromosomes (YACs), bacterial artificial chromosomes (BACs), bacterial P1 constructions, or single chromosome cDNA libraries (Harrington, J.J. et al. (1997) Nat. Genet. 15:345-

355; Price, C.M. (1993) Blood Rev. 7:127-134; Trask, B.J. (1991) Trends Genet. 7:149-154). Once mapped, the nucleic acid sequences may be used to develop genetic linkage maps, for example, which correlate the inheritance of a disease state with the inheritance of a particular chromosome region or restriction fragment length polymorphism (RFLP) (Lander, E.S. and D. Botstein (1986) Proc. Natl.
5 Acad. Sci. USA 83:7353-7357).

Fluorescent in situ hybridization (FISH) may be correlated with other physical and genetic map data (Heinz-Ulrich, et al. (1995) in Meyers, supra, pp. 965-968). Examples of genetic map data can be found in various scientific journals or at the Online Mendelian Inheritance in Man (OMIM) World Wide Web site. Correlation between the location of the gene encoding ENZM on a physical map and a specific disorder, or a predisposition to a specific disorder, may help define the region of DNA associated with that disorder and thus may further positional cloning efforts.

In situ hybridization of chromosomal preparations and physical mapping techniques, such as linkage analysis using established chromosomal markers, may be used for extending genetic maps.

Often the placement of a gene on the chromosome of another mammalian species, such as mouse, may reveal associated markers even if the exact chromosomal locus is not known. This information is valuable to investigators searching for disease genes using positional cloning or other gene discovery techniques. Once the gene or genes responsible for a disease or syndrome have been crudely localized by genetic linkage to a particular genomic region, e.g., ataxia-telangiectasia to 11q22-23, any sequences mapping to that area may represent associated or regulatory genes for further investigation (Gatti, R.A. et al. (1988) Nature 336:577-580). The nucleotide sequence of the instant invention may also be used to detect differences in the chromosomal location due to translocation, inversion, etc., among normal, carrier, or affected individuals.

In another embodiment of the invention, ENZM, its catalytic or immunogenic fragments, or oligopeptides thereof can be used for screening libraries of compounds in any of a variety of drug screening techniques. The fragment employed in such screening may be free in solution, affixed to a solid support, borne on a cell surface, or located intracellularly. The formation of binding complexes between ENZM and the agent being tested may be measured.

Another technique for drug screening provides for high throughput screening of compounds having suitable binding affinity to the protein of interest (Geysen, et al. (1984) PCT application

30 WO84/03564). In this method, large numbers of different small test compounds are synthesized on a solid substrate. The test compounds are reacted with ENZM, or fragments thereof, and washed. Bound ENZM is then detected by methods well known in the art. Purified ENZM can also be coated directly onto plates for use in the aforementioned drug screening techniques. Alternatively,

non-neutralizing antibodies can be used to capture the peptide and immobilize it on a solid support.

In another embodiment, one may use competitive drug screening assays in which neutralizing antibodies capable of binding ENZM specifically compete with a test compound for binding ENZM. In this manner, antibodies can be used to detect the presence of any peptide which shares one or more 5 antigenic determinants with ENZM.

In additional embodiments, the nucleotide sequences which encode ENZM may be used in any molecular biology techniques that have yet to be developed, provided the new techniques rely on properties of nucleotide sequences that are currently known, including, but not limited to, such properties as the triplet genetic code and specific base pair interactions.

Without further elaboration, it is believed that one skilled in the art can, using the preceding description, utilize the present invention to its fullest extent. The following embodiments are, therefore, to be construed as merely illustrative, and not limitative of the remainder of the disclosure in any way whatsoever.

The disclosures of all patents, applications, and publications mentioned above and below, 15 including U.S. Ser. No. 60/387,119, and U.S. Ser. No. 60/390,662, are hereby expressly incorporated by reference.

### EXAMPLES

#### Construction of cDNA Libraries I.

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Incyte cDNAs are derived from cDNA libraries described in the LIFESEQ database (Incyte, Palo Alto CA). Some tissues are homogenized and lysed in guanidinium isothiocyanate, while others are homogenized and lysed in phenol or in a suitable mixture of denaturants, such as TRIZOL (Invitrogen), a monophasic solution of phenol and guanidine isothiocyanate. The resulting lysates are centrifuged over CsCl cushions or extracted with chloroform. RNA is precipitated from the lysates 25 with either isopropanol or sodium acetate and ethanol, or by other routine methods.

Phenol extraction and precipitation of RNA are repeated as necessary to increase RNA purity. In some cases, RNA is treated with DNase. For most libraries, poly(A)+ RNA is isolated using oligo d(T)-coupled paramagnetic particles (Promega), OLIGOTEX latex particles (QIAGEN, Chatsworth CA), or an OLIGOTEX mRNA purification kit (QIAGEN). Alternatively, RNA is 30 isolated directly from tissue lysates using other RNA isolation kits, e.g., the POLY(A)PURE mRNA purification kit (Ambion, Austin TX).

In some cases, Stratagene is provided with RNA and constructs the corresponding cDNA libraries. Otherwise, cDNA is synthesized and cDNA libraries are constructed with the UNIZAP

vector system (Stratagene) or SUPERSCRIPT plasmid system (Invitrogen), using the recommended procedures or similar methods known in the art (Ausubel et al., supra, ch. 5). Reverse transcription is initiated using oligo d(T) or random primers. Synthetic oligonucleotide adapters are ligated to double stranded cDNA, and the cDNA is digested with the appropriate restriction enzyme or enzymes. For 5 most libraries, the cDNA is size-selected (300-1000 bp) using SEPHACRYL S1000, SEPHAROSE CL2B, or SEPHAROSE CL4B column chromatography (Amersham Biosciences) or preparative agarose gel electrophoresis. cDNAs are ligated into compatible restriction enzyme sites of the polylinker of a suitable plasmid, e.g., PBLUESCRIPT plasmid (Stratagene), PSPORT1 plasmid (Invitrogen, Carlsbad CA), PCDNA2.1 plasmid (Invitrogen), PBK-CMV plasmid (Stratagene), PCR2-10 TOPOTA plasmid (Invitrogen), PCMV-ICIS plasmid (Stratagene), pIGEN (Incyte, Palo Alto CA), pRARE (Incyte), or pINCY (Incyte), or derivatives thereof. Recombinant plasmids are transformed into competent E. coli cells including XL1-Blue, XL1-BlueMRF, or SOLR from Stratagene or DH5α, DH10B, or ElectroMAX DH10B from Invitrogen.

#### TT. Isolation of cDNA Clones

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Plasmids obtained as described in Example I are recovered from host cells by in vivo excision using the UNIZAP vector system (Stratagene) or by cell lysis. Plasmids are purified using at least one of the following: a Magic or WIZARD Minipreps DNA purification system (Promega); an AGTC Miniprep purification kit (Edge Biosystems, Gaithersburg MD); and QIAWELL 8 Plasmid, QIAWELL 8 Plus Plasmid, QIAWELL 8 Ultra Plasmid purification systems or the R.E.A.L. PREP 20 96 plasmid purification kit from QIAGEN. Following precipitation, plasmids are resuspended in 0.1 ml of distilled water and stored, with or without lyophilization, at 4°C.

Alternatively, plasmid DNA is amplified from host cell lysates using direct link PCR in a highthroughput format (Rao, V.B. (1994) Anal. Biochem. 216:1-14). Host cell lysis and thermal cycling steps are carried out in a single reaction mixture. Samples are processed and stored in 384-well 25 plates, and the concentration of amplified plasmid DNA is quantified fluorometrically using PICOGREEN dye (Molecular Probes, Eugene OR) and a FLUOROSKAN II fluorescence scanner (Labsystems Oy, Helsinki, Finland).

#### Sequencing and Analysis III.

Incyte cDNA recovered in plasmids as described in Example II are sequenced as follows. 30 Sequencing reactions are processed using standard methods or high-throughput instrumentation such as the ABI CATALYST 800 (Applied Biosystems) thermal cycler or the PTC-200 thermal cycler (MJ Research) in conjunction with the HYDRA microdispenser (Robbins Scientific) or the MICROLAB 2200 (Hamilton) liquid transfer system. cDNA sequencing reactions are prepared using

reagents provided by Amersham Biosciences or supplied in ABI sequencing kits such as the ABI
PRISM BIGDYE Terminator cycle sequencing ready reaction kit (Applied Biosystems).

Electrophoretic separation of cDNA sequencing reactions and detection of labeled polynucleotides are
carried out using the MEGABACE 1000 DNA sequencing system (Amersham Biosciences); the ABI
PRISM 373 or 377 sequencing system (Applied Biosystems) in conjunction with standard ABI
protocols and base calling software; or other sequence analysis systems known in the art. Reading
frames within the cDNA sequences are identified using standard methods (Ausubel et al., supra, ch.
7). Some of the cDNA sequences are selected for extension using the techniques disclosed in
Example VIII.

Polynucleotide sequences derived from Incyte cDNAs are validated by removing vector, 10 linker, and poly(A) sequences and by masking ambiguous bases, using algorithms and programs based on BLAST, dynamic programming, and dinucleotide nearest neighbor analysis. The Incyte cDNA sequences or translations thereof are then queried against a selection of public databases such as the GenBank primate, rodent, mammalian, vertebrate, and eukaryote databases, and BLOCKS, PRINTS, 15 DOMO, PRODOM; PROTEOME databases with sequences from Homo sapiens, Rattus norvegicus, Mus musculus, Caenorhabditis elegans, Saccharomyces cerevisiae, Schizosaccharomyces pombe, and Candida albicans (Incyte, Palo Alto CA); hidden Markov model (HMM)-based protein family databases such as PFAM, INCY, and TIGRFAM (Haft, D.H. et al. (2001) Nucleic Acids Res. 29:41-43); and HMM-based protein domain databases such as SMART 20 (Schultz, J. et al. (1998) Proc. Natl. Acad. Sci. USA 95:5857-5864; Letunic, I. et al. (2002) Nucleic Acids Res. 30:242-244). (HMM is a probabilistic approach which analyzes consensus primary structures of gene families; see, for example, Eddy, S.R. (1996) Curr. Opin. Struct. Biol. 6:361-365.) The queries are performed using programs based on BLAST, FASTA, BLIMPS, and HMMER. The Incyte cDNA sequences are assembled to produce full length polynucleotide sequences. 25 Alternatively, GenBank cDNAs, GenBank ESTs, stitched sequences, stretched sequences, or Genscan-predicted coding sequences (see Examples IV and V) are used to extend Incyte cDNA assemblages to full length. Assembly is performed using programs based on Phred, Phrap, and Consed, and cDNA assemblages are screened for open reading frames using programs based on GeneMark, BLAST, and FASTA. The full length polynucleotide sequences are translated to derive 30 the corresponding full length polypeptide sequences. Alternatively, a polypeptide may begin at any of the methionine residues of the full length translated polypeptide. Full length polypeptide sequences are subsequently analyzed by querying against databases such as the GenBank protein databases

(genpept), SwissProt, the PROTEOME databases, BLOCKS, PRINTS, DOMO, PRODOM, Prosite,

hidden Markov model (HMM)-based protein family databases such as PFAM, INCY, and TIGRFAM; and HMM-based protein domain databases such as SMART. Full length polynucleotide sequences are also analyzed using MACDNASIS PRO software (MiraiBio, Alameda CA) and LASERGENE software (DNASTAR). Polynucleotide and polypeptide sequence alignments are generated using default parameters specified by the CLUSTAL algorithm as incorporated into the MEGALIGN multisequence alignment program (DNASTAR), which also calculates the percent identity between aligned sequences.

Table 7 summarizes tools, programs, and algorithms used for the analysis and assembly of
Incyte cDNA and full length sequences and provides applicable descriptions, references, and threshold
parameters. The first column of Table 7 shows the tools, programs, and algorithms used, the second
column provides brief descriptions thereof, the third column presents appropriate references, all of
which are incorporated by reference herein in their entirety, and the fourth column presents, where
applicable, the scores, probability values, and other parameters used to evaluate the strength of a
match between two sequences (the higher the score or the lower the probability value, the greater the
identity between two sequences).

The programs described above for the assembly and analysis of full length polynucleotide and polypeptide sequences are also used to identify polynucleotide sequence fragments from SEQ ID NO:39-76. Fragments from about 20 to about 4000 nucleotides which are useful in hybridization and amplification technologies are described in Table 4, column 2.

# 20 IV. Identification and Editing of Coding Sequences from Genomic DNA

Putative enzymes are initially identified by running the Genscan gene identification program against public genomic sequence databases (e.g., gbpri and gbhtg). Genscan is a general-purpose gene identification program which analyzes genomic DNA sequences from a variety of organisms (Burge, C. and S. Karlin (1997) J. Mol. Biol. 268:78-94; Burge, C. and S. Karlin (1998) Curr. Opin.

25 Struct. Biol. 8:346-354). The program concatenates predicted exons to form an assembled cDNA sequence extending from a methionine to a stop codon. The output of Genscan is a FASTA database of polynucleotide and polypeptide sequences. The maximum range of sequence for Genscan to analyze at once is set to 30 kb. To determine which of these Genscan predicted cDNA sequences encode enzymes, the encoded polypeptides are analyzed by querying against PFAM models for enzymes. Petential enzymes are also identified by homology to Incyte cDNA sequences that have been annotated as enzymes. These selected Genscan-predicted sequences are then compared by BLAST analysis to the genpept and gbpri public databases. Where necessary, the Genscan-predicted sequences are then edited by comparison to the top BLAST hit from genpept to correct errors in the

sequence predicted by Genscan, such as extra or omitted exons. BLAST analysis is also used to find any Incyte cDNA or public cDNA coverage of the Genscan-predicted sequences, thus providing evidence for transcription. When Incyte cDNA coverage is available, this information is used to correct or confirm the Genscan predicted sequence. Full length polynucleotide sequences are obtained by assembling Genscan-predicted coding sequences with Incyte cDNA sequences and/or public cDNA sequences using the assembly process described in Example III. Alternatively, full length polynucleotide sequences are derived entirely from edited or unedited Genscan-predicted coding sequences.

# V. Assembly of Genomic Sequence Data with cDNA Sequence Data

### 10 "Stitched" Sequences

Partial cDNA sequences are extended with exons predicted by the Genscan gene identification program described in Example IV. Partial cDNAs assembled as described in Example III are mapped to genomic DNA and parsed into clusters containing related cDNAs and Genscan exon predictions from one or more genomic sequences. Each cluster is analyzed using an algorithm 15 based on graph theory and dynamic programming to integrate cDNA and genomic information, generating possible splice variants that are subsequently confirmed, edited, or extended to create a full length sequence. Sequence intervals in which the entire length of the interval is present on more than one sequence in the cluster are identified, and intervals thus identified are considered to be equivalent by transitivity. For example, if an interval is present on a cDNA and two genomic sequences, then all three intervals are considered to be equivalent. This process allows unrelated but consecutive genomic sequences to be brought together, bridged by cDNA sequence. Intervals thus identified are then "stitched" together by the stitching algorithm in the order that they appear along their parent sequences to generate the longest possible sequence, as well as sequence variants. Linkages between intervals which proceed along one type of parent sequence (cDNA to cDNA or genomic sequence to 25 genomic sequence) are given preference over linkages which change parent type (cDNA to genomic sequence). The resultant stitched sequences are translated and compared by BLAST analysis to the genpept and gbpri public databases. Incorrect exons predicted by Genscan are corrected by comparison to the top BLAST hit from genpept. Sequences are further extended with additional cDNA sequences, or by inspection of genomic DNA, when necessary.

### 30 "Stretched" Sequences

Partial DNA sequences are extended to full length with an algorithm based on BLAST analysis. First, partial cDNAs assembled as described in Example III are queried against public databases such as the GenBank primate, rodent, mammalian, vertebrate, and eukaryote databases

using the BLAST program. The nearest GenBank protein homolog is then compared by BLAST analysis to either Incyte cDNA sequences or GenScan exon predicted sequences described in Example IV. A chimeric protein is generated by using the resultant high-scoring segment pairs (HSPs) to map the translated sequences onto the GenBank protein homolog. Insertions or deletions 5 may occur in the chimeric protein with respect to the original GenBank protein homolog. The GenBank protein homolog, the chimeric protein, or both are used as probes to search for homologous genomic sequences from the public human genome databases. Partial DNA sequences are therefore "stretched" or extended by the addition of homologous genomic sequences. The resultant stretched sequences are examined to determine whether they contain a complete gene.

#### Chromosomal Mapping of ENZM Encoding Polynucleotides VI. 10

The sequences used to assemble SEQ ID NO:39-76 are compared with sequences from the Incyte LIFESEQ database and public domain databases using BLAST and other implementations of the Smith-Waterman algorithm. Sequences from these databases that matched SEQ ID NO:39-76 are assembled into clusters of contiguous and overlapping sequences using assembly algorithms such 15 as Phrap (Table 7). Radiation hybrid and genetic mapping data available from public resources such as the Stanford Human Genome Center (SHGC), Whitehead Institute for Genome Research (WIGR), and Généthon are used to determine if any of the clustered sequences have been previously mapped. Inclusion of a mapped sequence in a cluster results in the assignment of all sequences of that cluster, including its particular SEQ ID NO:, to that map location.

Map locations are represented by ranges, or intervals, of human chromosomes. The map position of an interval, in centiMorgans, is measured relative to the terminus of the chromosome's parm. (The centiMorgan (cM) is a unit of measurement based on recombination frequencies between chromosomal markers. On average, 1 cM is roughly equivalent to 1 megabase (Mb) of DNA in humans, although this can vary widely due to hot and cold spots of recombination.) The cM distances 25 are based on genetic markers mapped by Généthon which provide boundaries for radiation hybrid markers whose sequences were included in each of the clusters. Human genome maps and other resources available to the public, such as the NCBI "GeneMap'99" World Wide Web site (http://www.ncbi.nlm.nih.gov/genemap/), can be employed to determine if previously identified disease genes map within or in proximity to the intervals indicated above.

# 30 VII. Analysis of Polynucleotide Expression

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Northern analysis is a laboratory technique used to detect the presence of a transcript of a gene and involves the hybridization of a labeled nucleotide sequence to a membrane on which RNAs from a particular cell type or tissue have been bound (Sambrook and Russell, supra, ch. 7; Ausubel et

al., supra, ch. 4).

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Analogous computer techniques applying BLAST are used to search for identical or related molecules in databases such as GenBank or LIFESEQ (Incyte). This analysis is much faster than multiple membrane-based hybridizations. In addition, the sensitivity of the computer search can be modified to determine whether any particular match is categorized as exact or similar. The basis of the search is the product score, which is defined as:

BLAST Score x Percent Identity

5 x minimum {length(Seq. 1), length(Seq. 2)}

The product score takes into account both the degree of similarity between two sequences and the length of the sequence match. The product score is a normalized value between 0 and 100, and is calculated as follows: the BLAST score is multiplied by the percent nucleotide identity and the product is divided by (5 times the length of the shorter of the two sequences). The BLAST score is calculated by assigning a score of +5 for every base that matches in a high-scoring segment pair (HSP), and -4 for every mismatch. Two sequences may share more than one HSP (separated by gaps). If there is more than one HSP, then the pair with the highest BLAST score is used to calculate the product score. The product score represents a balance between fractional overlap and quality in a BLAST alignment. For example, a product score of 100 is produced only for 100% identity over the entire length of the shorter of the two sequences being compared. A product score of 70 is produced either by 100% identity and 100% overlap at one end, or by 88% identity and 100% overlap at the other. A product score of 50 is produced either by 100% identity and 50% overlap at one end, or 79% identity and 100% overlap.

Alternatively, polynucleotides encoding ENZM are analyzed with respect to the tissue sources
from which they are derived. For example, some full length sequences are assembled, at least in part,
with overlapping Incyte cDNA sequences (see Example III). Each cDNA sequence is derived from
a cDNA library constructed from a human tissue. Each human tissue is classified into one of the
following organ/tissue categories: cardiovascular system; connective tissue; digestive system;
embryonic structures; endocrine system; exocrine glands; genitalia, female; genitalia, male; germ cells;
hemic and immune system; liver; musculoskeletal system; nervous system; pancreas; respiratory
system; sense organs; skin; stomatognathic system; unclassified/mixed; or urinary tract. The number
of libraries in each category is counted and divided by the total number of libraries across all
categories. Similarly, each human tissue is classified into one of the following disease/condition
categories: cancer, cell line, developmental, inflammation, neurological, trauma, cardiovascular, pooled,

and other, and the number of libraries in each category is counted and divided by the total number of libraries across all categories. The resulting percentages reflect the tissue- and disease-specific expression of cDNA encoding ENZM. cDNA sequences and cDNA library/tissue information are found in the LIFESEQ database (Incyte, Palo Alto CA).

## 5 VIII. Extension of ENZM Encoding Polynucleotides

Full length polynucleotides are produced by extension of an appropriate fragment of the full length molecule using oligonucleotide primers designed from this fragment. One primer is synthesized to initiate 5' extension of the known fragment, and the other primer is synthesized to initiate 3' extension of the known fragment. The initial primers are designed using OLIGO 4.06 software

(National Biosciences), or another appropriate program, to be about 22 to 30 nucleotides in length, to have a GC content of about 50% or more, and to anneal to the target sequence at temperatures of about 68°C to about 72°C. Any stretch of nucleotides which would result in hairpin structures and primer-primer dimerizations is avoided.

Selected human cDNA libraries are used to extend the sequence. If more than one extension is necessary or desired, additional or nested sets of primers are designed.

High fidelity amplification is obtained by PCR using methods well known in the art. PCR is performed in 96-well plates using the PTC-200 thermal cycler (MI Research, Inc.). The reaction mix contains DNA template, 200 nmol of each primer, reaction buffer containing Mg<sup>2+</sup>, (NH<sub>4)2</sub>SO<sub>4</sub>, and 2-mercaptoethanol, Taq DNA polymerase (Amersham Biosciences), ELONGASE enzyme (Invitrogen), 20 and Pfu DNA polymerase (Stratagene), with the following parameters for primer pair PCI A and PCI B: Step 1: 94°C, 3 min; Step 2: 94°C, 15 sec; Step 3: 60°C, 1 min; Step 4: 68°C, 2 min; Step 5: Steps 2, 3, and 4 repeated 20 times; Step 6: 68°C, 5 min; Step 7: storage at 4°C. In the alternative, the parameters for primer pair T7 and SK+ are as follows: Step 1: 94°C, 3 min; Step 2: 94°C, 15 sec; Step 3: 57°C, 1 min; Step 4: 68°C, 2 min; Step 5: Steps 2, 3, and 4 repeated 20 times; Step 6: 68°C, 5 min; Step 7: storage at 4°C.

The concentration of DNA in each well is determined by dispensing 100 µl PICOGREEN quantitation reagent (0.25% (v/v) PICOGREEN; Molecular Probes, Eugene OR) dissolved in 1X TE and 0.5 µl of undiluted PCR product into each well of an opaque fluorimeter plate (Corning Costar, Acton MA), allowing the DNA to bind to the reagent. The plate is scanned in a Fluoroskan II

(Labsystems Oy, Helsinki, Finland) to measure the fluorescence of the sample and to quantify the concentration of DNA. A 5 µl to 10 µl aliquot of the reaction mixture is analyzed by electrophoresis on a 1 % agarose gel to determine which reactions are successful in extending the sequence.

The extended nucleotides are desalted and concentrated, transferred to 384-well plates,

digested with CviJI cholera virus endonuclease (Molecular Biology Research, Madison WI), and sonicated or sheared prior to religation into pUC 18 vector (Amersham Biosciences). For shotgun sequencing, the digested nucleotides are separated on low concentration (0.6 to 0.8%) agarose gels, fragments are excised, and agar digested with Agar ACE (Promega). Extended clones were religated 5 using T4 ligase (New England Biolabs, Beverly MA) into pUC 18 vector (Amersham Biosciences), treated with Pfu DNA polymerase (Stratagene) to fill-in restriction site overhangs, and transfected into competent E. coli cells. Transformed cells are selected on antibiotic-containing media, and individual colonies are picked and cultured overnight at 37°C in 384-well plates in LB/2x carb liquid media.

The cells are lysed, and DNA is amplified by PCR using Taq DNA polymerase (Amersham Biosciences) and Pfu DNA polymerase (Stratagene) with the following parameters: Step 1: 94°C, 3 min; Step 2: 94°C, 15 sec; Step 3: 60°C, 1 min; Step 4: 72°C, 2 min; Step 5: steps 2, 3, and 4 repeated 29 times; Step 6: 72°C, 5 min; Step 7: storage at 4°C. DNA is quantified by PICOGREEN reagent (Molecular Probes) as described above. Samples with low DNA recoveries are reamplified using the 15 same conditions as described above. Samples are diluted with 20% dimethysulfoxide (1:2, v/v), and sequenced using DYENAMIC energy transfer sequencing primers and the DYENAMIC DIRECT kit (Amersham Biosciences) or the ABI PRISM BIGDYE Terminator cycle sequencing ready reaction kit (Applied Biosystems).

In like manner, full length polynucleotides are verified using the above procedure or are used 20 to obtain 5' regulatory sequences using the above procedure along with oligonucleotides designed for such extension, and an appropriate genomic library.

### Identification of Single Nucleotide Polymorphisms in ENZM Encoding IX. Polynucleotides

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Common DNA sequence variants known as single nucleotide polymorphisms (SNPs) are 25 identified in SEQ ID NO:39-76 using the LIFESEQ database (Incyte). Sequences from the same gene are clustered together and assembled as described in Example III, allowing the identification of all sequence variants in the gene. An algorithm consisting of a series of filters is used to distinguish SNPs from other sequence variants. Preliminary filters remove the majority of basecall errors by requiring a minimum Phred quality score of 15, and remove sequence alignment errors and errors resulting from improper trimming of vector sequences, chimeras, and splice variants. An automated procedure of advanced chromosome analysis is applied to the original chromatogram files in the vicinity of the putative SNP. Clone error filters use statistically generated algorithms to identify errors introduced during laboratory processing, such as those caused by reverse transcriptase, polymerase, or

somatic mutation. Clustering error filters use statistically generated algorithms to identify errors resulting from clustering of close homologs or pseudogenes, or due to contamination by non-human sequences. A final set of filters removes duplicates and SNPs found in immunoglobulins or T-cell receptors.

Certain SNPs are selected for further characterization by mass spectrometry using the high throughput MASSARRAY system (Sequenom, Inc.) to analyze allele frequencies at the SNP sites in four different human populations. The Caucasian population comprises 92 individuals (46 male, 46 female), including 83 from Utah, four French, three Venezualan, and two Amish individuals. The African population comprises 194 individuals (97 male, 97 female), all African Americans. The 10 Hispanic population comprises 324 individuals (162 male, 162 female), all Mexican Hispanic. The Asian population comprises 126 individuals (64 male, 62 female) with a reported parental breakdown of 43% Chinese, 31% Japanese, 13% Korean, 5% Victnamese, and 8% other Asian. Allele frequencies are first analyzed in the Caucasian population; in some cases those SNPs which show no allelic variance in this population are not further tested in the other three populations.

#### Labeling and Use of Individual Hybridization Probes 15 X.

Hybridization probes derived from SEO ID NO:39-76 are employed to screen cDNAs, genomic DNAs, or mRNAs. Although the labeling of oligonucleotides, consisting of about 20 base pairs, is specifically described, essentially the same procedure is used with larger nucleotide fragments. Oligonucleotides are designed using state-of-the-art software such as OLIGO 4.06 20 software (National Biosciences) and labeled by combining 50 pmol of each oligomer, 250 μCi of [y-32P] adenosine triphosphate (Amersham Biosciences), and T4 polynucleotide kinase (DuPont NEN, Boston MA). The labeled oligonucleotides are substantially purified using a SEPHADEX G-25 superfine size exclusion dextran bead column (Amersham Biosciences). An aliquot containing 107 counts per minute of the labeled probe is used in a typical membrane-based hybridization analysis of 25 human genomic DNA digested with one of the following endonucleases: Ase I, Bgl II, Eco RI, Pst I, Xba I, or Pvu II (DuPont NEN).

The DNA from each digest is fractionated on a 0.7% agarose gel and transferred to nylon membranes (Nytran Plus, Schleicher & Schuell, Durham NH). Hybridization is carried out for 16 hours at 40°C. To remove nonspecific signals, blots are sequentially washed at room temperature 30 under conditions of up to, for example, 0.1 x saline sodium citrate and 0.5% sodium dodecyl sulfate. Hybridization patterns are visualized using autoradiography or an alternative imaging means and compared.

#### XI. Microarrays

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The linkage or synthesis of array elements upon a microarray can be achieved utilizing photolithography, piezoelectric printing (ink-jet printing; see, e.g., Baldeschweiler et al., supra), mechanical microspotting technologies, and derivatives thereof. The substrate in each of the aforementioned technologies should be uniform and solid with a non-porous surface (Schena, M., ed. 5 (1999) DNA Microarrays: A Practical Approach, Oxford University Press, London). Suggested substrates include silicon, silica, glass slides, glass chips, and silicon wafers. Alternatively, a procedure analogous to a dot or slot blot may also be used to arrange and link elements to the surface of a substrate using thermal, UV, chemical, or mechanical bonding procedures. A typical array may be produced using available methods and machines well known to those of ordinary skill in the art and may contain any appropriate number of elements (Schena, M. et al. (1995) Science 270:467-470; Shalon, D. et al. (1996) Genome Res. 6:639-645; Marshall, A. and J. Hodgson (1998) Nat. Biotechnol. 16:27-31).

Full length cDNAs, Expressed Sequence Tags (ESTs), or fragments or oligomers thereof may comprise the elements of the microarray. Fragments or oligomers suitable for hybridization can be 15 selected using software well known in the art such as LASERGENE software (DNASTAR). The array elements are hybridized with polynucleotides in a biological sample. The polynucleotides in the biological sample are conjugated to a fluorescent label or other molecular tag for ease of detection. After hybridization, nonhybridized nucleotides from the biological sample are removed, and a fluorescence scanner is used to detect hybridization at each array element. Alternatively, laser desorbtion and mass spectrometry may be used for detection of hybridization. The degree of complementarity and the relative abundance of each polynucleotide which hybridizes to an element on the microarray may be assessed. In one embodiment, microarray preparation and usage is described in detail below.

### Tissue or Cell Sample Preparation

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Total RNA is isolated from tissue samples using the guanidinium thiocyanate method and poly(A)+ RNA is purified using the oligo-(dT) cellulose method. Each poly(A)+ RNA sample is reverse transcribed using MMLV reverse-transcriptase, 0.05 pg/µl oligo-(dT) primer (21mer), 1X first strand buffer, 0.03 units/µl RNase inhibitor, 500 µM dATP, 500 µM dGTP, 500 µM dTTP, 40 µM dCTP, 40 μM dCTP-Cy3 (BDS) or dCTP-Cy5 (Amersham Biosciences). The reverse transcription 30 reaction is performed in a 25 ml volume containing 200 ng poly(A)+ RNA with GEMBRIGHT kits (Incyte). Specific control poly(A)+ RNAs are synthesized by in vitro transcription from non-coding yeast genomic DNA. After incubation at 37°C for 2 hr, each reaction sample (one with Cy3 and another with Cy5 labeling) is treated with 2.5 ml of 0.5M sodium hydroxide and incubated for 20

minutes at 85°C to the stop the reaction and degrade the RNA. Samples are purified using two successive CHROMA SPIN 30 gel filtration spin columns (BD Clontech, Palo Alto CA) and after combining, both reaction samples are ethanol precipitated using 1 ml of glycogen (1 mg/ml), 60 ml sodium acetate, and 300 ml of 100% ethanol. The sample is then dried to completion using a SpeedVAC (Savant Instruments Inc., Holbrook NY) and resuspended in 14 µl 5X SSC/0.2% SDS.

### Microarray Preparation

Sequences of the present invention are used to generate array elements. Each array element is amplified from bacterial cells containing vectors with cloned cDNA inserts. PCR amplification uses primers complementary to the vector sequences flanking the cDNA insert. Array elements are amplified in thirty cycles of PCR from an initial quantity of 1-2 ng to a final quantity greater than 5 μg. Amplified array elements are then purified using SEPHACRYL-400 (Amersham Biosciences).

Purified array elements are immobilized on polymer-coated glass slides. Glass microscope slides (Corning) are cleaned by ultrasound in 0.1% SDS and acetone, with extensive distilled water washes between and after treatments. Glass slides are etched in 4% hydrofluoric acid (VWR)

15 Scientific Products Corporation (VWR), West Chester PA), washed extensively in distilled water, and coated with 0.05% aminopropyl silane (Sigma-Aldrich, St. Louis MO) in 95% ethanol. Coated slides are cured in a 110°C oven.

Array elements are applied to the coated glass substrate using a procedure described in U.S.

Patent No. 5,807,522, incorporated herein by reference. 1  $\mu$ l of the array element DNA, at an average

concentration of 100 ng/ $\mu$ l, is loaded into the open capillary printing element by a high-speed robotic

apparatus. The apparatus then deposits about 5 nl of array element sample per slide.

Microarrays are UV-crosslinked using a STRATALINKER UV-crosslinker (Stratagene).

Microarrays are washed at room temperature once in 0.2% SDS and three times in distilled water.

Non-specific binding sites are blocked by incubation of microarrays in 0.2% casein in phosphate

buffered saline (PBS) (Tropix, Inc., Bedford MA) for 30 minutes at 60°C followed by washes in 0.2%

SDS and distilled water as before.

### Hybridization

Hybridization reactions contain 9  $\mu$ l of sample mixture consisting of 0.2  $\mu$ g each of Cy3 and Cy5 labeled cDNA synthesis products in 5X SSC, 0.2% SDS hybridization buffer. The sample mixture is heated to 65°C for 5 minutes and is aliquoted onto the microarray surface and covered with an 1.8 cm² coverslip. The arrays are transferred to a waterproof chamber having a cavity just slightly larger than a microscope slide. The chamber is kept at 100% humidity internally by the addition of 140  $\mu$ l of 5X SSC in a corner of the chamber. The chamber containing the arrays is incubated for about

6.5 hours at 60°C. The arrays are washed for 10 min at 45°C in a first wash buffer (1X SSC, 0.1% SDS), three times for 10 minutes each at 45°C in a second wash buffer (0.1X SSC), and dried.
Detection

Reporter-labeled hybridization complexes are detected with a microscope equipped with an

Innova 70 mixed gas 10 W laser (Coherent, Inc., Santa Clara CA) capable of generating spectral lines
at 488 nm for excitation of Cy3 and at 632 nm for excitation of Cy5. The excitation laser light is
focused on the array using a 20X microscope objective (Nikon, Inc., Melville NY). The slide
containing the array is placed on a computer-controlled X-Y stage on the microscope and rasterscanned past the objective. The 1.8 cm x 1.8 cm array used in the present example is scanned with a

resolution of 20 micrometers.

In two separate scans, a mixed gas multiline laser excites the two fluorophores sequentially.

Emitted light is split, based on wavelength, into two photomultiplier tube detectors (PMT R1477,

Hamamatsu Photonics Systems, Bridgewater NJ) corresponding to the two fluorophores. Appropriate
filters positioned between the array and the photomultiplier tubes are used to filter the signals. The
emission maxima of the fluorophores used are 565 nm for Cy3 and 650 nm for Cy5. Each array is
typically scanned twice, one scan per fluorophore using the appropriate filters at the laser source,
although the apparatus is capable of recording the spectra from both fluorophores simultaneously.

The sensitivity of the scans is typically calibrated using the signal intensity generated by a cDNA control species added to the sample mixture at a known concentration. A specific location on the array contains a complementary DNA sequence, allowing the intensity of the signal at that location to be correlated with a weight ratio of hybridizing species of 1:100,000. When two samples from different sources (e.g., representing test and control cells), each labeled with a different fluorophore, are hybridized to a single array for the purpose of identifying genes that are differentially expressed, the calibration is done by labeling samples of the calibrating cDNA with the two fluorophores and adding identical amounts of each to the hybridization mixture.

The output of the photomultiplier tube is digitized using a 12-bit RTI-835H analog-to-digital (A/D) conversion board (Analog Devices, Inc., Norwood MA) installed in an IBM-compatible PC computer. The digitized data are displayed as an image where the signal intensity is mapped using a linear 20-color transformation to a pseudocolor scale ranging from blue (low signal) to red (high signal). The data is also analyzed quantitatively. Where two different fluorophores are excited and measured simultaneously, the data are first corrected for optical crosstalk (due to overlapping emission spectral) between the fluorophores using each fluorophore's emission spectrum.

A grid is superimposed over the fluorescence signal image such that the signal from each spot

is centered in each element of the grid. The fluorescence signal within each element is then integrated to obtain a numerical value corresponding to the average intensity of the signal. The software used for signal analysis is the GEMTOOLS gene expression analysis program (Incyte). Array elements that exhibit at least about a two-fold change in expression, a signal-to-background ratio of at least about 2.5, and an element spot size of at least about 40%, are considered to be differentially expressed.

### Expression

SEQ ID NO:49 was downregulated in breast cancer tissue versus normal breast tissue as
determined by microarray analysis. Expression of SEQ ID NO:49 was decreased in diseased tissue
10 as compared with normal tissue from the same donor. Therefore, SEQ ID NO:49 can be used in
monitoring treatment of, and diagnostic assays for, breast cancer.

SEQ ID NO:39 and SEQ ID NO:54 were downregulated in colon cancer tissue versus normal colon tissue as determined by microarray analysis. Expression of SEQ ID NO:39 and SEQ ID NO:54 was decreased in comparison of normal tissue from a donor with diseased tissue from the same donor.

15 Therefore, SEQ ID NO:39 and SEQ ID NO:54 can be used in monitoring treatment of, and diagnostic assays for, colon cancer.

SEQ ID NO:48 and SEQ ID NO:51 were upregulated in prostate cancer cells versus normal prostate epithelial cells as determined by microarray analysis. Expression profiles of the prostate carcinoma lines LNCaP, PC-3, and DU 145 were compared to the expression profile of prostate epithelial cells (PrECs). DU 145 is a prostate carcinoma cell line isolated from a metastatic site in the brain of a donor with widespread metastatic prostate carcinoma. LNCaP is a prostate carcinoma cell line isolated from a lymph node biopsy of donor with metastatic prostate carcinoma. PC-3 is a prostate adenocarcinoma cell line that was isolated from a metastatic site in the bone of a donor with grade IV prostate adenocarcinoma. Expression of SEQ ID NO:48 was increased in all three cell lines tested. Expression of SEQ ID NO:51 was increased in DU 145 and LNCaP cells. Therefore, SEQ ID NO:48 and SEQ ID NO:51 can be used in monitoring treatment of, and diagnostic assays for, prostate cancer.

SEQ ID NO:49, SEQ ID NO:54, SEQ ID NO:55, and SEQ ID NO:56 were differentially regulated in lung cancer tissue versus normal lung tissue as determined by microarray analysis.

30 Expression of SEQ ID NO:49 and SEQ ID NO:54 was decreased in diseased tissue as compared with normal tissue from the same donor. Expression of SEQ ID NO:55 and SEQ ID NO:56 was increased in diseased tissue as compared with normal tissue from the same donor. Therefore, SEQ ID NO:49, SEQ ID NO:54, SEQ ID NO:55, and SEQ ID NO:56 can be used in monitoring treatment of, and

diagnostic assays for, lung cancer.

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SEQ ID NO:49 was downregulated in ovarian cancer tissue versus normal ovarian tissue as determined by microarray analysis. Expression of SEQ ID NO:49 was decreased in comparisons of normal and diseased tissue from the same donor. Therefore, SEQ ID NO:49 can be used in 5 monitoring treatment of, and diagnostic assays for, ovarian cancer.

SEQ ID NO:42 and SEQ ID NO:46 were differentially regulated in brain tissue associated with Alzheimer's disease (AD) versus normal brain tissue as determined by microarray analysis. In one experiment, specific dissected brain regions from the brain of a female with severe AD were compared to dissected regions from a normal female and two normal male brains. The diagnosis of 10 normal or severe AD was established by a certified neuropathologist based on microscopic examination of multiple sections throughout the brain. Expression of SEQ ID NO:42 was decreased in tissue affected by severe AD as compared with normal brain tissue. In a second experiment, specific dissected brain regions from the brain of a female with mild AD were compared to from a normal donor were compared to specific dissected brain regions from two normal male and a normal female 15 donor. The diagnosis of normal or mild AD was established by a certified neuropathologist based on microscopic examination of multiple sections throughout the brain. Expression of SEQ ID NO:46 was increased in tissue affected by mild AD compared with normal brain tissue. Therefore, SEQ ID NO:42 and SEQ ID NO:46 can be used in monitoring treatment of, and diagnostic assays for, Alzheimer's disease.

SEQ ID NO:55 and SEQ ID NO:56 were downregulated in tissue associated with Tangier disease versus normal tissue as determined by microarray analysis. Human fibroblasts were obtained from skin explants from both normal subjects and two patients with homozygous Tangier disease. Cell lines were immortalized by transfection with human papillomavirus 16 genes E6 and E7 and a neomycin resistance selectable marker. In addition, both types of cells were cultured in the presence 25 of cholesterol and compared with the same cell type cultured in the absence of cholesterol. TD derived cells are shown to be deficient in an assay of apoA-I mediated tritiated cholesterol efflux. Expression of SEQ ID NO:55 and SEQ ID NO:56 was decreased in fibroblasts from both patients with Tangier disease. Therefore, SEQ ID NO:55 and SEQ ID NO:56 can be used in monitoring treatment of, and diagnostic assays for, Tangier disease.

SEQ ID NO:45 were differentially regulated in C3A cells treated with gemfibrozil versus untreated C3A cells, as determined by microarray analysis. Early confluent C3A cells were treated with various amounts of Gemfibrozil (120, 600, 800, and 1200 µg/ml) dissolved in CMC for 1, 3, and 6 hours. Parallel samples of C3A cells were treated with 1% CMC only, as a control. Expression of

SEQ ID NO:45 was increased in C3A cells treated with gemfibrozil. Therefore, SEQ ID NO:45 can be used in monitoring treatment of, and diagnostic assays for, metabolic, cardiovascular, and liver disorders.

SEQ ID NO:55 and SEQ ID NO:56 were upregulated in C3A cells treated with a variety of

steroids versus untreated C3A cells, as determined by microarray analysis. Early confluent C3A cells
were treated for 1, 3, and 6 hours at concentrations of 1, 10, and 100 µM. Treated cells were
compared to untreated early confluent C3A cells. Expression of SEQ ID NO:55 and SEQ ID NO:56
was increased in C3A cells treated with progesterone, beclomethasone, medroxyprogesterone,
budesonide, prednisone, dexamethasone, and betamethasone. Therefore, SEQ ID NO:55 and SEQ ID

NO:56 can be used for the diagnosis and monitoring of liver, endocrine, and reproductive diseases and
in the diagnosis of and as a therapeutic target for inflammatory diseases and humoral immune
response.

In another example, SEQ ID NO:60 showed differential expression in prostate cancer cells versus normal prostate epithelial cells, as determined by microarray analysis. Primary prostate epithelial cells were compared with prostate carcinomas representative of the different stages of tumor progression. Cell lines compared included: a)PrEC, a primary prostate epithelial cell line isolated from a normal donor, b) DU 145, a prostate carcinoma cell line isolated from a metastatic site in the brain of a donor with widespread metastatic prostate carcinoma, c) LNCaP, a prostate carcinoma cell line isolated from a lymph node biopsy of a donor with metastatic prostate carcinoma, and d) PC-3, a prostate adenocarcinoma cell line isolated from a metastatic site in the bone of a donor with grade IV prostate adenocarcinoma. Cells grown under restrictive conditions were compared to normal PrECs grown under restrictive conditions. Expression of SEQ ID NO:60 was increased in all LNCaP samples and in 3 of 4 DU 145 samples tested. Therefore, SEQ ID NO:60 can be used in monitoring treatment of, and diagnostic assays for, prostate cancer.

In a further example, SEQ ID NO:75 showed differential expression in fibroblasts with Tangier disease versus normal fibroblasts, as determined by microarray analysis. Human fibroblasts were obtained from skin explants from both normal subjects and two patients with homozygous Tangier disease. Cell lines were immortalized by transfection with human papillomavirus 16 genes E6 and E7 and a neomycin resistance selectable marker. In addition, both types of cells were cultured in the presence of cholesterol and compared with the same cell type cultured in the absence of cholesterol. TD-derived cells are deficient in an assay of apoA-I mediated tritiated cholesterol efflux. Expression of SEQ ID NO:75 was increased by an average of more than twofold in fibroblasts obtained from donors with Tangier disease. Therefore, SEQ ID NO:75 can be used in diagnostic

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assays for Tangier disease.

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In addition, SEQ ID NO:39, SEQ ID NO:42, SEQ ID NO:44, SEQ ID NO:45, SEQ ID NO:49, SEQ ID NO:55, SEQ ID NO:55, SEQ ID NO:56, SEQ ID NO:61, and SEQ ID NO:69 showed tissue-specific expression. RNA samples isolated from a variety of normal human tissues were compared to a common reference sample. Tissues contributing to the reference sample were selected for their ability to provide a complete distribution of RNA in the human body and include brain (4%), heart (7%), kidney (3%), lung (8%), placenta (46%), small intestine (9%), spleen (3%), stomach (6%), testis (9%), and uterus (5%). The normal tissues assayed were obtained from at least three different donors. RNA from each donor was separately isolated and individually hybridized to the microarray. Since these hybridization experiments were conducted using a common reference sample, differential expression values are directly comparable from one tissue to another.

The expression of SEQ ID NO:39 was increased by at least two-fold in occipital cortex and in temporal cortex brain tissue, as compared to the reference sample. Therefore, SEQ ID NO:39 can be used as a tissue marker for brain.

The expression of SEQ ID NO:44, SEQ ID NO:45, SEQ ID NO:49, SEQ ID NO:54, SEQ ID NO:55, and SEQ ID NO:56 was increased by at least two-fold in liver as compared to the reference sample. Therefore, SEQ ID NO:44, SEQ ID NO:45, SEQ ID NO:49, SEQ ID NO:54, SEQ ID NO:55, and SEQ ID NO:56 can be used as a tissue marker for liver.

The expression of SEQ ID NO:54 and SEQ ID NO:45 was increased by at least two-fold in
kidney as compared to the reference sample. Therefore, SEQ ID NO:54 and SEQ ID NO:45 can be used as a tissue marker for kidney.

The expression of SEQ ID NO:54 was increased by at least two-fold in duodenum as compared to the reference sample. Therefore, SEQ ID NO:54 can be used as a tissue marker for duodenum.

The expression of SEQ ID NO:55 and SEQ ID NO:56 was increased by at least two-fold in psoas skeletal muscle as compared to the reference sample. Therefore, SEQ ID NO:55 and SEQ ID NO:56 can be used as a tissue marker for psoas skeletal muscle.

The expression of SEQ ID NO:42 was increased by at least two-fold in brain as compared to the reference sample. Therefore, SEQ ID NO:42 can be used as a tissue marker for brain.

The expression of SEQ ID NO:55 and SEQ ID NO:56 was increased by at least two-fold in left ventricle and right ventricle as compared to the reference sample. Therefore, SEQ ID NO:55 and SEQ ID NO:56 can be used as a tissue marker for left ventricle and right ventricle.

The expression of SEQ ID NO:61 was increased by at least two-fold in right ventricle tissue

and adrenal tissue, as compared to the reference sample. Therefore, SEQ ID NO:61 can be used as a tissue marker for right ventricle tissue and adrenal tissue.

The expression of SEQ ID NO:69 was increased by at least two-fold in liver and kidney tissue, as compared to the reference sample. Therefore, SEQ ID NO:69 can be used as a tissue

marker for liver and kidney tissue.

#### XII. Complementary Polynucleotides

Sequences complementary to the ENZM-encoding sequences, or any parts thereof, are used to detect, decrease, or inhibit expression of naturally occurring ENZM. Although use of oligonucleotides comprising from about 15 to 30 base pairs is described, essentially the same 10 procedure is used with smaller or with larger sequence fragments. Appropriate oligonucleotides are designed using OLIGO 4.06 software (National Biosciences) and the coding sequence of ENZM. To inhibit transcription, a complementary oligonucleotide is designed from the most unique 5' sequence and used to prevent promoter binding to the coding sequence. To inhibit translation, a complementary oligonucleotide is designed to prevent ribosomal binding to the ENZM-encoding transcript.

#### 15 XIII. Expression of ENZM

Expression and purification of ENZM is achieved using bacterial or virus-based expression systems. For expression of ENZM in bacteria, cDNA is subcloned into an appropriate vector containing an antibiotic resistance gene and an inducible promoter that directs high levels of cDNA transcription. Examples of such promoters include, but are not limited to, the trp-lac (tac) hybrid 20 promoter and the T5 or T7 bacteriophage promoter in conjunction with the lac operator regulatory element. Recombinant vectors are transformed into suitable bacterial hosts, e.g., BL21(DE3). Antibiotic resistant bacteria express ENZM upon induction with isopropyl beta-Dthiogalactopyranoside (IPTG). Expression of ENZM in eukaryotic cells is achieved by infecting insect or mammalian cell lines with recombinant Autographica californica nuclear polyhedrosis virus 25 (AcMNPV), commonly known as baculovirus. The nonessential polyhedrin gene of baculovirus is replaced with cDNA encoding ENZM by either homologous recombination or bacterial-mediated transposition involving transfer plasmid intermediates. Viral infectivity is maintained and the strong polyhedrin promoter drives high levels of cDNA transcription. Recombinant baculovirus is used to infect Spodoptera frugiperda (Sf9) insect cells in most cases, or human hepatocytes, in some cases. 30 Infection of the latter requires additional genetic modifications to baculovirus (Engelhard, E.K. et al. (1994) Proc. Natl. Acad. Sci. USA 91:3224-3227; Sandig, V, et al. (1996) Hum. Gene Ther. 7:1937-1945).

In most expression systems, ENZM is synthesized as a fusion protein with, e.g., glutathione S-

transferase (GST) or a peptide epitope tag, such as FLAG or 6-His, permitting rapid, single-step,
affinity-based purification of recombinant fusion protein from crude cell lysates. GST, a 26-kilodalton
enzyme from Schistosoma japonicum, enables the purification of fusion proteins on immobilized
glutathione under conditions that maintain protein activity and antigenicity (Amersham Biosciences).

5 Following purification, the GST moiety can be proteolytically cleaved from ENZM at specifically
engineered sites. FLAG, an 8-amino acid peptide, enables immunoaffinity purification using
commercially available monoclonal and polyclonal anti-FLAG antibodies (Eastman Kodak). 6-His, a
stretch of six consecutive histidine residues, enables purification on metal-chelate resins (QIAGEN).
Methods for protein expression and purification are discussed in Ausubel et al. (supra, ch. 10 and 16).

10 Purified ENZM obtained by these methods can be used directly in the assays shown in Examples
XVII, XVIII, and XIX, where applicable.

#### XIV. Functional Assays

ENZM function is assessed by expressing the sequences encoding ENZM at physiologically elevated levels in mammalian cell culture systems. cDNA is subcloned into a mammalian expression 15 vector containing a strong promoter that drives high levels of cDNA expression. Vectors of choice include PCMV SPORT plasmid (Invitrogen, Carlsbad CA) and PCR3.1 plasmid (Invitrogen), both of which contain the cytomegalovirus promoter. 5-10  $\mu g$  of recombinant vector are transiently transfected into a human cell line, for example, an endothelial or hematopoietic cell line, using either liposome formulations or electroporation. 1-2  $\mu g$  of an additional plasmid containing sequences 20 encoding a marker protein are co-transfected. Expression of a marker protein provides a means to distinguish transfected cells from nontransfected cells and is a reliable predictor of cDNA expression from the recombinant vector. Marker proteins of choice include, e.g., Green Fluorescent Protein (GFP; BD Clontech), CD64, or a CD64-GFP fusion protein. Flow cytometry (FCM), an automated, laser optics-based technique, is used to identify transfected cells expressing GFP or CD64-GFP and to 25 evaluate the apoptotic state of the cells and other cellular properties. FCM detects and quantifies the uptake of fluorescent molecules that diagnose events preceding or coincident with cell death. These events include changes in nuclear DNA content as measured by staining of DNA with propidium iodide; changes in cell size and granularity as measured by forward light scatter and 90 degree side light scatter; down-regulation of DNA synthesis as measured by decrease in bromodeoxyuridine 30 uptake; alterations in expression of cell surface and intracellular proteins as measured by reactivity with specific antibodies; and alterations in plasma membrane composition as measured by the binding of fluorescein-conjugated Annexin V protein to the cell surface. Methods in flow cytometry are discussed in Ormerod, M.G. (1994; Flow Cytometry, Oxford, New York NY).

The influence of ENZM on gene expression can be assessed using highly purified populations of cells transfected with sequences encoding ENZM and either CD64 or CD64-GFP. CD64 and CD64-GFP are expressed on the surface of transfected cells and bind to conserved regions of human immunoglobulin G (IgG). Transfected cells are efficiently separated from nontransfected cells using magnetic beads coated with either human IgG or antibody against CD64 (DYNAL, Lake Success NY). mRNA can be purified from the cells using methods well known by those of skill in the art. Expression of mRNA encoding ENZM and other genes of interest can be analyzed by northern analysis or microarray techniques.

# XV. Production of ENZM Specific Antibodies

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ENZM substantially purified using polyacrylamide gel electrophoresis (PAGE; see, e.g., Harrington, M.G. (1990) Methods Enzymol. 182:488-495), or other purification techniques, is used to immunize animals (e.g., rabbits, mice, etc.) and to produce antibodies using standard protocols.

Alternatively, the ENZM amino acid sequence is analyzed using LASERGENE software
(DNASTAR) to determine regions of high immunogenicity, and a corresponding oligopeptide is
synthesized and used to raise antibodies by means known to those of skill in the art. Methods for
selection of appropriate epitopes, such as those near the C-terminus or in hydrophilic regions are well
described in the art (Ausubel et al., supra, ch. 11).

Typically, oligopeptides of about 15 residues in length are synthesized using an ABI 431A peptide synthesizer (Applied Biosystems) using FMOC chemistry and coupled to KLH (Sigma-20 Aldrich, St. Louis MO) by reaction with N-maleimidobenzoyl-N-hydroxysuccinimide ester (MBS) to increase immunogenicity (Ausubel et al., supra). Rabbits are immunized with the oligopeptide-KLH complex in complete Freund's adjuvant. Resulting antisera are tested for antipeptide and anti-ENZM activity by, for example, binding the peptide or ENZM to a substrate, blocking with 1% BSA, reacting with rabbit antisera, washing, and reacting with radio-iodinated goat anti-rabbit IgG.

# 25 XVI. Purification of Naturally Occurring ENZM Using Specific Antibodies

Naturally occurring or recombinant ENZM is substantially purified by immunoaffinity chromatography using antibodies specific for ENZM. An immunoaffinity column is constructed by covalently coupling anti-ENZM antibody to an activated chromatographic resin, such as CNBr-activated SEPHAROSE (Amersham Biosciences). After the coupling, the resin is blocked and washed according to the manufacturer's instructions.

Media containing ENZM are passed over the immunoaffinity column, and the column is washed under conditions that allow the preferential absorbance of ENZM (e.g., high ionic strength buffers in the presence of detergent). The column is eluted under conditions that disrupt

antibody/ENZM binding (e.g., a buffer of pH 2 to pH 3, or a high concentration of a chaotrope, such as urea or thiocyanate ion), and ENZM is collected.

# XVII. Identification of Molecules Which Interact with ENZM

ENZM, or biologically active fragments thereof, are labeled with <sup>135</sup>I Bolton-Hunter reagent

(Bolton, A.E. and W.M. Hunter (1973) Biochem. J. 133:529-539). Candidate molecules previously arrayed in the wells of a multi-well plate are incubated with the labeled ENZM, washed, and any wells with labeled ENZM complex are assayed. Data obtained using different concentrations of ENZM are used to calculate values for the number, affinity, and association of ENZM with the candidate molecules.

Alternatively, molecules interacting with ENZM are analyzed using the yeast two-hybrid system as described in Fields, S. and O. Song (1989; Nature 340:245-246), or using commercially available kits based on the two-hybrid system, such as the MATCHMAKER system (BD Clontech).

ENZM may also be used in the PATHCALLING process (CuraGen Corp., New Haven CT)
which employs the yeast two-hybrid system in a high-throughput manner to determine all interactions
between the proteins encoded by two large libraries of genes (Nandabalan, K. et al. (2000) U.S.
Patent No. 6,057,101).

## XVIII. Demonstration of ENZM Activity

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ENZM activity is demonstrated through a variety of specific enzyme assays; some of which

20 are outlined below.

ENZM oxidoreductase activity is measured by the increase in extinction coefficient of NAD(P)H coenzyme at 340 nm for the measurement of oxidation activity, or the decrease in extinction coefficient of NAD(P)H coenzyme at 340 nm for the measurement of reduction activity (Dalziel, K. (1963) J. Biol. Chem. 238:2850-2858). One of three substrates may be used: Asn-βGal, biocytidine, or ubiquinone-10. The respective subunits of the enzyme reaction, for example, cytochrome e<sub>1</sub>-b oxidoreductase and cytochrome c, are reconstituted. The reaction mixture contains a)1-2 mg/ml ENZM; and b) 15 mM substrate, 2.4 mM NAD(P)\* in 0.1 M phosphate buffer, pH 7.1 (oxidation reaction), or 2.0 mM NAD(P)H, in 0.1 M Na<sub>2</sub>HPO<sub>4</sub> buffer, pH 7.4 (reduction reaction); in a total volume of 0.1 ml. Changes in absorbance at 340 nm (A<sub>340</sub>) are measured at 23.5 °C using a recording spectrophotometer (Shimadzu Scientific Instruments, Inc., Pleasanton, CA). The amount of NAD(P)H is stoichiometrically equivalent to the amount of substrate initially present, and the change in A<sub>340</sub> is a direct measure of the amount of NAD(P)H produced; ΔA<sub>340</sub> = 6620[NADH]. ENZM activity is proportional to the amount of NAD(P)H prosent in the assay.

Aldo/keto reductase activity of ENZM is proportional to the decrease in absorbance at 340 nm as NADPH is consumed (or increased absorbance if NADPH is produced, i.e., if the reverse reaction is monitored). A standard reaction mixture is 135 mM sodium phosphate buffer (pH 6.2-7.2 depending on enzyme), 0.2 mM NADPH, 0.3 M lithium sulfate, 0.5-2.5 mg ENZM and an appropriate level of substrate. The reaction is incubated at 30°C and the reaction is monitored continuously with a spectrophotometer. ENZM activity is calculated as mol NADPH consumed/mg of ENZM.

Acyl-CoA dehydrogenase activity of ENZM is measured using an anaerobic electron transferring flavoprotein (ETF) assay. The reaction mixture comprises 50 mM Tris-HCl (pH 8.0), 0.5% glucose, and 50 μM acyl-CoA substrate (i.e., isovaleryl-CoA) that is pre-warmed to 32 °C. The mixture is depleted of oxygen by repeated exposure to vacuum followed by layering with argon. Trace amounts of oxygen are removed by the addition of glucose oxidase and catalase followed by the addition of ETF to a final concentration of 1 μM. The reaction is initiated by addition of purified ENZM or a sample containing ENZM and exciting the reaction at 342 nm. Quenching of fluorescence caused by the transfer of electrons from the substrate to ETF is monitored at 496 nm. 1 unit of acyl-CoA dehydrogenase activity is defined as the amount of ENZM required to reduce 1 μmol of ETF per minute (Reinard, T. et al. (2000) J. Biol. Chem. 275:33738-33743).

Alcohol dehydrogenase activity of ENZM is measured by following the conversion of NAD+ to NADH at 340 nm (e<sub>340</sub> = 6.22 mM<sup>+</sup> cm<sup>-1</sup>) at 25°C in 0.1 M potassium phosphate (pH 7.5), 0.1 M glycine (pH 10.0), and 2.4 mM NAD+. Substrate (e.g., ethanol) and ENZM are then added to the reaction. The production of NADH results in an increase in absorbance at 340 nm and correlates with the oxidation of the alcohol substrate and the amount of alcohol dehydrogenase activity in the ENZM sample (Svensson, S. (1999) J. Biol. Chem. 274:29712-29719).

Aldehyde dehydrogenase activity of ENZM is measured by determining the total hydrolase + dehydrogenase activity of ENZM and subtracting the hydrolase activity. Hydrolase activity is first determined in a reaction mixture containing 0.05 M Tris-HCl (pH 7.8), 100 mM 2-mercaptoethanol, and 0.5-18 

µM substrate, e.g., 10-HCO-HPteGlu (10-formyletrahydrofolate; HPteGlu, tetrahydrofolate) or 10-FDDP (10-formyl-5,8-dideazafolate). Approximately 1 

µg of ENZM is added in a final volume of 1.0 ml. The reaction is monitored and read against a blank cuvette, containing all components except enzyme. The appearance of product is measured at either 295 nm for 5,8-dideazafolate or 300 nm for HPteGlu using molar extinction coefficients of 1.89x10<sup>4</sup> and 2.17x10<sup>4</sup> for 5,8-dideazafolate and HPteGlu, respectively. The addition of NADP<sup>+</sup> to the reaction mixture allows the measurement of both dehydrogenase and hydrolase activity (assays are performed as before). Based on the production of product in the presence of NADP<sup>+</sup> and the production of product

in the absence of the cofactor, aldehyde dehydrogenase activity is calculated for ENZM. In the alternative, aldehyde dehydrogenase activity is assayed using propanal as substrate. The reaction mixture contains 60 mM sodium pyrophosphate buffer (pH 8.5), 5 mM propanal, 1 mM NADP\*, and ENZM in a total volume of 1 ml. Activity is determined by the increase in absorbance at 340 nm, resulting from the generation of NADPH, and is proportional to the aldehyde dehydrogenase activity in the sample (Krupenko, S.A. et al. (1995) J. Biol. Chem. 270:519-522).

6-phosphogluconate dehydrogenase activity of ENZM is measured by incubating purified ENZM, or a composition comprising ENZM, in 120 mM triethanolamine (pH 7.5), 0.1 mM EDTA, 0.5 mM NADP\*, and 10-150 μM 6-phosphogluconate as substrate at 20-25°C. The production of NADPH is measured fluorimetrically (340 nm excitation, 450 nm emission) and is indicative of 6-phosphogluconate dehydrogenase activity. Alternatively, the production of NADPH is measured photometrically, based on absorbance at 340 nm. The molar amount of NADPH produced in the reaction is proportional to the 6-phosphogluconate dehydrogenase activity in the sample (Tetaud et al., supra).

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Ribonucleotide diphosphate reductase activity of ENZM is determined by incubating purified ENZM, or a composition comprising ENZM, along with dithiothreitol, Mg\*\*, and ADP, GDP, CDP, or UDP substrate. The product of the reaction, the corresponding deoxyribonucleotide, is separated from the substrate by thin-layer chromatography. The reaction products can be distinguished from the reactants based on rates of migration. The use of radiolabeled substrates is an alternative for increasing the sensitivity of the assay. The amount of deoxyribonucleotides produced in the reaction is proportional to the amount of ribonucleotide diphosphate reductase activity in the sample (note that this is true only for pre-steady state kinetic analysis of ribonucleotide diphosphate reductase activity, as the enzyme is subject to negative feedback inhibition by products) (Nutter and Cheng, supra).

Dihydrodiol dehydrogenase activity of ENZM is measured by incubating purified ENZM, or a

25 composition comprising ENZM, in a reaction mixture comprising 50 mM glycine (pH 9.0), 2.3 mM

NADP\*, 8% DMSO, and a trans-dihydrodiol substrate, selected from the group including but not

limited to, (±)-trans-naphthalene-1,2-dihydrodiol, (±)-trans-phenanthrene-1,2-dihydrodiol, and (±)-transchrysene-1,2-dihydrodiol. The oxidation reaction is monitored at 340 nm to detect the formation of

NADPH, which is indicative of the oxidation of the substrate. The reaction mixture can also be

analyzed before and after the addition of ENZM by circular dichroism to determine the

stereochemistry of the reaction components and determine which enantiomers of a racemic substrate

composition are oxidized by the ENZM (Penning, supra).

Glutathione S-transferase (GST) activity of ENZM is determined by measuring the ENZM

catalyzed conjugation of GSH with 1-chloro-2,4-dinitrobenzene (CDNB), a common substrate for most GSTs. ENZM is incubated with 1 mM CDNB and 2.5 mM GSH together in 0.1M potassium phosphate buffer, pH 6.5, at 25°C. The conjugation reaction is measured by the change in absorbance at 340 nm using an ultraviolet spectrophometer. ENZM activity is proportional to the change in absorbance at 340 nm.

15-oxoprostaglandin 13-reductase (PGR) activity of ENZM is measured following the separation of contaminating 15-hydroxyprostaglandin dehydrogenase (15-PGDH) activity by DEAE chromatography. Following isolation of PGR containing fractions (or using the purified ENZM), activity is assayed in a reaction comprising 0.1 M sodium phosphate (pH 7.4), 1 mM 2-10 mercaptoethanol, 20 μg substrate (e.g., 15-oxo derivatives of prostaglandins PGE<sub>1</sub>, PGE<sub>2</sub>, and PGE<sub>20</sub>), and 1 mM NADH (or a higher concentration of NADPH). ENZM is added to the reaction which is then incubated for 10 min at 37°C before termination by the addition of 0.25 ml 2 N NaOH. The amount of 15-oxo compound remaining in the sample is determined by measuring the maximum absorption at 500 nm of the terminated reaction and comparing this value to that of a terminated control reaction that received no ENZM. 1 unit of enzyme is defined as the amount required to catalyze the oxidation of 1 μmol substrate per minute and is proportional to the amount of PGR activity in the sample.

Choline dehydrogenase activity of ENZM is identified by the ability of E. coli, transformed with an ENZM expression vector, to grow on media containing choline as the sole carbon and nitrogen source. The ability of the transformed bacteria to thrive is indicative of choline dehydrogenase activity (Magne Østerås, M. (1998) Proc. Natl. Acad. Sci. USA 95:11394-11399).

ENZM thioredoxin activity is assayed as described (Luthman, M. (1982) Biochemistry 21:6628-6633). Thioredoxins catalyze the formation of disulfide bonds and regulate the redox environment in cells to enable the necessary thiol:disulfide exchanges. One way to measure the thiol:disulfide exchange is by measuring the reduction of insulin in a mixture containing 0.1 M potassium phosphate, pH 7.0, 2 mM EDTA, 0.16 µM insulin, 0.33 mM DTT, and 0.48 mM NADPH. Different concentrations of ENZM are added to the mixture, and the reaction rate is followed by monitoring the oxidation of NADPH at 340 nM.

ENZM transferase activity is measured through assays such as a methyl transferase assay in
which the transfer of radiolabeled methyl groups between a donor substrate and an acceptor substrate
is measured (Bokar, J.A. et al. (1994) J. Biol. Chem. 269:17697-17704). Reaction mixtures (50 µl
final volume) contain 15 mM HEPES, pH 7.9, 1.5 mM MgCl<sub>2</sub>, 10 mM dithiothreitol, 3%
polyvinylalcohol, 1.5 µCl [methyl-³H]AdoMet (0.375 µM AdoMet) (DuPont-NEN), 0.6 µg ENZM, and

acceptor substrate (0.4  $\mu g$  [35S]RNA or 6-mercaptopurine (6-MP) to 1 mM final concentration). Reaction mixtures are incubated at 30°C for 30 minutes, then at 65°C for 5 minutes. The products are separated by chromatography or electrophoresis and the level of methyl transferase activity is determined by quantification of methyl-3H recovery.

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Aminotransferase activity of ENZM is assayed by incubating samples containing ENZM for 1 hour at 37°C in the presence of 1 mM L-kynurenine and 1 mM 2-oxoglutarate in a final volume of 200 μl of 150 mM Tris acetate buffer (pH 8.0) containing 70 μM PLP. The formation of kynurenic acid is quantified by HPLC with spectrophotometric detection at 330 nm using the appropriate standards and controls well known to those skilled in the art. In the alternative, L-3-hydroxykynurenine is used as 30 substrate and the production of xanthurenic acid is determined by HPLC analysis of the products with UV detection at 340 nm. The production of kynurenic acid and xanthurenic acid, respectively, is indicative of aminotransferase activity (Buchli et al., supra).

In another alternative, aminotransferase activity of ENZM is measured by determining the activity of purified ENZM or crude samples containing ENZM toward various amino and oxo acid 15 substrates under single turnover conditions by monitoring the changes in the UV/VIS absorption spectrum of the enzyme-bound cofactor, pyridoxal 5'-phosphate (PLP). The reactions are performed at 25°C in 50 mM 4-methylmorpholine (pH 7.5) containing 9 µM purified ENZM or ENZM containing samples and substrate to be tested (amino and oxo acid substrates). The half-reaction from amino acid to oxo acid is followed by measuring the decrease in absorbance at 360 nm and the increase in 20 absorbance at 330 nm due to the conversion of enzyme-bound PLP to pyridoxamine 5' phosphate (PMP). The specificity and relative activity of ENZM is determined by the activity of the enzyme preparation against specific substrates (Vacca, supra).

ENZM chitinase activity is determined with the fluorogenic substrates 4-methylumbelliferyl chitotriose, methylumbelliferyl chitobiose, or methylumbelliferyl N-acetylglucosamine. Purified ENZM 25 is incubated with 0.5uM substrate at pH 4.0 (0.1M citrate buffer), pH 5.0 (0.1M phosphate buffer), or pH 6.0 (0.1M Tris-HCL). After various times of incubation, the reaction is stopped by the addition of 0.1M glycine buffer, pH 10.4, and the concentration of free methylumbelliferone is determined fluorometrically. Chitinase B from Serratia marcescens may be used as a positive control (Hakala, supra).

ENZM isomerase activity is determined by measuring 2-hydroxyhepta-2,4-diene,1,7 dioate isomerase (HHDD isomerase) activity, as described by Garrido-Peritierra, A. and R.A. Cooper (1981; Eur. J. Biochem. 17:581-584). The sample is combined with 5-carboxymethyl-2-oxo-hex-3-ene-1,5, dioate (CMHD), which is the substrate for HHDD isomerase. CMHD concentration is monitored by

measuring its absorbance at 246 nm. Decrease in absorbance at 246 nm is proportional to HHDD isomerase activity of ENZM.

ENZM isomerase activity such as peptidyl prolyl cis/trans isomerase activity can be assayed by an enzyme assay described by Rahfeld (supra). The assay is performed at 10°C in 35 mM

5 HEPES buffer, pH 7.8, containing chymotrypsin (0.5 mg/ml) and ENZM at a variety of concentrations. Under these assay conditions, the substrate, Suc-Ala-Xaa-Pro-Phe-4-NA, is in equilibrium with respect to the prolyl bond, with 80-95% in trans and 5-20% in cis conformation. An aliquot (2 µl) of the substrate dissolved in dimethyl sulfoxide (10 mg/ml) is added to the reaction mixture described above. Only the cis isomer is a substrate for cleavage by chymotrypsin. Thus, as the substrate is isomerized by ENZM, the product is cleaved by chymotrypsin to produce 4-nitroanilide, which is detected by its absorbance at 390 nm. 4-Nitroanilide appears in a time-dependent and a ENZM concentration-dependent manner.

Alternatively, peptidyl prolyl *cis-trans* isomerase activity of ENZM can be assayed using a chromogenic peptide in a coupled assay with chymotrypsin (Fischer, G. et al. (1984) Biomed.

Biochim. Acta 43:1101-1111).

UDP glucuronyltransferase activity of ENZM is measured using a colorimetric determination of free amine groups (Gibson, G.G. and P. Skett (1994) Introduction to Drug Metabolism. Blackie Academic and Professional, London). An amine-containing substrate, such as 2-aminophenol, is incubated at 37°C with an aliquot of the enzyme in a reaction buffer containing the necessary cofactors (40 mM Tris pH 8.0, 7.5 mM MgCl<sub>2</sub>, 0.025% Triton X-100, 1 mM ascorbic acid, 0.75 mM UDP-glucuronic acid). After sufficient time, the reaction is stopped by addition of ice-cold 20% trichloroacetic acid in 0.1 M phosphate buffer pH 2.7, incubated on ice, and centrifuged to clarify the supernatant. Any unreacted 2-aminophenol is destroyed in this step. Sufficient freshly-prepared sodium nitrite is then added; this step allows formation of the diazonium salt of the glucuronidated product. Excess nitrite is removed by addition of sufficient ammonium sulfamate, and the diazonium salt is reacted with an aromatic amine (for example, N-naphthylethylene diamine) to produce a colored azo compound which can be assayed spectrophotometrically (at 540 nm, for example). A standard curve can be constructed using known concentrations of aniline, which will form a chromophore with similar properties to 2-aminophenol glucuronide.

Adenylosuccinate synthetase activity of ENZM is measured by synthesis of AMP from IMP. The sample is combined with AMP. IMP concentration is monitored spectrophotometrically at 248 nm at 23°C (Wang, W. et al. (1995) J. Biol. Chem. 270:13160-13163). The increase in IMP concentration is proportional to ENZM activity.

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In another alternative, xenobiotic carboxylic acid:CoA ligase activity of ENZM is measured by combining the sample with  $\gamma^{33}$ P-ATP and measuring the formation of  $\gamma^{-33}$ P- pyrophosphate with time (Vessey, D.A. et al. (1998) J. Biochem. Mol. Toxicol. 12:151-155).

Protein phosphatase (PP) activity can be measured by the hydrolysis of P-nitrophenyl phosphate (PNPP). ENZM is incubated together with PNPP in HEPES buffer pH 7.5, in the

10 presence of 0.1% β-mercaptoethanol at 37°C for 60 min. The reaction is stopped by the addition of 6 ml of 10 N NaOH (Diamond, R.H. et al. (1994) Mol. Cell. Biol. 14:3752-62).

Alternatively, acid phosphatase activity of ENZM is demonstrated by incubating ENZM containing extract with 100  $\mu$ l of 10 mM PNPP in 0.1 M sodium citrate, pH 4.5, and 50  $\mu$ l of 40 mM NaCl at 37°C for 20 min. The reaction is stopped by the addition of 0.5 ml of 0.4 M glycine/NaOH, pH 10.4 (Saftig, P. et al. (1997) J. Biol. Chem. 272:18628-18635). The increase in light absorbance at 410 nm resulting from the hydrolysis of PNPP is measured using a spectrophotometer. The increase in light absorbance is proportional to the activity of ENZM in the assay.

In the alternative, ENZM activity is determined by measuring the amount of phosphate removed from a phosphorylated protein substrate. Reactions are performed with 2 or 4 nM ENZM in a final volume of 30 µl containing 60 mM Tris, pH 7.6, 1 mM EDTA, 1 mM EGTA, 0.1% 2-mercaptoethanol and 10 µM substrate, <sup>32</sup>P-labeled on serine/threonine or tyrosine, as appropriate. Reactions are initiated with substrate and incubated at 30° C for 10-15 min. Reactions are quenched with 450 µl of 4% (w/v) activated charcoal in 0.6 M HCl, 90 mM Na<sub>4</sub>P<sub>2</sub>O<sub>7</sub>, and 2 mM NaH<sub>2</sub>PO<sub>4</sub>, then centrifuged at 12,000 × g for 5 min. Acid-soluble <sup>32</sup>Pi is quantified by liquid scintillation counting (Sinclair, C. et al. (1999) J. Biol. Chem. 274:23666-23672).

The adenosine deaminase activity of ENZM is determined by measuring the rate of deamination that occurs when adenosine substrate is incubated with ENZM. Reactions are performed with a predetermined amount of ENZM in a final volume of 3.0 ml containing 53.3 mM potassium phosphate and 0.045 mM adenosine. Assay reagents excluding ENZM are mixed in a quartz cuvette and equilibrated to 25° C. Reactions are initiated by the addition of ENZM and are mixed immediately by inversion. The decrease in light absorbance at 265 nm resulting from the hydrolysis of adenosine to inosine is measured using a spectrophotometer. The decrease in the A<sub>265 nm</sub> is recorded for approximately 5 minutes. The decrease in light absorbance is proportional to the activity of ENZM in

the assay.

ENZM hydrolase activity is measured by the hydrolysis of appropriate synthetic peptide substrates conjugated with various chromogenic molecules in which the degree of hydrolysis is quantified by spectrophotometric (or fluorometric) absorption of the released chromophore (Beynon and Bond, supra, pp.25-55). Peptide substrates are designed according to the category of protease activity as endopeptidase (serine, cysteine, aspartic proteases), aminopeptidase (leucine aminopeptidase), or carboxypeptidase (Carboxypeptidase A and B, procollagen C-proteinase).

An assay for carbonic anhydrase activity of ENZM uses the fluorescent pH indicator 8-hydroxypyrene-1,3,6-trisulfonate (pyranine) in combination with stopped-flow fluorometry to measure 10 carbonic anhydrase activity (Shingles, et al. 1997, Anal. Biochem. 252:190-197). A pH 6.0 solution is mixed with a pH 8.0 solution and the initial rate of bicarbonate dehydration is measured. Addition of carbonic anhydrase to the pH 6.0 solution enables the measurement of the initial rate of activity at physiological temperatures with resolution times of 2 ms. Shingles et al. (supra) used this assay to resolve differences in activity and sensitivity to sulfonamides by comparing mammalian carbonic anhydrase isoforms. The fluorescent technique's sensitivity allows the determination of initial rates with a protein concentration as little as 65 ng/ml.

Decarboxylase activity of ENZM is measured as the release of CO<sub>2</sub> from labeled substrate. For example, ornithine decarboxylase activity of ENZM is assayed by measuring the release of CO<sub>2</sub> from L-[1-<sup>14</sup>C]-ornithine (Reddy, S.G et al. (1996) J. Biol. Chem. 271:24945-24953). Activity is measured in 200 µl assay buffer (50 mM Tris/HCl, pH 7.5, 0.1 mM EDTA, 2 mM dithiothreitol, 5 mM NaF, 0.1% Brij35, 1 mM PMSF, 60 µM pyridoxal-5-phosphate) containing 0.5 mM L-ornithine plus 0.5 µCi L-[1-<sup>14</sup>C]ornithine. The reactions are stopped after 15-30 minutes by addition of 1 M citric acid, and the <sup>14</sup>CO<sub>2</sub> evolved is trapped on a paper disk filter saturated with 20 µl of 2 N NaOH. The radioactivity on the disks is determined by liquid scintillation spectography. The amount of <sup>14</sup>CO<sub>2</sub> released is proportional to ornithine decarboxylase activity of ENZM.

AdoHCYase activity of ENZM in the hydrolytic direction is performed spectroscopically by measuring the rate of the product (homocysteine) formed by reaction with 5,5'-Dithiobis(2-nitrobenzoic acid) (DTNB). To 800 μl of an enzyme solution containing 4.7 μg of ENZM and 4 units of adenosine dearninase in 50 mM potassium phosphate buffer, pH 7.2, containing 1 mM EDTA (buffer A), is added 200 μl of S-Adenosyl-L-homocysteine (500 μM) containing 250 μM DTNB in buffer A. The reaction mixture is incubated at 37°C for 2 minutes. Hydrolytic activity is monitored at 412 mm continuously using a diode array UV spectrophotometer. Enzyme activity is defined as the amount of enzyme that can hydrolyze 1 μmol of S-Adenosyl-L-homocysteine/minute (Yuan, C-S et al. (1996) J.

Biol. Chem. 271:28009-28015).

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AdoHCYase activity of ENZM can be measured in the synthetic direction as the production of S-adenosyl homocysteine using 3-deazaadenosine as a substrate (Sganga et al. supra). Briefly, ENZM is incubated in a 100 μl volume containing 0.1 mM 3-deazaadenosine, 5 mM homocysteine, 20 mM HEPES (pH 7.2). The assay mixture is incubated at 37°C for 15 minutes. The reaction is terminated by the addition of 10 μl of 3 M perchloric acid. After incubation on ice for 15 minutes, the mixture is centrifuged for 5 minutes at 18,000 x g in a microcentrifuge at 4°C. The supernatant is removed, neutralized by the addition of 1 M potassium carbonate, and centrifuged again. A 50 μl aliquot of supernatant is then chromatographed on an Altex Ultrasphere ODS column (5 μm particles, 4.6 x 250 mm) by isocratic elution with 0.2 M ammonium dihydrogen phosphate (Aldrich) at a flow rate of 1 ml/min. Protein is determined by the bicinchoninic acid assay (Pierce).

Alternatively, AdoHCYase activity of ENZM can be measured in the synthetic direction by a TLC method (Hershfield, M.S. et al. (1979) J. Biol. Chem. 254:22-25). In a preincubation step, 50 μM [8<sup>14</sup>C]adenosine is incubated with 5 molar equivalents of NAD\* for 15 minutes at 22°C. Assay 15 samples containing ENZM in a 50 μl final volume of 50 mM potassium phosphate buffer, pH 7.4, 1 mM DTT, and 5 mM homocysteine, are mixed with the preincubated [8<sup>14</sup>C]adenosine/NAD\* to initiate the reaction. The reaction is incubated at 37°C, and 1 μl samples are spotted on TLC plates at 5 minute intervals for 30 minutes. The chromatograms are developed in butanol-1/glacial acetic acid/water (12:3:5, v/v) and dried. Standards are used to identify substrate and products under ultraviolet light. The complete spots containing [1<sup>14</sup>C]adenosine and [1<sup>14</sup>C]SAH are then detected by exposing x-ray film to the TLC plate. The radiolabeled substrate and product are then cut from the chromatograms and counted by liquid scintillation spectrometry. Specific activity of the enzyme is determined from the linear least squares slopes of the product vs time plots and the milligrams of protein in the sample (Bethin, K.E. et al. (1995) J. Biol. Chem. 270:20698-20702).

Asparaginase activity of ENZM can be measured in the hydrolytic direction by determining the amount of radiolabeled L-asparatate released from 0.6 mM N\*.9'-N-acetylglucosaminyl-L-asparagine substrate when it is incubated at 25 °C with ENZM in 50 mM phosphate buffer, pH 7.5 (Kaartinen, V. et al. (1991) J. Biol. Chem. 266:5860-5869).

Acyl CoA Acid Hydrolase activity of ENZM in the hydrolytic direction is performed

spectroscopically by monitoring the appearance of the product (CoASH) formed by reaction of

substrate (acyl-CoA) and ENZM with 5,5'-Dithiobis(2-nitrobenzoic acid) (DTNB). The final reaction

volume is 1 ml of 0.05 M potassium phosphate buffer, pH 8, containing 0.1 mM DTNB, 20 µg/ml

bovine serum albumin, 10 µM of acyl-CoA of different lengths (C6-CoA, C10-CoA, C14-CoA and

C18-CoA, Sigma), and ENZM. The reaction mixture is incubated at 22 °C for 7 minutes. Hydrolytic activity is monitored spectrophotometrically by measuring absorbance at 412 nm (Poupon, V. et al. (1999) J. Biol. Chem. 274:19188-19194).

ENZM activity of ENZM can be measured spectrophotometrically by determining the amount
of solubilized RNA that is produced as a result of incubation of RNA substrate with ENZM. 5 μ1 (20
μg) of a 4 mg/ml solution of yeast tRNA (Sigma) is added to 0.8 ml of 40 mM sodium phosphate, pH
7.5, containing ENZM. The reaction is incubated at 25 °C for 15 minutes. The reaction is stopped by
addition of 0.5 ml of an ice-cold fresh solution of 20 mM lanthanum nitrate plus 3% perchloric acid.
The stopped reaction is incubated on ice for at least 15 min, and the insoluble tRNA is removed by
centrifugation for 5 min at 10,000 g. Solubilized tRNA is determined as UV absorbance (260 mm) of
the remaining supernatant, with Λ<sub>200</sub> of 1.0 corresponding to 40 μg of solubilized RNA (Rosenberg,
H.F. et al. (1996) Nucleic Acids Research 24:3507-3513).

ENZM activity can be determined as the ability of ENZM to cleave <sup>32</sup>P internally labeled T.

thermophila pre-tRNA<sup>GB</sup>. ENZM and substrate are added to reaction vessels and reactions are

carried out in MBB buffer (50 mM Tris-HCl (pH 7.5), 10 mM MgCl<sub>2</sub>) for 1 hour at 37°C. Reactions

are terminated with the addition of an equal volume of sample loading buffer (SLB: 40 mM EDTA, 8

M urea, 0.2% xylene cyanol, and 0.2% bromophenol blue). The reaction products are separated by

electrophoresis on 8 M urea, 6% polyacrylamide gels and analyzed using detection instruments and

software capable of quantification of the products. One unit of ENZM activity is defined as the

amount of enzyme required to cleave 10% of 28 fmol of T. thermophila pre-tRNA<sup>GB</sup> to mature

products in 1 hour at 37°C (True, H.L. et al. (1996) J. Biol. Chem. 271:16559-16566).

Alternatively, cleavage of <sup>32</sup>P internally labeled substrate tRNA by ENZM can be determined in a 20 µl reaction mixture containing 30 mM HEPES-KOH (pH 7.6), 6 mM MgCl<sub>2</sub>, 30 mM KCl, 2 mM DTT, 25 µg/ml bovine serum albumin, 1 unit/µl rRNasin, and 5,000-50,000 cpm of gel-purified substrate RNA. 3.0 µl of ENZM is added to the reaction mixture, which is then incubated at 37°C for 30 minutes. The reaction is stopped by guanidinium/phenol extraction, precipitated with ethanol in the presence of glycogen, and subjected to denaturing polyacrylamide gel electrophoresis (6 or 8% polyacrylamide, 7 M urea) and autoradiography (Rossmanith, W. et al. (1995) J. Biol. Chem. 270:12885-12891). The ENZM activity is proportional to the amount of cleavage products detected.

ENZM activity can be measured by determining the amount of free adenosine produced by the hydrolysis of AMP, as described by Sala-Newby et al., *supra*. Briefly, ENZM is incubated with AMP in a suitable buffer for 10 minutes at 37 °C. Free adenosine is separated from AMP and measured by reverse phase HPLC.

30

Alternatively. ENZM activity is measured by the hydrolysis of ADP-ribosylarginine (Konczalik, P. and J. Moss (1999) J. Biol. Chem. 274:16736-16740). 50 ng of ENZM is incubated with 100 µM ADP-ribosyl-[14Clarginine (78,000 cpm) in 50 mM potassium phosphate, pH 7.5, 5 mM dithiothreitol, 10 mM MgCl2 in a final volume of 100 µl. After 1 h at 37° C, 90 µl of the sample is 5 applied to a column (0.5 × 4 cm) of Affi-Gel 601 (boronate) equilibrated and eluted with five 1-ml portions of 0.1 M glycine, pH 9.0, 0.1 M NaCl, and 10 mM MgCl<sub>2</sub>. Free <sup>14</sup>C-Arg in the total eluate is measured by liquid scintillation counting.

Epoxide hydrolase activity of ENZM can be determined with a radiometric assay utilizing [H3]-labeled trans-stilbene oxide (TSO) as substrate. Briefly, ENZM is preincubated in Tris-HCl pH 7.4 buffer in a total volume of 100 μl for 1 minute at 37°C. 1 μl of [H³l-labeled TSO (0.5 μM in EtOH) is added and the reaction mixture is incubated at 37°C for 10 minutes. The reaction mixture is extracted with 200 \(\mu\) n-dodecane. 50 \(\mu\)] of the aqueous phase is removed for quantification of diol product in a liquid scintillation counter (LSC). ENZM activity is calculated as nmol diol product/min/mg protein (Gill, S.S. et al. (1983) Analytical Biochemistry 131:273-282).

Lysophosphatidic acid acyltransferase activity of ENZM is measured by incubating samples containing ENZM with 1 mM of the thiol reagent 5,5'-dithiobis(2-nitrobenzoic acid) (DTNB), 50 µm LPA, and 50 µm acyl-CoA in 100 mM Tris-HCl, pH 7.4. The reaction is initiated by addition of acyl-CoA, and allowed to reach equilibrium. Transfer of the acyl group from acyl-CoA to LPA releases free CoA, which reacts with DTNB. The product of the reaction between DTNB and free CoA 20 absorbs at 413 nm. The change in absorbance at 413 nm is measured using a spectrophotometer, and is proportional to the lysophosphatidic acid acyltransferase activity of ENZM in the sample.

15

N-acyltransferase activity of ENZM is measured using radiolabeled amino acid substrates and measuring radiolabel incorporation into conjugated products. ENZM is incubated in a reaction buffer containing an unlabeled acyl-CoA compound and radiolabeled amino acid, and the radiolabeled acyl-25 conjugates are separated from the unreacted amino acid by extraction into n-butanol or other appropriate organic solvent. For example, Johnson, M. R. et al. (1990; J. Biol. Chem. 266:10227-10233) measured bile acid-CoA:amino acid N-acyltransferase activity by incubating the enzyme with cholyl-CoA and 3H-glycine or 3H-taurine, separating the tritiated cholate conjugate by extraction into n-butanol, and measuring the radioactivity in the extracted product by scintillation. Alternatively, N-30 acyltransferase activity is measured using the spectrophotometric determination of reduced CoA (CoASH) described below.

N-acetyltransferase activity of ENZM is measured using the transfer of radiolabel from [14C]acetyl-CoA to a substrate molecule (for example, see Deguchi, T. (1975) J. Neurochem.

24:1083-5). Alternatively, a newer spectrophotometric assay based on DTNB reaction with CoASH may be used. Free thiol-containing CoASH is formed during N-acetyltransferase catalyzed transfer of an acetyl group to a substrate. CoASH is detected using the absorbance of DTNB conjugate at 412 nm (De Angelis, J. et al. (1997) J. Biol. Chem. 273:3045-3050). ENZM activity is proportional to the rate of radioactivity incorporation into substrate, or the rate of absorbance increase in the spectrophotometric assay.

Galactosyltransferase activity of ENZM is determined by measuring the transfer of galactose from UDP-galactose to a GlcNAc-terminated oligosaccharide chain in a radioactive assay. (Kolbinger, F. et al. (1998) J. Biol. Chem. 273:58-65.) The ENZM sample is incubated with 14 μl of assay stock solution (180 mM sodium cacodylate, pH 6.5, 1 mg/ml bovine serum albumin, 0.26 mM UDP-galactose, 2 μl of UDP-PHlgalactose), 1 μl of MnCl<sub>4</sub> (500 mM), and 2.5 μl of GlcNAcβO-(CH<sub>2</sub>)<sub>1</sub>-CO<sub>2</sub>Me (37 mg/ml in dimethyl sulfoxide) for 60 minutes at 37 °C. The reaction is quenched by the addition of 1 ml of water and loaded on a C18 Sep-Pak cartridge (Waters), and the column is washed twice with 5 ml of water to remove unreacted UDP-PHlgalactose. The [H]galactosylated 15 GlcNAcβO-(CH<sub>2</sub>)<sub>1</sub>-CO<sub>2</sub>Me remains bound to the column during the water washes and is eluted with 5 ml of methanol. Radioactivity in the eluted material is measured by liquid scintillation counting and is proportional to galactosyltransferase activity of ENZM in the starting sample.

Phosphoribosyltransferase activity of ENZM is measured as the transfer of a phosphoribosyl group from phosphoribosylprophosphate (PRPP) to a purine or pyrimidine base. Assay mixture (20 20 µl) containing 50 mM Tris acetate, pH 9.0, 20 mM 2-mercaptoethanol, 12.5 mM MgCl<sub>2</sub>, and 0.1 mM labeled substrate, for example, [¹¹C]uracil, is mixed with 20 µl of ENZM diluted in 0.1 M Tris acetate, pH 9.7, and 1 mg/ml bovine serum albumin. Reactions are preheated for 1 min at 37°C, initiated with 10 µl of 6 mM PRPP, and incubated for 5 min at 37°C. The reaction is stopped by heating at 100°C for 1 min. The product [¹⁴C]UMP is separated from [¹⁴C]uracil on DEAE-cellulose paper (Turner, 25 R.J. et al. (1998) J. Biol. Chem. 273:5932-5938). The amount of [¹⁴C]UMP produced is proportional to the phosphoribosyltransferase activity of ENZM.

ADP-ribosyltransferase activity of ENZM is measured as the transfer of radiolabel from adenine-NAD to agmatine (Weng, B. et al. (1999) J. Biol. Chem. 274:31797-31803). Purified ENZM is incubated at 30°C for 1 hr in a total volume of 300 μl containing 50 mM potassium phosphate (pH. 30 7.5), 20 mM agmatine, and 0.1 mM [adenine-U-<sup>14</sup>C]NAD (0.05 mCi). Samples (100 μl) are applied to Dowex columns and [<sup>14</sup>C]ADP-ribosylagmatine eluted with 5 ml of water for liquid scintillation counting. The amount of radioactivity recovered is proportional to ADP-ribosyltransferase activity of ENZM.

An ENZM activity assay measures aminoacylation of tRNA in the presence of a radiolabeled substrate. SYNT is incubated with [<sup>14</sup>C]-labeled amino acid and the appropriate cognate tRNA (for example, [<sup>14</sup>C]alanine and tRNA<sup>4a</sup>) in a buffered solution. <sup>14</sup>C-labeled product is separated from free [<sup>14</sup>C]amino acid by chromatography, and the incorporated <sup>14</sup>C is quantified using a scintillation counter.

5 The amount of <sup>14</sup>C-labeled product detected is proportional to the activity of ENZM in this assay (Ibba, M. et al. (1997) Science 278:1119-1122).

Alternatively, argininosuccinate synthase activity of ENZM is measured based on the conversion of [³H]aspartate to [³H]argininosuccinate. ENZM is incubated with a mixture of [³H]aspartate, citrulline, Tris-HCl (pH 7.5), ATP, MgCl<sub>2</sub>, KCl, phosphoenolpyruvate, pyruvate kinase, myokinase, and pyrophosphatase, and allowed to proceed for 60 minutes at 37 °C. Enzyme activity was terminated with addition of acetic acid and heating for 30 minutes at 90 °C. [³H]argininosuccinate is separated from un-catalyzed [³H]aspartate by chromatography and quantified by liquid scintillation spectrometry. The amount of [³H]argininosuccinate detected is proportional to

Alternatively, the esterase activity of ENZM is assayed by the hydrolysis of p-nitrophenylacetate (NPA). ENZM is incubated together with  $0.1~\mu M$  NPA in 0.1~M potassium phosphate buffer (pH 7.25) containing 150 mM NaCl. The hydrolysis of NPA is measured by the increase of absorbance at 400 nm with a spectrophotometer. The increase in light absorbance is proportional to the activity of ENZM (Probst, M.R. et al. (1994) J. Biol. Chem. 269:21650-21656).

the activity of ENZM in this assay (O'Brien, W. E. (1979) Biochemistry 18:5353-5356).

### 20 XIX. Identification of ENZM Agonists and Antagonists

15

Agonists or antagonists of ENZM activation or inhibition may be tested using the assays described in section XVIII. Agonists cause an increase in ENZM activity and antagonists cause a decrease in ENZM activity.

Various modifications and variations of the described compositions, methods, and systems of
the invention will be apparent to those skilled in the art without departing from the scope and spirit of
the invention. It will be appreciated that the invention provides novel and useful proteins, and their
encoding polynucleotides, which can be used in the drug discovery process, as well as methods for
using these compositions for the detection, diagnosis, and treatment of diseases and conditions.
Although the invention has been described in connection with certain embodiments, it should be
understood that the invention as claimed should not be unduly limited to such specific embodiments.
Nor should the description of such embodiments be considered exhaustive or limit the invention to the
precise forms disclosed. Furthermore, elements from one embodiment can be readily recombined with
elements from one or more other embodiments. Such combinations can form a number of

embodiments within the scope of the invention. It is intended that the scope of the invention be defined by the following claims and their equivalents.

# Table 1

Incute Project ID	Polynentide	Incyte	Polynucleotide	Incyte	
and a soften and a soften a	SEQ ID NO:	ptide ID	SEQ ID NO:	Polynucleotide	
					Incyte Full Length Clones
7511289	-	7511289CD1	39	7511289CB1	
7511056	2	7511056CD1	40	7511056CB1	1443673CA2
7511567	3	7511567CD1	41	7511567CB1	
7511651	4	7511651CD1	42		
7511881	5	7511881CD1	43	7511881CB1	95003710CA2
7512181	9	7512181CD1	44	7512181CB1	90139166CA2
7511726	7	7511726CD1	45	7511726CB1	
7511057	. 00	7511057CD1	46	7511057CB1	1449639CA2, 95164104CA2, 95164112CA2, 95164144CA2,
					95164160CA2, 95164244CA2, 95164276CA2
7511078	6	7511078CD1	47	7511078CB1	1930540CA2
7511505	10	7511505CD1	48	7511505CB1	
7511552	11	7511552CD1	49	7511552CB1	
7511722	12	7511722CD1	50	7511722CB1	
7511489	13	7511489CD1	51	7511489CB1	
7511497	14	7511497CD1	52		
7511498	15	7511498CD1	53		95133328CA2
7511612	16	7511612CD1	54		6604874CA2
7511624	17	7511624CD1	55	7511624CB1	
7511626	18	7511626CD1	26	7511626CB1	
7512885	19	7512885CD1	57	7512885CB1	90025580CA2
7511965	20	7511965CD1	58	7511965CB1	
7512403	21	7512403CD1	59	7512403CB1	
7512564	22	7512564CD1	09	7512564CB1	95110656CA2
7512646	23	7512646CD1	19	7512646CB1	6883320CA2
7512700	24	7512700CD1	62	7512700CB1	
7512707	25	7512707CD1	63	7512707CB1	95078276CA2
7512710	26	7512710CD1	64	7512710CB1	
7512884	27	7512884CD1	65	7512884CB1	
7512931	28	7512931CD1	99	7512931CB1	

# Table 1

Incote Project ID	Polypeptide	Incyte	Polynucleotide	Incyte	
,	SEO ID NO:	Polypeptide ID   SEQ ID NO:	SEQ ID NO:	Polynucleotide	
				D	Incyte Full Length Clones
7512033	29	7512933CD1	29	7512933CB1	95137927CA2, 95137951CA2, 95137967CA2, 95138043CA2
7512942	30	7512942CD1	89	7512942CB1	
7513736	31	7513736CD1	69	7513736CB1	90089783CA2, 90089883CA2
7512779	32	7512779CD1	70	7512779CB1	
7512877	33	7512877CD1	71	7512877CB1	90065442CA2
7510780	34	7512782CD1	72	7512782CB1	
7510784	3,5	7512784CD1	73	7512784CB1	
7512794	36	7512794CD1	74	7512794CB1	8683815CA2
7512886	37	7512886CD1	75	7512886CB1	
0000000	00	1	7.6	7512020CR1	8131571CA2

Polypeptide SEQ Incyte ID NO: Polype	Incyte Polypeptide ID	GenBank ID NO: Probability or PROTEOME Score ID NO:	bility	Amotation
	7511289CD1	599428 FLJ2048 1.9E-33	1.9E-33	[Homo sapiens][Small molecule-binding protein] Protein containing two EF hand domains, which are found in signaling, buffering or transport proteins
2	7511056CD1	g11120435	6.2E-71	[Homo sapiens] alpha-aminoadipic semialdehyde dehydrogenase-phosphopantetheinyl transferase
		569688 AASDHP 1.9E-56 PT	1.9E-56	[Homo sapiens][Transferase] Member of the 4'-phosphopantetheinyl transferase (holo-acyl carrier protein synthase, ACPS) superfamily
3	7511567CDI	g2725625	2.0E-287	[Homo sapiens] acetolactate synthase
		715550 T26C12.   1.9E-152	1.9E-152	[Caenorhabditis elegans][Lyase][Cytoplasmic] Member of the decarboxylase (tentative) protein family
		609176 Hpcl	3.4E-43	[Mus musculus][Lyase][Cytoplasmic; Peroxisome] Protein with strong similarity to human
				fatty acids to make formyl-CoA and 2-methyl-branched fatty aldehydes, member of the
				thiamine pyrophosphate-dependent enzyme family
4	7511651CD1	g337463	4.1E-123	[Homo sapiens] uroporphyrinogen III synthase (EC 4.2.1.75)
				Isai, S. F. et al., Human uroporphyrinogen III synthase: molecular cloning, nucleotide
				sequence, and expression of a full-length cDNA, Proc. Natl. Acad. Sci. U.S.A. 85, 7049- 7053 (1988)
		339612 UROS	3.3E-124	[Homo sapiens][Lyase] Uroporphyrinogen III synthase (congenital crythropoletic
				porphyria), catalyzes the conversion of hydroxymethylbilane to uroporphyrinogen-III in
				heme biosynthesis; genetic mutation is associated with congenual crythropoictic porphyria
				Deybach, J. C. et al., Point mutations in the uroporphyrinogen III synthase gene in congenital erythropoietic porphyria (Gunther's disease), Blood 75, 1763-5 (1990).
		581807 Uros	1.1E-95	[Mus musculus][Lyase] Uroporphyrinogen III synthase, catalyzes the conversion of hydroxymethylbilane to uroporphyrinogen-III in heme bioxintesis; mutation of the human hydroxymethylbilane to uroporphyrinogen-III in heme bioxintesis;
				UROS gene is associated with congenital crythropoletic purphyria

Table 2

				_		_		_			_	_		T	-7	_		$\neg$		1
Amotation	Aizencang, G. I. et al. Uropomblyrinogen III synthase. An alternative promoter controls erythroid-specific expression in the murine gene., J Biol Chem 275, 2295-304 (2000).	[Homo sapiens] leukotriene C4 synthase	Penrose, J. F. et al., Molecular cloning of the gene for human leukotrene C4 synthase. Organization, uncloside sequence, and chromosomal localization to 5q35, J. Biol. Chem. 2711, 1156-11361 (1996)	[Homo sapiens][Transferase][Endoplasmic reticulum; Microsomal fraction; Cytoplasmic;	Unspecified membrane] Leukouriene C4 synthase, an integral membrane protein that conjugates the epoxide intermediate leukortiene A4 to glutathione, forming the	proinflammatory mediator leukotriene C4	Sjolinder, M. et al., Abernant expression of active leukouriene C(4) synthase in CD16(+) neutrophils from patients with chronic myeloid leukemia., Blood 95, 1456-64 (2000).	Hsieh. F. H. et al., T. Helper Cell Type 2 Cytokines Coordinately Regulate Immunoglobulin	E-dependent Cysteinyl Leukotriene Production by Human Cord Blood-derived Mast Cells. Descend induction of Janotriene et al, synthase expression by interfeukin 4., J Exp Med	193, 123-134. (2001).	[Mus musculus][Transferase] Leukotriene C4 synthase, an integral membrane protein that	conjugates the epoxide intermediate leukotriene A4 to glutathione, forming the	proinflammatory mediator leukotriene C4; involved in increasing vascular permeability	during inflammatory responses	[Homo sapiens] paraoxanase-3	[Homo sapiens][Hydrolase] Paraoxonase 3, member of paraoxonase protein family, which	hydrolyze toxic organophosphates, may protect low density lipoprotein against oxidative	modification	Primo-Parmo, S. L. et al., The human serum paraoxonase/arylesterase gene (PON1) is one	member of a multigene family, Commission, 100
Probability Score		5.4E-73		4.4E-74							3.8E-35				2.0E-120	3.3E-114				
GenBank ID NO: Probability or PROTEOME Score ID NO:		g1314483		344532ILTC4S							581981 Ltc4s				g12751374	348104 PON3				
ptide ID		7511881CD1													7512181CD1					
Polypeptide SEQ Incyte ID NO: Polype		5													9					

Table 2

Polypeptide SEQ Incyte ID NO: Polype	Incyte Polypeptide ID	GenBank ID NO: Probability or PROTEOME Score ID NO:	oility	Amountion
		337086 PON2	5.1E-79	[Homo sapiens][Hydrolase] Paraoxonase 2 (ur)lesterase 2), an antioxidant that protects low density hipoprotein against peroxidation, member of a family of proteins that hydrolyze toxic organoptospates; gene mutations are associated with an increased risk of coronary heart disease.
				Sunghera, D. K. et al., DNA, polymorphisms in two paraoxonase genes (POM1 and POM2) are associated with the risk of coronary heart disease, Am J Hum Genet 62, 36-44, (1998).
7	7511726CD1	g1922287	2.2E-107	[Homo sapiens] enoyl-CoA hydratase
				Jansen, U. et al., Human mitochondrial encyl-CoA hydratase gane (BCES1); structural organization and assignment to chromosome 10q26.2-q26.3, Genomics 40, 470-475 (1997)
		763627[Echs1	1.7E-112	[Rattus norvegicus][Lyase][Cytoplasmic; Mitochondrial] Short chain enoyl-Coenzyme A
		•		hydratase 1, catalyzes the hydration of trans-2-encyl-CoA forming 3-hydroxyacyl-CoA in
				mitochondrial fatty acid beta-oxidation, a member of the low similarity hydratase/isomerase
				enzyme superfamily
				Minami-Ishii, N. et al., Molecular cloning and sequence analysis of the cDNA for rat
				mitochondrial enoyl-CoA hydratase. Structural and evolutionary relationships linked to the
				bifunctional enzyme of the peroxisomal beta- oxidation system., Eur J Biochem 185, 73-8
				(1969). To be a Meteoratic and anomalouised studies of the hydratese and isomerase
				Actema, 1. K. et al., Mutagenic and enzymotogical studies of the hydratase and isomorties of 2-enoyl-CoA hydratase-1., Biochemistry 38, 2991-9. (1999).
		662412/ECHS1	1.8E-108	[Homo sapiens][Lyase][Cytoplasmic; Mitochondrial] Short chain encyl-Coenzyme A
		-		hydratase 1, catalyzes the second step in mitochondrial fatty acid beta-oxidation
				Sakata, M. et al., Messenger RNA differential display reverse-transcriptase-polymerase-
				chain-reaction analysis of a progestogen-suppressive gene in a human endometrial-cancer
				cell line., Int J Cancer 78, 125-9 (1998).

Polypeptide SEQ Incyte ID NO: Polype	ptide ID	GenBank ID NO: Probability or PROTEOME Score ID NO:		Amotation
,		732665 Echs1	2.IE-89	Mus musculus] Protein with strong similarity to strort chain enoyl-Coenzyme A hydratase 1 (rat Echs1), which catalyzes the hydration of trans-2-enoyl-CoA in mitochoodrial fatty acid beta-exidation, member of the enoyl-CoA hydratase or isomerase family
· ·	7511057CD1	87159801 2.8E-59 599822 LOC5586 1.2E-56	2.8E-59 1.2E-56	[Homo septems] d1551k20.2.1 (novel encoyl cooknoyl cook hydratuse/detydrogenase type represent selection.  Home septems [1], year, Oxidoreductase] Uncharacterized hypothalmus protein HCDASE, amenher of the encoyl-Cook hydratase/snoarese family, which function in fatty add an ententer of the encoyl-Cook hydratase/snoarese family, which function in fatty add
		1		metabolism
6	7511078CD1	g18478646 3.1E-168 731657/FLJ1449 2.5E-169 7	3.1E-168 2.5E-169	[Homo sapions] p33-responsive gene 3 The most superal Member of the pyridine medeoide-disulfide oxidoreductase family of Rivoproteins, has week similarity to C. albicans orfic.6088
		641934 orf6.6088 2.1E-18	2.1E-18	[Candida albicans][Oxidoreductase] Member of the pyridine nucleotide-disulphide oxidoreductase family of flavoproteins, has low similarity to uncharacterized S. cerevisine Ymr074a
				Huang, S. et al., Specificity of cotranslational amino-terminal processing of proteins in yeast., Biochemistry 26, 8242-6 (1987).
10	7511505CD1	g4106362	1.4E-251	Homo sapiens Jdatydrockolesterol reductase Waterham, H. R. et al., Smith-Lemli-Opizz syndrome is caused by mutations in the 7- dehydrocholesterol reductase gene, h. Him. Genet. 63, 329-338 (1998) dehydrocholesterol reductase gene, h. Him. Genet. 63, 329-338 (1998)
-		,		Yu, H. et al., Spectrum of Deftal/Jednytrocholossico) Foundates an analysis a parametric the smith-lemil-opitz (RSH) syndrome, Hum. Mol. Genet. 9, 1385-1391 (2000)
		335038 DHCR7 5.0E-252	5.0E-252	Home superal[Originatelase] Depolprisme relations: Cytoplasmic Dispolified membrane] Adelty depolprisme relations: Cytoplasmic Dispolified of membrane] Adelty develociolescon (reductor, catalyzes the reduction of the C7-C8 (data A) double bond of 7-dehydrocholescurol in the last stop of cholescurol biosynthesis; mutations in the corresponding gene are associated with Smith-Lemii-Optix syndrome

Table 2

	邑 一	o z	a 24.		П		e e		to to	18,
Amotation	Kim, J. H. et al., Cholesterol biosynthesis from lanosterol. A concerted role for Spl. and NF-Y-binding stee for succl-mediated regulation of raf 7-dahydrocholesterol reductase gene expression, J Biol Chem 276, 1815-561, (2001).	[Mas muscalus][Oxidoreductase] 7-dehydrocholestreol reductase, catalyzes the reduction of the CJ-C8 (delta, 7) double board of 7-dehydrocholestreol in the last step of cholestreol the CJ-C8 (delta, 7) double board of 7-dehydrocholestreol in the last step of cholestreol capacity and easily and the human DHCR7 gene are associated with Smith-Lemil-Optiz condonner.	Wassif, C. A. et al., Biochemical, phenotypic and neurophysiological characterization of a Wassif, c. A. et al., Biochemical, phenotypic syndrome., Hum Mol Genet 10, 535-64. (2001).	Kim, J. H. et al. (supra)	[Homo sapiens] aldehyde dehydrogenase	Hsu, L. C. et al., Genomic structure of the human cytosolic aldehyde dehydrogenase gene, Genomics 5, 857-865 (1989)	Fibron segriesal (Oxidoreductases Small molecule-birtining protein) [Cytoplasmic] Addebyde daylogenase I hanly member AI, a broad-specificity ofrostolic NAD-dependent enzyme that oxidizes addebydes to carboxylic acids, oxidizes retinal to retinoic acid, detoxifies xenobiotics, and contributes to cyclophosphamide resistance	Magni, M. et al., Induction of cyclophosphamide-resistance by aldehyde-dehydrogenase sene transfer Blood 87, 1097-103. (1996).	[Buts muscrilus][Oxidoroductaso][Cytoplasmic] Aldebyde debydrogenase 1 family member Al. 1 broad specificity cytosolite NAL1 a broad specificity cytosolite NAL2-dependent enzyme flatt oxidizes aldebydes to acrossyme alea, oxidizes retinal to retinoie acid, detoxifies xerobiotics, and contributes to evel-obsorbamide resistance	Greene, W. K. et al., The T-cell oncogenic protein HOX11 activates Addit expression in NIH 313 cells bur represses its expression in mouse spleen development, Mol Cell Biol 18, 7009-7 (1998).
		1.7E-227			3.3E-113		8.9E-114		2.4E-95	
GenBank ID NO: Probability or PROTEOME Score ID NO:		583989 Dhcr7			g178372		334128 ALDH1A 8.9E-114		586863 Aldh1a1 2.4E-95	
ptide ID					7511552CD1					
Polypeptide SEQ Incyte ID NO: Polype					11					

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Amotation	[Brucella melitensis] HYDROXYPYRUVATE ISOMERASE	DeIVecchio, V. G. et al., The genome sequence of the facultative intracellular pathogen	Iffucella incluents, r loc. tvat. Acad. Sci. C. Str. Str. Str. Str. Str. Str. Str. Str	elvoxylate-induced protein	Reinke, V. et al., A global profile of germline gene expression in C. elegans., Mol Cell 6,	605-16 (2000).	[Homo sapiens] squalene synthetase	McKenzie, T. L. et al., Molecular cloning, expression, and characterization of the cDNA for	the rat hepatic squalene synthase, J. Biol. Chem. 267, 21368-21374 (1992)	Jiang, G. et al., Transcriptional regulation by lovastatin and 25-hydroxycholesterol in	HepG2 cells and molecular cloning and expression of the cDNA for the human hepatic	squalene synthase, J. Biol. Chem. 268, 12818-12824 (1993)	[Homo sapiens][Transferase] Squalene synthase (farnesyl-diphosphate farnesyltransferase 1), catalyzes the conversion of farnesyl diphosphate to squalene in cholesterol biosynthesis,	may be a potential target for cholesterol lowering therapy	Tozawa, R. et al., Embryonic lethality and defective neural tube closure in mice lacking	squalene synthase., J Biol Chem 274, 30843-8 (1999).	[Mus musculus][Transferase] Squalene synthase (famesyl-diphosphate famesyltransferase	<ol> <li>catalyzes the conversion of ramesyl unanosphare to squarent in choiceach orosphares.</li> </ol>	Sakakura, Y. et al., Sterol regulatory element-binding proteins induce an entire pathway of	cholesterol synthesis., Biochem Biophys Res Commun 286, 176-83. (2001).	[Homo sapiens] methyltransferase	
Probability Score	4.7E-41		1 GE-34	1.05-34			3.2E-190						2.6E-191				3.1E-171				1.2E-44	
GenBank ID NO: Probability or PROTEOME Score ID NO:	g17985315		226614[C05[D11 1 6E-34	238014 CU3D11.			g307432						340542 FDFT1				584775 Fdft1				g2911587	
ptide ID	7511722CD1						7511489CD1														7511497CD1	l
Polypeptide SEQ Incyte ID NO:	12						13														14	

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Polypeptide SEQ Incyte ID NO: Polype	ptide ID	GenBank ID NO: Probability or PROTEOME Score ID NO:	pility	Аподалол
				Stanchi, F. et al., Characterization of 16 novel human genes showing high similarity to yeast
				sequences, Yeast 18, 69-80 (2001)
		240044 C27F2.4 3.6E-23	3.6E-23	[Caenorhabditis elegans][Unknown] Putative ortholog of S. cerevisiae Y cr04/p, a protein
	_			with similarity to rat methylglycine transferase
				Wang, S. P. et al., Structure-function analysis of the mRNA cap methyltransferase of
				Saccharomyces cerevisiae., J Biol Chem 272, 14683-9 (1997).
		2051 BUD23	9.7E-21	[Saccharomyces cerevisiae][Transferase] Protein with similarity to rat methylglycine
				transferase, may be involved in bud site selection/bipolar budding
				Moerschell, R. P. et al., The specificities of yeast methionine aminopeptidase and
				acetylation of amino-terminal methionine in vivo. Processing of altered iso-1-cytochromes or
				Lealed of organizations removed the second s
15	7511498CD1	g16226067	2.2E-114	[Homo sapiens] putative methyltransferase WBMT
2	1	240044IC27F2.4 4.0E-63	4.0E-63	[Caenorhabditis elegans][Unknown] Putative ortholog of S. cerevisiae Ycr047p, a protein
				with similarity to rat methylglycine transferase
				Wang, S. P. et al. (supra)
		2051BUD23	5.8E-55	[Saccharomyces cerevisiae] [Transferase] Protein with similarity to rat methylglycine
		-		transferase, may be involved in bud site selection/bipolar budding
				Slonimski, P. P. et al. (supra)
				Moerschell, R. P. et al. (supra)
16	7511612CD1	g12653701	3.2E-67	[Homo sapiens] acetyl-Coenzyme A acyltransferase 1 (peroxisomal 3-oxoacyl-Coenzyme A
1				thiolase)
		333998ACAA1 2.6E-68	2.6E-68	[Homo sapiens][Transferase][Cytoplasmic; Peroxisome] Peroxisomal 3-oxoacyl-coenzyme
				A thiolase (peroxisomal 3-ketoacyl-Coenzyme A thiolase), functions in peroxisomal peta-
				oxidation of fatty acids, deficiency is associated with rhizomelic chondrodysplassa punctata
				(RCDP)

Polypeptide SEQ Incyte	Incyte	GenBank ID NO: Probability	Probability	Annotation
ID NO:	Polypeptide ID	or PROTEOME Score ID NO:	Score	
				Heikoop, J. C. et al., Rhizomelic chondrodysplasia punctata. Deficiency of 3-excaeryl- cenzyme. A thiolase in peroxisomes and impaired processing of the enzyme. J. Clin Invest 86. 126-30, (1990).
		430496 Acaa	2.9E-51	[Rattus norvegicus] [Transferase] [Cytoplasmic; Peroxisonue] Peroxisonal 3-ketonacy-CoA thionse A. (1), catalyzes the firm I step in the rept sold between expression is only weakly networked by administration of peroxisone proliferators
				Hijikata, M. et al., Rat peroxisomal 3-ketoacyl-CoA thiolase gene. Occurrence of two closely related but differentially regulated genes., J Biol Chem 266, 4600-6 (1990).
17	[7511624CD1	g179104	1.6E-199	[Homo sapiens] aspartate aminotransferase precursor (2.6.1.1)
		0		Pol. S. et al., Nucleotide sequence and itsue distribution of the human mitochondrial asparate aminorumsferase mRNA, Biochem. Biophys. Res. Commun. 157, 1309-1315 (1989).
		343324 GOT2	1.3E-200	[Homo supierse][Transfearse][Cytoplasmie; Mirochondrial] Mirochondrial aspurate aminoransfearse (glutamic oxaloacete transaminase), catalyzes the reversible transfer of the amino group from aspurate to 2-oxoglutarate to form oxaloacetate and glutamate
				Salomon, A. R. et al., Understanding and exploiting the mechanistic basis for selectivity of polyketide inhibitors of POFI-ATPase, Proc Natl Acad Sci U S A 97, 14766-71 (2000).
		584895 Got2	1.1E-192	[Mus musculus][Transferase][Cytoplasmic; Mitochondrial] Mitochondrial aspartate aminoransferase, cashyes the reversible travière of the amino group from aspartate to 2-coordinates to form oxfolocetute and guinante.
				Memon, R. A. et al., Regulation of putative fatty acid transporters and Acyl-CoA synthetase in liver and adipose tissue in ob/ob mice., Diabetes 48, 121-7 (1999).
18	7511626CD1	g179104	1.1E-209	[Homo sapiens] aspartate aminotransferase precursor (2.6.1.1)  Pol, S. et al. (supra)

Table 2

Polypeptide SEQ Incyte ID NO: Polype	ptide ID	GenBank ID NO: Probability or PROTEOME Score ID NO:	bility	Amotation
		343324 GOT2	8.9E-211	[Homo sapiens][Transferaso][Cyroplasmic; Mitochondrial] Mitochondrial aspartite aminormasferase (glutamic oxaloacetic transaminase), catalyzes the reversible transfer of the amino group from asparate to 2-cooglutarate to form oxaloacetate and glutamate
				Salomon, A. R. et al. (supra)
		584895 Got2	4.2E-202	[Mus musculus][Transferase][Cytoplasmic; Mitochondrial] Mitochondrial aspartate aminotransferase, catalyzes the reversible transfer of the amino group from aspartate to 2-
				oxoglutarate to form oxaloacetate and glutamate  Memon, R. A. et al. (supra)
19	7512885CD1	569696  DKFZp5660084	6.1E-138	[Homo sapiens][Oxidoreductass] Member of the short-chain dehydrogenass/reductase family
		251240 T25G12. 7.0E-28	7.0E-28	[Caenoriabditis elegans][Oxidoreductase] Putative dehydrogenase, has strong similarity over the N-terminal region to C. elegans T25G12.7
				Bateman, A. et al., Pfam 3.1: 1313 multiple alignments and profile HMMs match the majority of proteins., Nucleic Acids Res 27, 260-2 (1999).
		754109 Hsd17b1 2.7E-18	2.7E-18	[Mus musculus] Membor of the short-chain dehydrogenase-reductase family, which are NAD- or NADP-dependent oxidoreductases, has low similarity to short-chain.
				dehydrogenase-reductase 1 (numan SDK1), which catalyzes the reduction of air-datast contents to retinol
20	7511965CD1	g3150035	1.9E-146	[Homo sapiens] aldose reductase-like peptide  Cao, D. et al., Identification and characterization of a novel human aldose reductase-like
				gene, J. Biol. Chem. 273, 11429-11435 (1998)
		690816 AKR1B1 5.0E-147	5.0E-147	[Homo sapiens][Oxidoreductase] Aldose reductase-like protein (human small intestine
				detoxify reactive digested aldehydes or act in steroid metabolism, overexpressed in some
				liver cancers

Polypeptide SEQ Incyte ID NO: Polype	ptide ID	GenBank ID NO: Probability or PROTEOME Score ID NO:	oility	Amotation
				Lefrancois Martinez, A. M. et al., Product of side-chain cleavage of cholesterol, iso-aproaldebyde, is an endogenous specific substrate of mouse vas eleferous protein, an aldose reductuse-like protein in adrenocurical cells., J Biol Chem 274, 32875-80 (1999).
	7511965CD1	589811 Akr1b1	9.6E-110	[Ratus norvegicus][Oxidoreductase] Aldose recircuse, member of the NADPH-dependent addo-keto reductase superfamily, reduces glucose and other carbonyl-containing substrates as part of the polyol pathway, metabolizes xenobiotics, and may contribute to diabetic complications.
				Takahashi, M. et al., Elevation of aldose reductase gene expression in rat primary begatoma and bepatoma and lenserian cell three: implication in detoxification of cyrotoxic alidehydes, Int J Cancer 62, 7465-54 (1992).
21	7512403CD1	g1870244	7.1E-197	[Homo sapiens] aldehyde dehydrogenasc
				Chang, C. et al., Human fatty aldehyde dehydrogenase gene (ALDH10): organization and tissue-dependent expression, Genomics 40, 80-85 (1997)
		339660 ALDH3A   5.6E-198		Homo supieus [I Oxidoreductase] Aldahyde dehytogenuse 3 family member A2, catalyzes the oxidation of long chain fatty aldehydes and leukoricness; mutation of the corresponding causes Stogen Lanson syndrome, a disorder marked by mental retardation, spisiticity, and ichthyosis.
				Kraus, C. et al., RNA-based mutation screening in German families with Sjogren-Larsson syndrome, Eur J Hum Genet 8, 299-306. (2000).
		711692 Aldh3a2   6.7E-167		Ratus norvegious [Oxidorelucuese] [Endoplasmic reticulum; Cytoplasmic; Plasma membrane] Aldebyed elaylungeanse family 3 subfamily A2, catalyzes the oxidation of long and medium chain eliphatic aldebydes derived from ligid metabolism; mutation of the luman ALDH3A2 gene cuasse Sjogen Larson syndrome
				Rizzo, W. B. et al., Fatty aldehyde dehydrogenase: genomic structure, expression and mitation analysis in Sjegren-Larson syndrome., Chem Biol Interact 130, 297-307. (2001).

# Table 2

				1.7
Annotation	[Homo sepiens] androgen-regulated short-chain dehydrogenase/reduciase 1  11. Re of al., Prostate short-chain dehydrogenase-reduciase 1 (PSD/RL): a new marther of the short-chain steroid dehydrogenas-erhodicuse family highly expressed in normal and mooplassic prostate opithelium, Cancer Res. 61, 1611-1618 (2001)	[Homo sapiens] Protein containing EGF-like domains, which are found in some secreted proteins and extractellular domains of transmendrane proteins proteins and extractellular domains of transmendrane proteins. [Homo sapiens] Member of the since claim of adolydrogenase-reductase family, which are NAD- or NAD-dependent oxidoreductases, has low similarity to 17 beta-hydroxysteroid delabydrogenase type 7 (mouse Had17b7), which converts estrone to estradiol in the corpus linears.	[Homo supiosa] peroxitedoxin 3. a mitochondrial antioxidant protein flamo supiosal peroxitedoxin 3. a mitochondrial antioxidant protein involved in oxygen and radical metabolism, has an antiapoproteic function involved in oxygen and radical metabolism, has an antiapoproteic function protein. J Biol Chem 276, 21870-7, (2021).  [Mus musculus] Peroxitedoxin 3. a putative antioxidant protein, has ally! hydroperoxide protein. Then, it involved in red ifferentiation of inflementation of the antioxidant protein and antioxidant protein and antioxidant protein and includence activity, involved in red ifferentiation of the protein and antioxidant protein complements ally/hydroperoxide reductase effects.	[Homo suptores] Gut 44-3-coxostoral ben-reuched by the CDNA of human delin 4-3-oxostoral 5 Kondo, K. He at., Cloning and expression of cDNA of human delin 4-3-oxostoral 5 Kondo, K. He at., Cloning and expression of cDNA of human delin 4-3-oxostoral 219-357-367 (1994)  150 (1994)  Homo supteral [Dristoredurense] Delin 4-3-oxostoral 5 beta-reductase, catalyzes the Floron supteral [Dristoredurense] Delin 4-3-oxostoral 5 beta-reductase, catalyzes the reduction of the Delin 4 doubte bond of bile acid intermediates and steroid hormones; deficiency results in neonatal liver failure with associated homodromatosis
Probability	6.7E-76	5.2E-77 7.0E-42	2.1E-95 2.1E-96 1.1E-78	4.0E-144 3.1B-145
GenBank ID NO: Probability or PROTEOME Score ID NO:	g9622124 (	475723 LOC5110 5.2E-77 9 613361 PAN2 7.0E-42	g18203832 2.7E-95 428166[PRDX3 2.1E-96 583587[Prdx3 1.1E-78	8431857 4.0E-144 343204 AKRIDI 3.1E-145
ptide ID	7512564CD1		7512646CD1	7512700CD1
Polypeptide SEQ Incyte ID NO:	22		33	24

Table 2

Amotation	Smeider, B. L. et al., Delta 4-5 constenoid 5 bear-eductuse deficiency causting necontail Simeider, B. L. et al., Delta 4-5 constenoid 5 bear-eductuse deficiency causting necontail reference for the construction of the Delta 4-5 construction of the construction of the Delta 4 double bond of this end intermediates and steroid hormones; reduciency of human ARRID1 results in monatual liver fullure with associated deficiency of human ARRID1 results in monatual liver fullure with associated benochromatosis.	Kondo, K. H. et al., Clouing and expression of cDNA of human delia 4-3-exosterou'd 20 Bear- reductase and substrate specificity of the expressed enzyme., Eur J Biochem 219, 357-453 (1994).  [Residenments sarrientosal Probabioe PAD-dependent monocoxygenase	It sourcements to the state of the flavoprotein monooxygenase family of aromatic ring [Homo sapiens] Member of the flavoprotein monotoxygenase family of aromatic ring hydroxylases, has moderate similarity to uncharacterized C. elegans K07B1.2	[Caenorhabditis elegans][Oxidoreductase][Mitochondrial] Protein with strong similarity to S. cerevisiae Coqôp, a monocoygenase required for coenzyme Q (thiquinone) biosynthesis	Hill, A. A. et al., Genomic analysis of gene expression in C. elegans., Science 290, 809-12. (2000).	[Homo sapiens] Member of the flavoprotein monooxygenase family of acouatic ring hydroxyjaxes, has moderate similarity to uncharacterized C. Delegans KOTB1.2 hydroxyjaxes, has moderate similarity to uncharacterized C. Delegans KOTB1.2 hydroxyjaxes, has moderate similarity to uncharacterized for the control of the control	[Saccharomyces cerevisiae][Oxidoredudase][Unitocholidinal] aromotory general coenzyme Q (tibiquinone) biosynthesis	Ifan, A. Y. et al., Genetic evidence for a mutu-subanii comprox in taco ancory, steps of coemzyme Q biosynthesis., Biochim Biophys Acta 1484, 287-297 (2000).	[Homo sapiens] cysteine dioxygenase	[Homo supposts [Doudoroutense] Cysteme unalgement, camparation of saffine as a sufficient of saffine and the regulation of saffine production of saffine production
robability	5.6E-118	10H 33	3.4E-153	6.2E-42		1.5E-217	4.5E-54		7.9E-41	2.7E-41
GenBank ID NO: Probability or PROTEOME Score ID NO:	332522Rn.25716 5.6B-118	000000	89951529 1.9E-54 475553[LOC5100 3.4E-153	247658 K07B1.2 6.2E-42		475553 LOC5100 1.5E-217 4	7170 C0Q6		g1747331	334614 CDO1
ptide ID			7512707CD1			7512710CD1			7512884CD1	
Polypeptide SEQ Incyte ID NO:			25			56			27	

Table 7

oility	Touboyama-Kasaoka, N. et al., Human cysteine dioxy genase gene: structural organization, Issue appeare experience to generate the control to a control to a control to a control to the co	4.5E-39 [Rattus norvegicus][Oxidoreductase] Cysteine dioxygenase 1, catalyzes the conversion of cysteine to 3-sulfinoalanine in cysteine catabolism	Parsons, R. B. et al., Cysteine dioxygenase: regional expression of activity in rat oran., Neurosci Let 248, 101-4, (1998).	1.1E-242 [Homo sapiens] protoporphyrinogen oxidase	Dailey, T. A. et al., Expression of a cloned protoporphyrmogen oxidase, J. Biol. Cucui. 209, 813-815 (1994)	8.9E-244 [Homo sapiens][Outdoreductase][Cytroplasmic, Mitochondrial] Protoporphymogen oxidase, catalyzes the oxidation of protoporphymicogen IX to protoporphymic IX in heme biosynthesis, inhibited by the ternaphydrophthalimide and diplomyl ether herbicides; genetic munition is detected in patients with variegate porphyria	Whatley, S. D. et al., Variegate porphyria in Western Europe, identification of PPOX gene musations in 104 families, extent of allelic heterogeneity, and absence of correlation between phenotype and type of musation, An J Hum Genet (5, 984-94, (1999).	2.5E-215 [Mas musculus][Oxidoreductuse] Protoporphyrinogen oxidase, catalyzes the oxidation of protoporphyrinogen IX to protoporphyrin IX, inhibited by agents used in light treatment of lumns and by diphenyl ether herbicides; deficiency of humn PPOX is associated with variente contributa.	Finger, V. Le et al., Photodynamic therapy using a protoporphyrinogen oxidase inhibitor, Cancer Res 57, 4551-6. (1997).	3.5E-94 [Homo sapiens] protoporphyrinogen oxidase	
GenBank ID NO: Probability or PROTEOME Score ID NO:		751686 Cdo1 4.5		g837328   1.1		337132JPPOX 8.9		582571Ppox 25		2837328	
ptide ID				7512931CD1		,				7512933CD1	
Polypeptide SEQ Incyte ID NO:				28						20	177

protoporphyrinogen IX to protoporphyrin IX, inhibited by agents used in light treatment of protoporphyrinogen IX to protoporphyrin IX, inhibited by agents used in light treatment of biosynthesis, inhibited by the tetrahydrophthalimide and diphenyl ether herbicides; genetic biosynthesis, inhibited by the tetrahydrophthalimide and diphenyl cther herbicides; genetic Mus musculus][Oxidoreductase] Protoporphyrinogen oxidase, catalyzes the oxidation of [Mus musculus][Oxidoreductase] Protoporphyrinogen oxidase, catalyzes the oxidation of oxidase, catalyzes the oxidation of protoporphyrinogen IX to protoporphyrin IX in heme oxidase, catalyzes the oxidation of protoporphyrinogen IX to protoporphyrin IX in heme tumors and by diphenyl ether herbicides; deficiency of human PPOX is associated with tumors and by diphenyl ether herbicides; deficiency of human PPOX is associated with Persoxisomal 2-Hydroxy Acid Oxidases, J. Biol. Chem. (2000) 275(17):12590-12597 Homo sapiens][Oxidoreductasc][Cytoplasmic; Mitochondrial] Protoporphyrinogen [Homo sapiens][Oxidoreductase][Cytoplasmic; Mitochondrial] Protoporphyrinogen Jones, J. M. et al., Identification of HAOX1, HAOX2, and HAOX3, Three Human [Homo sapiens] long-chain 2-hydroxy acid oxidase HAOX2 mutation is detected in patients with variegate porphyria nutation is detected in patients with variegate porphyria [Homo sapiens] protoporphyrinogen oxidase Whatley, S. D. et al. (supra) Whatley, S. D. et al. (supra) Fingar, V. H. et al. (supra) Fingar, V. H. et al. (supra) Dailey, T. A. et al. (supra) variegate porphyria variegate porphyria Annotation Probability 8.1E-146 9.1E-56 7.1E-57 5.7E-48 2.8E-95 6.9E-82 Score or PROTEOME ID NO: GenBank ID NO: 337132PPOX 337132|PPOX 582571|Ppox 582571|Ppox g7208438 g837328 Polypeptide ID 7513736CD1 7512942CD1 Polypeptide SEQ Incyte

			Belmouden, A. et al., Molecular cloning and molecitide sequence of QDNA encoding rat kidney long-chain L-2-thydroxy acid oxidas. Expression of the eathyrically active recombinant protein as a chimaera., Eur J Biochem 214, 17-25 (1993).	Homo suplents  cysteine sulfinic acid decarboxylase-related protein 3   Ratus norvegicus [Lyase] Cysteine sulfinic acid decarboxylase, an enzyme involved in the bioxymbists of turner, expressed by the proposed and estrogen, and control acid turner of turner and estrogen, and control acid turner beneficiaries.	Expression is summand out up a representation of Kishimoto, T. et al., Owerexpression of cysteine sulfinic acid decarboxylase stimulated by impeptocarcinogenesis results in autoantbody production in rats., Cancer Res S6, 5230-7 (1996).	14 [Hono sapiens [Lyase] Glutamate decarboxylase (Legittamate 1-carboxy-lyase), converts Liguramic acid to gamma-aminobuyric acid; an autounitgen in insulin-dependent diabetes mellinus, and stiff-man syndrome; murine Gad2 is associated with epilepsy	Mally, M. I. et al., Ontogeny and tissue distribution of human GAD expression., Diabetes 45, 496-501 (1996).	3 [rHomo sarriens] 3-hvdroxyacyl-CoA dehydrogenase
Probabil Score	6.4E-14	7.7E-108		4.7E-217 6.8E-203		7.0E-114		1 11 43
GenBank ID NO: Probability or PROTEOME Score ID NO:	475383 HAOX2 6.4E-147	747907 Hao3		g4894560 624468 Csad		335440 GAD2		-1402511
ptide ID				7512779CD1				1 COTTOCION
Polypeptide SEQ Incyte ID NO:				32				

	-8- -8-	in the	1ase 108,			2-201	si uois	eptic	nic	
Amotation	Vredendanl, P. I. et al., Human short-chain L3-hydroxyacyl-CoA dehydrogenses: cloning and characterization of the coding sequence, Biochem. Biophys. Res. Commun. 223, 718-792 roar stoops.	Homo supplement   Homo suppl	Papiloxyou-typeanim: Papiloxyou-typeanim: Alpperinsulinism in short-chain L.3-hydroxyacyl-CoA debydrogenase Geficiency reveals the importance of beta-oxidation in insulin secretion. J Clin Invest 108, 4eef.ec. 2001.	437-03. (2001). Protein Data Bankl L-3-Hydroxyacyl-Coa Dehydrogenase	[Homo sapiens] quinolinate phosphoribosyl transferase	Fukuoka, S. I. et al., Characterization and functional expression of the cDNA encoding human brain quinolinate phosphoribosyltransferase, Biochim. Biophys. Acta 1395, 192-201.	(1998) [Homo sapiens][Transferase] Quinolinate phosphoribosyltransferase, catabolizas quinolinate, an intermediate in NAD synthesis and a neuron extitotoxin, excess expression is associated, with neurodegenerative disorders and decreased activity may play a role in	epilepsy Feldblum, S. et al., Quinolinic-phosphoribosyl transferase activity is decreased in epileptic	human brain tissue, Epilepsas 29, 225-9, 14989). [Saccharomyoss czevisiae][Transferase] Putative quinolinic acid phosphorobystransferase, active in the pathway of tryptophan degradation and nicotinic conf combrosis.	men of the special guinolinate phosphoribosyl transferase Fuktooka, S. I. et al. (supra)
	7 S <		0.4	60F-44			1.0E-55		1.76-31	2.5E-53
GenBank ID NO: Probability or PROTEOME Score ID NO:		342104 HADHS 8.6E-45 C		70/21/25 1-F10 A	Ш		569398 QPRT		9762 YFR047C	g1060907
ptide ID					7512782CD1					7512784CD1
Polypeptide SEQ Incyte ID NO:					34	ţ.				35

		-		<b>—</b>	-		0	_	Te	,	-		1				•	
Amotation	[Homo sapiens][Thansferase] Quinolinate phosphorhoxyltrunsferase, cataboblizes quinolinate, an intermediate in NAD synthesis and a neuron extinouxin; excess expression is seasoned with neurodegenerative disorders and decreased activity may play a role in epilepsy	Feldblum, S. et al. (supra)	[Saccharomyces cerevisiae][Transferase] Putative quinolinic acid phosphoribosyltransferase, active in the pathway of tryptophan degradation and nicotinic	acid synthesis	[Homo sapiens] S-adenosylmethionine decarboxylase proenzyme (EC 4.1.1.50) old gene name 'AMD'	Pajunen, A. et al., Structure and regulation of mammalian S-adenosylmethionine Arcarbov lose. J. Biol. Chem. 263, 17040-17049 (1988)	[Homo sapiens][Lyase] S-adenosylmethionine decarboxylase 1, catalyzes the removal of the	carboxylate group of S-adenosylmethionine to form S-adenosyl-5'-3-methylpropylamine in	the polyamine biosynthesis pathway	Bettuzzi, S. et al., Tumor progression is accompanied by significant changes in the levels of	expression of polyamine metabolism regulatory genes and clusterin (sunated glycopiotenin	<ol> <li>in human prostate cancer specimens (published effatum appears in Cancer New Zood and 1: 60(5):14721 Cancer Res 60, 28-34 (2000).</li> </ol>	Homo sapiens] PIG-L	Watanabe, R. et al., Mammalian PIG-L and its yeast homologue Gpi12p are N-	acetylglucosaminylphosphatidylinositol dc-N-acetylases essential in	glycosylphosphatidylinositol biosynthesis, Biochem. J. 339 (Pt 1), 185-192 (1999)	[Homo sapiens][Hydrolase] Protein with high similarity to N- acertolehroceaminyInhosnhatidvlinositol de-N-acetylase (rat Rn.10774), which catalyzes the	second step in GPI anchor synthesis
Probability Score	2.0E-54		3.0E-24		1.9E-15		1.5E-16						9 OF-88				7.1E-89	
GenBank ID NO: Probability or PROTEOME Score ID NO:	569398 QPRT		9762 YFR047C 3.6B-24		g178518		343256 AMD1						230086	0			341062[PIGL	
ptide ID					7512794CD1								7517006CD1					
Polypeptide SEQ Incyte ID NO:					36								te	100				

[Rattus norvegicus][Hydrolase] N-acetylglucosaminylphosphatidylinositol de-N-acetylase, catalyzes the N-deacylation of N-acetylglucosaminyl phosphatidylinositol (GlcNAc-PI) to Nakamura, N. et al., Expression cloning of PIG-L, a candidate N-acetylglucosaminyl-Smith, T. K. et al., Specificity of GlcNAc-PI de-N-acetylase of GPI biosynthesis and synthesis of parasite-specific suicide substrate inhibitors., Embo Journal 20, 3322-32. form glucosaminyl phosphatidylinositol (GlcN-PI) in the second step of GPI anchor phosphatidylinositol deacetylase, J Biol Chem 272, 15834-40 (1997). Table 2 biosynthesis Annotation (2001). GenBank ID NO: Probability 1.0E-64 Score or PROTEOME ( ID NO: 331106|Pigl Polypeptide SEQ Incyte
D NO: Polypeptide ID

184

[Mus musculus] UDP-glucuronic acid decarboxylase

1.0E-40

g15150358

7512929CD1

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Analytical Methods and Databases	SPSCAN	HMMER	HMMER_TIGRFAM	TMHMMER	MOTIFS	SPSCAN	BLAST_PRODOM	HMMER	HMMER PFAM	HMMER_PFAM	HMMER_TIGRFAM	TMHMMER
Signature Sequences, Domains and Motifs	Signal_oleavage: M1-L67	Signal Peptide: L46-L67	AGP_acyltm: 1-acyl-sn-glycerol-3-phosphate acyltransferases: L107-S237	Cytosolic domain: M1-F39; Transmembrane domain: C40-W62; Non-cytosolic domain: P63-P256	Leucine zipper pattern: L46-L67	Signal_cleavage: M1-A31	PROTEIN BIOSYNTHESIS ANTIBIOTIC SURFACTIN LIPOPERTIDE ANTIBIOTICS ITURIN A GSP PRODUCTION PRODUCTION PRO05239; 140-1130	Signal Peptide: M1-A28	Thiamine pyrophosphate enzyme, central: S269-L422	Thiamine pyrophosphate enzyme, N-terminus: S50-K221	menD: 2-succinyl-6-hydroxy-2,4-cyclohexadiene-1- carboxylic acid synthase/2-oxoglutarate decarboxylase: V46- F169	Cytosolic domain: M1-G11; Transmembrane domain: S12- TMHMMER
Potential Glycosylation Sites	N152					N108		N336				
ylation	SS S148 S171 S177 N152 S207 T131 T190					S29 T87		S50 S178 S367 S527 S571 S638 S648 S671 T95 T188 T299 T352 T432 T457				
no Acid dues	256											
ncyte Polypeptide D	7511289CD1 256					7511056CD1   138		7511567CD1 674				
SEQ Inc ID Pol NO: ID	_					2		3	1			

Table 3

Analytical Methods and Databases	BLIMPS_BLOCKS	PROFILESCAN	BLAST_PRODOM	BLAST_PRODOM	BLAST_DOMO	BLAST_DOMO	BLAST_DOMO	BLAST_DOMO	MOTIFS	SPSCAN	HIMIMER_PFAM
Signature Sequences, Domains and Motifs	Thiamine pyrophosphate dependent enzyme IPB000399: L62-C85, A105-A122, P330-S359, G494-A506, P511-G548	Thiamine pyrophosphate enzymes signature: P484-R542	THIAMINE PYROPHOSPHATE FLAVOROTEIN SYNTHASE LYASE PYRIVATE ACETOLACTATE DECARBOXYLASE ACID SUBUNIT PD000376: 1.62- 1539	ACETOLACTATE SYNTHASE FLAVOPROTEIN LYASE THIAMINE PYROPHOSPHATE HOMOLOG PD039458: ML-V61	THIAMINE PYROPHOSPHATE ENZYMES DM00391[P08142]10-559: G54-K230, E254-T533, P9-G37	THIAMINE PYROPHOSPHATE ENZYMES DM00391[p39994]1-559: E56-L213, P270-R536	THIAMINE PYROPHOSPHATE ENZYMES DM00391[P00893]1-563: G54-P215 Q272-1539 R561-R578	THIAMINE PYROPHOSPHATE ENZYMES DM00391P40149/7-564: L62-P209 Q256-1539	Thiamine pyrophosphate enzymes signature: L504-A523	Signal_cleavage: M1-A41	Uroporphyrinogen-III synthase HemD: K2-G226
Potential Glycosylation Sites										N50 N73	
Potential Phosphorylation Sites										S23 S35 S58 S176 N50 N73 T86 T200	
Amino Acid Potential Residues Phospho Sites										237	
yte ypeptide										7511651CD1 237	
SEQ Inc.										4	

Analytical Methods and Databases	BLAST_PRODOM	BLAST_DOMO	BLAST_DOMO	FS	4N	ER.	HIMMER PFAM	TMHMMER	BLIMPS_BLOCKS	BLAST_PRODOM	S	N.	3R
Analy and D	BLAS	BLAS	BLAS	MOTIFS	SPSCAN	HMMER	HIMIN		BLIM	BLAS	MOTIFS	SPSCAN	HMMER
Signature Sequences, Domains and Motifs	UROPORPHYRINOGENIII SYNTHASE UROS UROPORPHYRINOGEN III COSYNTHETASE HYDROXYMETHYLBLANG HYDROLYASE CYCLIZING UROIIIS PD017488. M.IR21 S3-1227	UROPORPHYRINOGEN-III SYNTHASE DM07953[P10746]1-264: M1-R21 R21-C237	UROPORPHYRINOGEN-III SYNTHASE DM07953[P51163]1-264; M1-R21 R21-C237	Aldehyde dehydrogenases cysteine active site: L84-E95	Signal_cleavage: M1-L18	Signal Peptide: M1-A20	MAPEG family: P44-A161	Cytosolic domains: Q25-S57, Q123-A142; Transmembrane domains: V5-L24, E38-V80, C100-F122, L143-A165; Non-cytosolic domains: MI-E4, G81-P99, A166-A178	MAPEG (Membrane-associated proteins in eicosanoid and glutathione metabolism) IPB001129: G42-G79	TRANSMEMBRANE LEUKOTRUENE BIOSTNTHESIS PROTEIN TRANSPERANS 51, POXY GENASE ACTIVATING FLAP MK88/GBINDING MICROSOMAL PD009387: MI-H75 A107-1173	FLAP/GST2/LTC4S family signature: G42-C56	Signal_cleavage: M1-V17	Signal Peptide: M1-S15
Potential Glycosylation Sites					NSS							N29	
Potential Phosphorylation Sites					S28 S139							S50 S138 S165 S191 S208 T157	
Amino Acid Potential Residues Phosphor Sites					178							245	
SEQ Incyte ID Polypeptide NO: ID					7511881CD1							7512181CD1	
SEQ NO:					5							9	

Analytical Methods and Databases	HMMER	HMMER	HMMER PFAM	TMHMMER		BLIMPS_PRODOM	BLAST_PRODOM			BLAST PRODOM				BLAST DOMO		BLAST_DOMO		HMMER PFAM		BLIMPS_BLOCKS	PROFILESCAN
Signature Sequences, Domains and Motifs	Signal Peptide: M1-G18	Signal Peptide: M1-A23	Arylesterase: G2-G245	Cytosolic domain: M1-L4; Transmembrane domain: V5-F24; TMHMMER	Non-cytosolic domain: R25-G245	SERUM PARAOXONASE/ARYLESTERASE P. PD02637: BLIMPS_PRODOM R32-P86, F106-V140, E141-1169, V170-K217, G218-G245	SERUM PARAOXONASE/ARYLESTERASE PON	ARTICULAR TERHOSPHATASE AESTERASE	AROMATIC ESTERASE HYDROLASE GLYCOPROTEIN Signal PD005046: F49-v233	PARAOXONASE/ARYLESTERASE SERUM PON	ARYLDIAKYLPHOSPHATASE AESTERASE	AROMATIC ESTERASE HYDROLASE	GLYCOPROTEIN Signal PD005529; M1-E46	PARAOXONASE; ARYLESTERASE; SERUM;	DM07178 P54832 1-353: M1-Y233	PARAOXONASE; ARYLESTERASE; SERUM;	DM07178 P27169 1-353; K3-Y233	Enoyl-CoA hydratase/isomerase family: G48-E214		Enoyl-CoA hydratase/isomerase IPB001753: IS0-L61, G83-BLMPS_BLOCKS N105, K128-A154, G175-E214	Enoyl-CoA hydratase/isomerase signature: K118-T169
Potential Glycosylation Sites																		N44 N105			
Potential Phosphorylation Sites																		S107 S113 S199	S216 T88 T176 T211		
Amino Acid Potential Residues Phospho Sites																		250			
Incyte Polypeptide ID																		7511726CDI 250			
2 2 3 3 3 3 3 3 3 3 3 3 3 3 3 3 3 3 3 3														_	j		~	_			

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Analytical Methods and Databases	BLAST_PRODOM	BLAST_PRODOM	BLAST_DOMO	BLAST_DOMO	BLAST_DOMO	BLAST_DOMO	MOTIFS	BLIMPS_BLOCKS	SPSCAN	HMMER_PFAM	TMINMER	BLIMPS_PRINTS
Signature Sequences, Domains and Motifs	PROTEIN HYDRATASE ENOYLCOA ACID FATTY LYASE ISOMERASE METABOLISM 3HYDROXYACYLCOA DEHYDROGENASE PD000432: V47-A203	HYDRATASE ENOYLCOA MITOCHONDRIAL PRECURSOR SHORT CHAIN SCEH FATTY ACID METABOLISM PD000523: MI-G48	ENOYL-COA HYDRATASFISOMERASE DM00366[P30084]34-285: E34-A206, A200-N246	ENOYL-COA HYDRATASE/ISOMERASE DM00366[P34559]32-283: E34-Q205	ENOYL-COA HYDRATASEISOMERASE DM00366[P52046]1-255: 136-L218	ENOYL-COA HYDRATASE/ISOMERASE DM00366[137195]76-334: K43-K204, L187-R243	Enoyl-CoA hydratase/isomerase signature: 1131-1151	Enoyl-CoA hydratase/isomerase IPB001753: L61-F72, K94 BLIMPS_BLOCKS S116	Signal_cleavage: M1-A26	Pyridine nucleotide-disulphide oxidoreductase: H13-Y256 HMMER_PFAM	Cytosolic domain: M1-G10, Transmembrane domain: A11-L33; Non-cytosolic domain: N34-P333	FAD-dependent pyridine nucleotide reductase signature PR00368: H13-V35, V199-N208, F104-E129, N197-A211, V341-A347
Potential Glycosylation Sites								N89	N208			
Potential Phosphorylation Sites								S12 S107	S147 S166 S170 S272 S324 T72 T188 T251 T327			
Amino Acid Potential Residues Phosphor Sites								145	333			
SEQ Incyte ID Polypeptide NO: ID								7511057CD1 145	7511078CD1 333			
SEQ Inc ID Pol NO: ID ID								, ,	0			

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Analytical Methods and Databases	HMMER_PFAM	TMHMMER		BLIMPS BLOCKS	1	BLAST PRODOM				BLAST_PRODOM			BLAST_PRODOM	BLAST_DOMO		BLAST_DOMO		BLAST_DOMO		BLAST_DOMO	
Signature Sequences, Domains and Motifs	Ergosterol biosynthesis ERG4/ERG24 family: P7-F443	Cytosolic domains: M1-A118, F169-H231, N323-M381;	Transmembrane domains: W119-I141, I146-Y168, S232-W254, L300-A322, G382-M404; Non-cytosolic domains: F142-W145, N255-0299, A405-F443	Ergosterol biosynthesis ERG4/ERG24 enzymes IPB001171: BLIMPS BLOCKS	G181-R196, K199-V234, E256-P297, G315-R330, C348- C390, G391-F443	REDUCTASE STEROL TRANSMEMBRANE	OXIDOREDUCTASE BIOSYNTHESIS C14	C14REDUCTASE PROTEIN LAMIN B PD004179: L67-	L396, A389-F443	REDUCTASE STEROL TRANSMEMBRANE	OXIDOREDUCTASE BIOSYNTHESIS C14 LAMIN B	RECEPTOR C14REDUCTASE PD149902: Y398-F443	DELTA7STEROL REDUCTASE OXIDOREDUCTASE PD140411: M1-A32, C34-T61	ERGOSTEROL BIOSYNTHESIS ERG4/ERG24 FAMILY BLAST_DOMO	DM01860 A53616 196-614: L67-F443	ERGOSTEROL BIOSYNTHESIS ERG4/ERG24 FAMILY BLAST_DOMO	DM01860 P23913 190-607: K63-Y436	ERGOSTEROL BIOSYNTHESIS ERG4/ERG24 FAMILY BLAST_DOMO	DM01860JrC4057/3-423: Y111-F443, A48-R330	ERGOSTEROL BIOSYNTHESIS ERG4/ERG24 FAMILY BLAST_DOMO	DMULGOOK 300/911-409: L0/-6391, A309-1443
Potential Glycosylation Sites	N220 N255																				
Potential Phosphorylation Sites	S81 S173 T61 T351 N220 N255 T409 Y350 Y430																				
Amino Acid Potential Residues Phosphor Sites	443																				
Incyte Polypeptide ID	7511505CD1 443																				
SEQ Incyte ID Polype NO: ID	10																				

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Analytical Methods and Databases	BLIMPS_BLOCKS	BLAST_PRODOM	BLAST_DOMO	BLAST_DOMO	BLAST_DOMO	BLAST_DOMO	HMMER	HMMER	BLAST_PRODO		BLAST_DOMO	BLAST_DOMO	BLAST_DOMO	BLAST_DOMO
Signature Sequences, Domains and Motifs	Aldehyde dehydrogenase family IPB002086: P159-P172	DEHYDROGENASE OXIDOREDUCTASE ALDEHYDE NAD PROTEIN CLASS SEMIALDEHYDE PRECURSOR TRANSIT PEFTIDE PD000218: E28-E216	ALDEHYDE DEHYDROGENASES GLUTAMIC ACID DM00100JP0035217-471: 118-A212	ALDEHYDE DEHYDROGENASES GLUTAMIC ACID DM00100 P15437 17-471: Q19-A212	ALDEHYDE DEHYDROGENASES GLUTAMIC ACID DM00100[PS1977[17-471: Q19-4212	ALDEHYDE DEHYDROGENASES GLUTAMIC ACID DM00100JP48644[17-471: Q19-A212	Signal Peptide: M1-A26	Signal Peptide: M1-G28	PROTEIN OF SECTION OLYOXYLATE-INDUCED GBD BLAST_PRODOM	5 REGION ORF4 PHBRPOS INTERGENIC REGION PD150048: R5-G237	GLYOXYLATE; INDUCED; TRANSIENT; POTENTIAL; BLAST_DOMO DM03647[i39562]1-257; RS-G237	GLYOXYLATE; INDUCED; TRANSIENT; POTENTIAL; BLAST_DOMO DM03647 P30147 1-257: L4-G237	GLYOXYLATE; INDUCED; TRANSIENT; POTENTIAL; BLAST_DOMO DM03647IC641081-257; R5-G237	GLYOXYLATE; INDUCED; TRANSIENT; POTENTIAL; BLAST_DOMO DM03647P36951 1-260: L4-G237
Potential Glycosylation Sites							98IN 6N							
ylation	S34 S83 T45 T129 T155						S30 S214 Y161							
Amino Acid Potential Residues Phosphor Sites	247						237							
SEQ Incyte  ID Polypeptide  NO: ID	7511552CD1 247						7511722CD1  237							
SEQ NO NO	=						12							

Analytical Methods and Databases	HMMER_PFAM	BLIMPS_BLOCKS	BLAST_PRODOM	BLAST_PRODOM	BLAST_PRODOM	BLAST_DOMO	BLAST_DOMO	BLAST_DOMO	BLAST_DOMO	MOTIFS
Signature Sequences, Domains and Motifis	Squalene/phytoene synthase: L47-V275	Squalene and phytoene synthases IPB002060: S51-T81, Y171-G182, M207-P232	FARNESYLDIPHOSPHATE FARNESYLTRANSFERASE SQS SS PPP: FPP MULTIFUNCTION ENZYME	I MANAGHAS FUAZZAZA PEDA SYNTHASE TRANSTERASE BIOSYNTHESIS MULTIEUNCTIONAL ENZYME PHYTOENE SQUALENE CAROTEMODI SOPRENE PROTEIN PIDNOIROS, I AT-RCA, NAGAVJA	FARNESYLDIPHOSPHATIE FARNESYLTRANSFERASE BLAST_PRODOM PRANESYLDIRLOSS SYMPINES SOGS SS FPP. FPP MULTIPHOSTONAL ENZYME TRANSFERASE PDC2380-M1-Y46.	SQUALENE AND PHYTOENE SYNTHASES DM02785[P37268]1-403: M1-V235 P232-L345	SQUALENE AND PHYTOENE SYNTHASES DM02785[P53800]1-408: L7-V235 P232-L337	SQUALENE AND PHYTOENE SYNTHASES DM02785IP3659611-435: M28-V235 P232-K299	SQUALENE AND PHYTOENE SYNTHASES DM02785[P53799]1-409: L7-V235 P232-L342	Squalene and phytoene synthases signature 1: Y171-L186
Potential Glycosylation Sites	N48									
ylation	S39 S164 S188 S190 S297 T50 T81 T201 T306									
Amino Acid Potential Residues Phosphor Sites	358									
Incyte Polypeptide ID	7511489CD1 358									
SEQ NO NO										

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Analytical Methods and Databases	MOTIFS	BLAST_DOMO	BLAST_DOMO	BLIMPS_BLOCKS	BLAST_PRODOM	BLAST_DOMO		BLAST_DOMO	SPSCAN	PROFILESCAN	BLAST_PRODOM				BLAST_PRODOM			BLAST_DOMO	BLAST_DOMO	BLAST_DOMO
Signature Sequences, Domains and Motifs	Squalene and phytoene synthases signature 2: M207-P232	DM08732 METHYLTRANSFERASE P25627 10-189: P14-  BLAST_DOMO A89	DM08732 METHYLTRANSFERASE Q10162 10-189: P14- BLAST_DOMO V97	ubiE/COQ5 methyltransferase family IPB000339: L58-S69, BLIMPS_BLOCKS D82-R94	METHYLTRANSFERASE TRANSFERASE SIMILAR S CEREVISTAE CHNE YCR47C PD03 1003; L128-0170	DM08732 METHYLTRANSFERASE P25627 10-189: P14- BLAST_DOMO	S194	DM08732 METHYLTRANSFERASE Q10162 10-189: P14-  BLAST_DOMO	Signal cleavage: M1-A35	Thiolases signatures: A104-A153	3KETOACYLCOA THIOLASE PEROXISOMAL	PRECURSOR EC 2.3.1.16 BETA KETOTHIOLASE	ACELLICOA ACTE-IRANSFERASE SOAGACTE CON FATTY ACID METABOLISM TRANSFERASE	PEROXISOME TRANSIT PEPTIDE PD060746: M1-A36	TRANSFERASE SYNTHASE POLYKETIDE	ACTLIKANSFERASE PROTEIN I INOLASE BIOSYNTHESIS ACETYLCOA ACID PUTATIVE	PD000145: D37-A134	THIOLASES DM00548[P07871]34-421: S34-G135	THIOLASES DM00548 S57792 48-430: D37-A134	THIOLASES DM00548 S33637 45-434: D37-A134
Potential Glycosylation Sites																				
Potential Phosphorylation Sites		S102 S121 T132 Y48		S194 Y48					034 T50 TKO	001 (01 100										
Amino Acid Potential Residues Phosphor Sites		140		248					152	CCY										
Incyte Polypeptide TD		7511497CD1 140		7511498CD1 248					76116190TD1	1211014011										
SEQ Inc	į.	14		15					1	DŢ										

Analytical Methods and Databases	BLAST_DOMO	SPSCAN		HMMER	HMMER	HMMER	HMMER_PFAM	BLIMPS_BLOCKS	NA COST TECCO	TWO TENSOR	BLIMPS PRINTS			BLAST_PRODOM					BLAST_DOMO			BLAST_DOMO			BLAST_DOMO		
Signature Sequences, Domains and Motifs	THIOLASES DM00548 Q05493 26-413: D37-A134	Signal_cleavage: M1-A24		Signal Peptide: M1-A22	Signal Peptide: M1-A29	Signal Peptide: M1-A24	Aminotransferase class I and II: E111-T375	Aminotransferases class-I IPB001511: G63-G72, L154-	NIOU, OZGA-OZGO	Aminotransferases class-1 pyridoxai-pitospirate attactiment site: K202-0253	Aspartate aminotransferase signature PR00799; L153-W172, BLIMPS_PRINTS	L184-G198, C220-V240, S252-L277, I320-E338, I346-	V364	AMINOTRANSFERASE PYRIDOXAL PHOSPHATE	SYNTHASE TRANSFERASE ASPARTATE	TRANSAMINASE	1AMINOCYCLOPROPANEICARBOXYLATE A ACC	PD001174: S30-L145	AMINOTRANSFERASES CLASS-I PYRIDOXAL-	PHOSPHATE ATTACHMENT SI DM00997 P12344 32-	429: W32-K146, L121-K376	AMINOTRANSFERASES CLASS-I PYRIDOXAL-	PHOSPHATE ATTACHMENT SI DM00997 P46643 32-	429: W32-K146, P148-K376	AMINOTRANSFERASES CLASS-I PYRIDOXAL-	PHOSPHATE ATTACHMENT SI DM00997 S56678 29-	427: W32-F144, P148-K376
Potential Glycosylation Sites																										-	
Potential Phosphorylation	ones	S6 S26 S57 S79	S123 T34 T55	1107 007 0011																							
Amino Acid Potential Residues Phosphor		376										_															
Incyte Polypeptide	m.	7511624CD1 376																									
SEQ.	Ö	17			T	Ī					I			I													

Analytical Methods BLIMPS\_BLOCKS Aspartate aminotransferase signature PR00799: L164-W183, BLIMPS\_PRINTS HMMER PFAM BLAST DOMO PROFILESCAN BLAST\_DOMO BLAST\_DOMO BLAST\_DOMO and Databases HMMER MOTIFS HMMER HMMER SPSCAN PHOSPHATE ATTACHMENT SI DM00997|P46645|4-404; Aminotransferases class-I pyridoxal-phosphate attachment Aminotransferases class-I pyridoxal-phosphate attachment PHOSPHATE ATTACHMENT SI DM00997/P12344/32-PHOSPHATE ATTACHMENT SI DM00997|P46643|32-PHOSPHATE ATTACHMENT SI DM00997|S56678|29-Aminotransferases class-I IPB001511: G63-G72, L165-.195-G209, C231-V251, S263-L288, I331-E349, I357-AMINOTRANSFERASES CLASS-I PYRIDOXAL-AMINOTRANSFERASES CLASS-I PYRIDOXAL AMINOTRANSFERASES CLASS-I PYRIDOXAL-AMINOTRANSFERASES CLASS-I PYRIDOXAL Signature Sequences, Domains and Motifs Aminotransferase class I and II: V76-T386 429: P43-K387, W32-K82 429: W32-K82, R81-K387 427: W32-K82, R81-K387 Signal\_cleavage: M1-A24 Signal Peptide: M1-A22 Signal Peptide: M1-A29 Signal Peptide: M1-A24 W33-L155, P148-K376 R179, S233-G246 site: K213-0264 site: S222-G235 V375 Glycosylation Sites Potential Phosphorylation S6 S26 S57 S79 T34 T55 T174 Amino Acid Potential Y358 Sites Residues 7511626CD1 Polypeptide Incyte Θ NO. ₽

Table 3

Analytical Methods and Databases	BLAST_DOMO	MOTIFS	SPSCAN	HIMMER	HIMMER	HMMER	HMMER	HIMMER	HIMMER PFAM	TIMEMMER			BLIMPS_PRINTS	BLIMPS_PRINTS	BLAST_DOMO		BLAST DOMO		DI A CT. DOMOO	DLAST DOMO
Signature Sequences, Domains and Motifs	AMINOTRANSFERASES CLASS-I PYRIDOXAL- PHOSPHATE ATTACHMENT SI DM00997[P46248[52- 452: V36-K82, G72-1382	Aminotransferases class-I pyridoxal-phosphate attachment site: S233-G246	Signal_cleavage: M1-G65	Signal Peptide: M16-G33	Signal Peptide: M16-G36	Signal Peptide: M16-F38	Signal Peptide: M16-R39	Signal Peptide: M16-G46	short chain dehydrogenase: R51-R264	Cytosolic domain: M1-T20	Transmembrane domain: S21-W43	Non-cytosolic domain: V44-S274	Alcohol dehydrogenase superfamily signature PR00080: G134-I145, G187-I195	Glucose/ribitol dehydrogenase family signature PR00081: V54-V71, G134-I145, M181-G197	SHORT-CHAIN ALCOHOL DEHYDROGENASE	FAMILY DM00034[P54554[1-237: 157-A213, P218-1742	SHORT-CHAIN ALCOHOL DEHYDROGENASE	FAMILY DM00034 P00335 9-246: L50-1201,	SUDDIT CHAIN AT COUCH DEHANDOCENASE	FAMILY DM00034[Q10783]48-289: L50-A213
Potential Glycosylation Sites																				
Potential Phosphorylation Sites			S146 S217 T6 T106 T150 T154 Y167																	
Amino Acid Potential Residues Phosphor Sites		,	274																	
Incyte Polypeptide ID			7512885CD1																	
SEQ ID NO:			19																	

	Analytical Methods and Databases	BLAST_DOMO	MOTIFS	SPSCAN	HMMER PFAM	BLIMPS BLOCKS	BLIMPS_BLOCKS		PROFILESCAN	BLIMPS_PRINTS	BLAST_PRODOM	!		BLAST_DOMO		BLAST_DOMO	BLAST DOMO		BLAST DOMO		MOTTES
	Signature Sequences, Domains and Motifs	SHORT-CHAIN ALCOHOL DEHYDROGENASE FAMILY DM00034[P14802]1-235; L50-5244		Signal_cleavage: M1-G26	Aldo/keto reductase family: L7-W273	Ribosomal protein L10E IPB001197: I182-N234	Aldo/keto reductase family IPB001395: L7-L18, A35-Q60, BLIMPS_BLOCKS	V68-W80, T96-L109, K154-S188, 1214-M263	Aldo/keto reductase family signatures: 1211-A275	Aldo-keto reductase signature PR00069: A35-I59, K95-P113, V158-Y187, L205-F229	PROTEIN OXIDOREDUCTASE REDUCTASE	CHANNEL NADP DEHYDROGENASE ALDEHYDE	IONIC ALDOSE PUTATIVE PD000288: V28-L153, E143- N272, L7-D72, E249-M263	ALDO/KETO REDUCTASE FAMILY DM00192P07943 1-BLAST_DOMO	29/: AZ-INIOU, DI43-KZ/4	ALDO/KETO REDUCTASE FAMILY DM00192[1494842- BLAST_DOMO 298: A2-N160, E143-R274	ALDO/KETO REDUCTASE FAMILY DM00192[P31210]7- BLAST DOMO	308: V5-N160, E143-R274	ALDO/KETO REDUCTASE FAMILY DM00192 P51857 7- BLAST_DOMO	308: V5-N160, E143-R274	Aldo/keto reductase family signature 1: G39-G56
	Potential Glycosylation Sites			N234																	
	Potential Phosphorylation Sites			S8 S103 S133 S204 N234 S259 S282 T3 T20 T82 T96 T118 T137 T222 Y40 Y47																	
	Amino Acid Potential Residues Phosphor			293																	
,	D Polypeptide NO: ID			7511965CD1 293																	
CLO	g e ë			8																Ī	

Analytical Methods BLIMPS BLOCKS BLIMPS\_BLOCKS BLAST\_PRODOM BLAST\_PRODOM BLAST\_PRODOM HIMMER PFAM BLAST\_DOMO BLAST DOMO BLAST DOMO and Databases **TMHIMMER** DEHYDROGENASE OXIDOREDUCTASE ALDEHYDE NAD PROTEIN CLASS SEMIAL DEHYDE PRECURSOR ENDOPLASMIC RETICULUM MICROSOME PD020122: ALDEHYDE DEHYDROGENASES GLUTAMIC ACID ALDEHYDE DEHYDROGENASES GLUTAMIC ACID Aldehyde dehydrogenase family IPB002086: P101-P114, ALDEHYDE DEHYDROGENASES GLUTAMIC ACID Gamma-glutamyl phosphate reductase IPB000965: G15-TRANSIT PEPTIDE PD000218: V5-Q153, G161-K319 MICROSOMAL CLASS OXIDOREDUCTASE NAD ALDEHYDE DEHYDROGENASE LIKE PROTEIN DEHYDROGENASE FATTY ALDEHYDE Aldehyde dehydrogenase family: M1-S320 Signature Sequences, Domains and Motifs Fransmembrane domain: L355-A373 DM00100[P51648]1-405: M1-Y160, DM00100|P30838|1-408: V5-Y160, DM00100P43353|1-408: V5-Y160, Non-cytosolic domain: M1-K354 Ovtosolic domain: E374-Y376 E67, L106-T143, E222-L250 T201-L250, V279-G303 PD065537: L139-I365 321-K372 Q157-G297 0157-G297 K149-G297 Glycosylation Sites Potential Phosphorylation F143 T210 T275 S14 S184 S320 S337 T80 T90 Amino Acid Potential Sites Residues 7512403CD1 Polypeptide ncyte ë

Table 3

Analytical Methods and Databases	BLAST_DOMO	SPSCAN	HMMER	HMMER	HMMER	HMMER	HMMER PFAM	BLIMPS_PRINTS	BLAST_DOMO	BLAST_DOMO	BLAST_DOMO	BLAST_DOMO	SPSCAN	HMMER PFAM	BLIMPS_PRODOM
Signature Sequences, Domains and Motifs	ALDEHYDB DEHYDROGENASES GLUTAMIC ACID DM00100[P12693]1-432: L3-L151, L139-G297	Signal_cleavage: M1-A20	Signal Peptide: M5-P22	Signal Peptide: M5-K26	Signal Peptide: MI-R25	Signal Peptide: M1-K26	short chain dehydrogenase: P40-R233	Alcohol dehydrogenase superfamily signature PR00080: K119-V130	SHORT-CHAIN ALCOHOL DEHYDROGENASE FAMILY DM00034 S42651 28-318: V45-T157, G152-L243	SHORT-CHAIN ALCOHOL DEHYDROGENASE FAMILY DM00034 S30167  82-368: V44-G152, H150-1187	SHORT-CHAIN ALCOHOL DEHYDROGENASE FAMILY DM00034[S39394[69-356: V43-G152, H150-1187	SHORT-CHAIN ALCOHOL DEHYDROGENASE FAMILY DM00034P13653 69-356: V43-G152, H150-1187	Signal_cleavage: M1-A51	AhpC/TSA family: V65-R198	PROTEIN ANTIOXIDANT PEROXIDASE REDUCTASE BLIMPS_PRODOM PD00210: V96-E111
Potential Glycosylation Sites		N228													
Potential Phosphorylation Sites		S106 T57											S86 S116 T146 T195		
Amino Acid Potential Residues Phosphor Sites		248											198		
Incyte Polypeptide ID		7512564CD1											7512646CD1 198		
S D S		22											23 7		

Analytical Methods and Databases	BLAST_PRODOM	BLAST_PRODOM	BLAST_DOMO	BLAST_DOMO	BLAST_DOMO	BLAST_DOMO	MOTIFS	SPSCAN	HMMER PFAM		PROFILESCAN	BLIMPS_PRINTS
Signature Sequences, Domains and Motifs	PROTEIN ANTIOXIDANT PEROXIDASE REDUCTASE BLAST_PRODOM PROBABLE OXIDOREDICTASE THIOREDOXIN PEROXIDE THOREDOXIN-DEPENDENT ALKYI. PROMOSS: A64-A182	MITOCHONDRIAL THIOREDOXIN-DEPENDENT PEROXIDE REDUCTASE PRECURSOR ANTIOXIDANT PROTEIN AOPI MITOCHONDRION TRANSIT PRO19666. PM-A64	ALKYL HYDROPEROXIDASE C22 PROTEIN DM00437[P20108]62-250: H61-R184	ALKYL HYDROPEROXIDASE C22 PROTEIN DM00437JIC2258J1-188: A62-R184	ALKYL HYDROPEROXIDASE C22 PROTEIN DM00437 A57716 4-192: A69-R184	ALKYL HYDROPEROXIDASE C22 PROTEIN DM00437[P35703]4-193: A69-R184		Signal_cleavage: M1-G24	Aldo/keto reductase family: L12-V250	Aldo/keto reductase family IPB001395: L12-L23, T49-162, T94-P127, K130-S164, L191-M240	Aldoketo reductase family signatures: D73-0137, L188-R251	Aldo-keto reductase signature PR00069: R48-P66, M98-F115, V134-Y163, L182-P206
Potential Glycosylation Sites								N176 N196				,
Potential Phosphorylation Sites								S7 S234 T25 T49 T168 T236 Y76				
Amino Acid Potential Residues Phosphor Sites												
SEQ Incyte  ID Polypeptide  NO: ID								7512700CD1 270				
ğ a ë							- 11	74				

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Analytical Methods and Databases	BLAST_PRODOM	BLAST_DOMO	BLAST_DOMO	BLAST_DOMO	BLAST_DOMO	MOTIFS	MOTIFS	SPSCAN	BLIMPS BLOCKS	BLIMPS_BLOCKS	BLIMPS_PRINTS	BLAST_PRODOM
Signature Sequences, Domains and Motifs	PROTEIN OXIDOREDUCTASE REDUCTASE CHANNEL NADP DEHYDROGENASE ALDEHYDE IONIC ALDOSE PUTATIVE PD00288: L12-Y26, T25- 1255	ALDOKETO REDUCTASE FAMILY DM00192 P51857 7-BLAST_DOMO 308: S7-831, L32-V253	ALDO/KETO REDUCTASE FAMILY DM00192[P31210]7- BLAST_DOMO 308: S7-K30, L32-V253	ALDO/KETO REDUCTASE FAMILY DM00192/JH0575[5] BLAST_DOMO 305: S7-M41, L32-F252	ALDO/KETO REDUCTASE FAMILY DM00192 P51652 5-  BLAST_DOMO 305: R9-L32, L32-F252	Aldo/keto reductase family signature 2: M98-F115	Aldo/keto reductase family putative active site signature: 1215-1230	Signal_cleavage: M1-P16	GMC oxidoreductase IPB000172: D37-G55	UbiH/COQ6 monooxygenase family IPB000689: D37-L54, BLIMPS_BLOCKS K202-L236	Aromatic-ring hydroxylase (flavoprotein monooxygenase) signature PR00420: D37-H59, Q200-R215	PROTEIN OXIDOREDUCTASE MONOOXY GENASE FLA VOPROTEIN FAD UBIQUINONE BIOSYNTHESIS PUTATIVE MONOOXGENASE COQ6 PD00974: D37- B157
Potential Glycosylation Sites								N222				
Potential Phosphorylation	Salics							S23 S28 S84 S160   N222 S283 S295 T34 T192 T273 Y141				
Amino Acid Potential Residues Phospho								306				
Incyte Polypeptide	1							7512707CD1 306				
SEQ EQ	. Ok							52				

Analytical Methods and Databases	BLAST_PRODOM	SPSCAN	HIMMER_PFAM	TIMEMIMER		BLIMPS_BLOCKS	BLIMPS_BLOCKS	BLIMPS_BLOCKS	BLIMPS_PRINTS	BLIMPS_PRINTS	BLIMPS_PRINTS
Signature Sequences, Domains and Motifs	UBIQUINONE BIOSYNTHESIS MONOOXGENASE COQE OXIDOREDICTASE MONOOXYGENASE ILAVOPROTEIN FAD PUTATIVE PROTEIN PD 155418: 1005-1281	Signal_cleavage: M1-P16	Monooxygenase: Q155-D383	Cytosolic domain: M1-D37	Transmembrane domain: V38-D60 Non-cytosolic domain: H61-K423	UbiH/COQ6 monooxygenase family IPB000689: D37-L54, BLIMPS_BLOCKS IX157-L191, F301-L344	Glucose-methanol-choline oxidoreductase IPB000172: D37- BLIMPS_BLOCKS G55	Furnarate roductase/succinate dehydrogenase, FAD-binding BLIMPS_BLOCKS site IPB000464: D37-A58	FAD-dependent glycerol-3-phosphate dehydrogenase family BLIMPS_PRINTS signature PR01001: Y36-A48	Aromatic-ring hydroxylase (flavoprotein monooxygenase) signature PR00420: D37-W59, Q155-R170, R313-P328, P328-L344, Y367-D383	Flavin-containing amine oxidase signature PR00757: D37- F56
Potential Glycosylation Sites		77IN									
Potential Phosphorylation Sites		S23 S28 S115 S238 N177 S254 S282 T34 T147 T228 T365 T369 Y96									
Amino Acid Potential Residues Phosphor Sites											
Incyte Polypeptide ID		7512710CD1   423									
SEQ D D		56									

spi		M					M				FAM	KS	KS	
Analytical Methods	abases	BLAST_PRODOM	BLAST_DOMO	BLAST_DOMO	BLAST_DOMO	S	PRODC	S	<u>~</u>	HMMER_PFAM	HMMER_TIGREAM	BLIMPS_BLOCKS	BLIMPS_BLOCKS	
Analytic	and Databases	BLAST	BLAST.	BLAST	BLAST	MOTIF	BLAST	MOTIFS	HIMMER	HIMME	HMME	BLIMP	BLIMP	
Signature Sequences, Domains and Motifs		UBIQUINONE BIOSYNTHESIS MONOOXGENASE COQ6 OXIDOREDICTASE MONOOXYGENASE FLAVOPROTEIN PAD PUTATIVE PROTEIN PD155418: 1160-1236	FAD NP_BIND DM00811 P25535 207-346: G210-P252, V290-L379	FAD NP_BIND DM00811[P53318[282-431: P287-L379	FAD NP_BIND DM00811 P25534 205-343: A295-L379	ubiH/COQ6 monooxygenase family signature: H327-D340 MOTIFS	DIOXX GENASE CYSTEINE CDO OXIDOREDUCTASE   BLAST_PRODOM IRON CDOI SIMILAR HUMAN PD016803: L15-G82	Riboflavin synthase alpha chain family signature: L19-E31	Signal Peptide: MI-A24	Flavin containing amine oxidoreductase: 112-L452	proto_IX_ox: PROTO-PORPHYRINOGEN oxidase: M1- S458	D-amino acid oxidase, DAO IPB000927: T4-A16	Glucose-methanol-choline oxidoreductase IPB000172: T4-	
Potential	Glycosylation Sites													
Potential	Phosphorylation Sites						T111		S36 S45 S160 S209 S252 S256 S318 S413 S443 T174 T226					
몆	Residues						88							
SEQ Incyte	Polypeptide ID						7512884CD1 88		7512931CD1   458					
SEQ	βŘ						27		28					

BLAST PRODOM BLAST PRODOM BLAST PRODOM BLIMPS BLOCKS Analytical Methods BLIMPS PRINTS BLAST\_DOMO BLAST\_DOMO BLAST\_DOMO BLAST\_DOMO and Databases MOTIFS HMMER OXIDOREDUCTASE PPO PORPHYRIN BIOSYNTHESIS OXIDOREDUCTASE PPO PORPHYRIN BIOS YNTHESIS Glucose-methanol-choline oxidoreductase IPB000172: T4-Flavin-containing amine oxidase signature PR00757: T4-FLAVOPROTEIN FAD HEME MITOCHONDRION FLAVOPROTEIN FAD HEME MITOCHONDRION PD004706: V101-R188, W208-S353, I53-K117 OXIDOREDUCTASE FLAVOPROTEIN FAD DM03709|Q10062|1-353: T4-L112, A104-S318 DM03709|P50336|3-340: R3-P118, G111-D321 PROTO-PORPHYRINOGEN OXIDASE PPO MITOCHONDRION PD041529: W354-H396 PROTO-PORPHYRINOGEN; OXIDASE; PROTO-PORPHYRINOGEN: OXIDASE: PROTO-PORPHYRINOGEN; OXIDASE; PROTO-PORPHYRINOGEN; OXIDASE; Signature Sequences, Domains and Motifs OXIDASE PROTO-PORPHYRINOGEN OXIDASE PROTO-PORPHYRINOGEN PORPHYRIN BIOSYNTHESIS HEME DM04318|P50336|365-476: V346-S458 DM03709|P32397|6-344: R47-D321 Cell attachment sequence: R88-D90 Signal\_cleavage: M1-A17 Signal Peptide: M1-A24 PD006423: G383-G453 S22 R23 Glycosylation Sites Potential S36 S45 S140 S170 Phosphorylation 5 cm Potential Amino Acid Residues 7512933CD1 Polypeptide Incyte

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Analytical Methods	and Databases	BLIMPS_PRINTS	BLAST PRODOM				BLAST_PRODOM				BLAST DOMO		BLAST_DOMO		SPSCAN	HMMER	BLIMPS_BLOCKS	BLIMPS_BLOCKS	BLIMPS PRINTS		BLIMPS PRINTS		BLAST_PRODOM			
Signature Sequences, Domains and Motifs		Flavin-containing monooxygenase (FMO) signature PR00370: R3-Y19	OXIDASE PROTO-PORPHYRINOGEN	OXIDOREDUCTASE PPO PORPHYRIN BIOSYNTHESIS	FLAVOPROTEIN FAD HEME MITOCHONDRION	PD006423: G110-G180	PROTO-PORPHYRINOGEN OXIDASE PPO	PORPHYRIN BIOSYNTHESIS HEME	OXIDOREDUCTASE FLAVOPROTEIN FAD	MITOCHONDRION PD041529: W81-H123	PROTO-PORPHYRINOGEN; OXIDASE;	DM04318 P50336 365-476: V75-S185	PROTO-PORPHYRINGGEN; OXIDASE;	DM03709 P50336 3-340: R3-V75	Signal_cleavage: M1-A17	Signal Peptide: M1-A24	-	Glucose-methanol-choline oxidoreductase IPB000172: T4- S22	Flavin-containing amine oxidase signature PR00757: T4-	R23	Flavin-containing monooxygenase (FMO) signature	PR00370: R3-Y19		OXIDOREDUCTASE PPO PORPHYRIN BIOSYNTHESIS	FLA VOPROTEIN FAD HEME MITOCHONDRION	PD006423: G65-G135
Potential	Glycosylation Sites																									_
Potential	Phosphorylation Sites														S95 S125											
Amino Acid Potential	Residues																									
SEQ Incyte	Polypeptide ID														7512942CD1 140											
SEQ.	g ë			_										7	30											

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Analytical Methods and Databases	BLAST_PRODOM	BLAST_DOMO	BLAST_DOMO	SPSCAN	HIMMER PFAM	BLIMPS_BLOCKS	BLAST_PRODOM	BLAST_PRODOM	BLAST_DOMO	BLAST_DOMO
Signature Soquences, Domains and Motifs	PROTO-PORPHYRINOGEN OXIDASE PPO PORPHYRIN BIOSYNTHEISS HEMB OXIDOREDUCTASE FLAVOROGIEN FAD MITOCHONDRION PDG41529: W36-H78	PROTO-PORPHYRINOGEN; OXIDASE; DM04318[P50336]365-476: V30-S140	PROTO-PORPHYRINOGEN; OXIDASE; DM03709JP50336J3-340: R3-M69	Signal_cleavage: M1-S18	FMN-dependent dehydrogenase: A13-F278	FMN-dependent alpha-hydroxy acid dehydrogenases IPB000262: V204-E247	OXIDASE OXIDOREDUCTASE DEHYDROGENASE FLAYOPROTEIN PAN LLACTATE GLYCOLATE PROTEIN CYTOCHROME CHAIN PD003201: A13-S184	OXIDASE OXIDOREDUCTASE PROTEIN FLAVOPROTEIN FAM DELYDROGENASE FORTHERME LIACTATE CHAIN GLYCOLATE PD00673C: C208-N272	IMN-DEPENDENT AI PHA-HYDROXY ACID DEHYDROGENASES DM01725 P00175 201-561: K199- 1229, S2-R170, V198-5279	FMN-DEPENDENT ALPHA-HYDROXY ACID DEHYDROGENASES DM01725P054142-357;
Potential Glycosylation Sites				9/IN						
ylation				S20 S56 S112 S184 N176 S267 T34 T92 T178 T212						
Amino Acid Potential Residues Phosphor Sites										
Incyte Polypeptide ID				7513736CD1   281						
g a g				31						

Analytical Methods and Databases	BLAST_DOMO	BLAST_DOMO	MOTIFS	HMMER_PFAM	BLIMPS_BLOCKS	BLAST_PRODOM	BLAST_PRODOM	BLAST_PRODOM	BLAST_DOMO	BLAST_DOMO
Signature Sequences, Domains and Motifs	FMN-DEPENDENT ALPHA-HYDROXY ACID DEHYDROGENASES DM01725 P09437 186-540: E114- N138. L3-R170. 1188-V276	FMN-DEPENDENT ALPHA-HYDROXY ACID DEHYDROGENASES DM01725[Q07523]1-350: L3-K187, 1188-R280	Bacterial regulatory proteins, lysR family signature: R94- L124	Pyridoxal-dependent decarboxylase consorved domain: P49- HMMER_PFAM P417	Pyridoxal-dependent decarboxylase family IPB002129: G144-O166, P181-L203, WZ69-G278, G292-D320	GLUTAMATE DECARBOXYLASE PD114206. L10-N112 BLAST_PRODOM	DECARBOXYI_ASE LYASE PYRIDOXAL PHOSPHATE BLAST_PRODOM MULTGENE FAMILY DOPA GLUTAMATIA ACID AROMATICL-AMINOACID PD001960: P49-E406	DECARBOXYI.ASE CYSTEINE SULFINOALANINE CYSTEINESULFINATB LYASE PYRIDOXAL PHOSPHATE SULFINIC ACID SULFINATE PD153956: M.1-248.	DDC / GAD / HDC / TYRDC PYRIDOXAL-PHOSPHATE   BLAST_DOMO ATTACHMENT SITE DM00568 H0627 84-575: L10-E406	DDC / GAD / HDC / TYRDC PYRIDOXAL-PHOSPHATE   BLAST_DOMO ATTACHMENT SITE DM00568 P14748  101-591: E5-E406
Potential Glycosylation Sites				N112						
Potential Phosphorylation Sites				S40 S62 S66 S81 S95 S142 S207 T186 T285						
Amino Acid Potential Residues Phosphor Sites				463						
Incyte Polypeptide				7512779CD1 463						
SEQ EQ				32						

Analytical Methods and Databases	BLAST_DOMO	BLAST_DOMO	MOTIFS	HMMER_PFAM	BLIMPS_BLOCKS	BLAST_PRODOM	0000	BLAST_DOMO	BLAST_DOMO	SPSCAN	HMMER	HMMER_PFAM	HMMER_PFAM	HMMER_TIGRFAM
Signature Sequences, Domains and Motifs	DDC / GAD / HDC / TYRDC PYRIDOXAL. PHOSPHATE   BLAST_DOMO ATTACHMENT SITE DM00568 Q05329 95-582: S4-E406	DDC / GAD / HDC / TYRDC PYRIDOXAL-PHOSPHATE BLAST_DOMO ATTACHMENT SITE DM00568 S55689 5-478: L7-P407	Eukaryotic putative RNA-binding region RNP-1 signature: K196-L203	3-hydroxyacyl-CoA dehydrogenase, NAD binding: E43- Q149	3-hydroxyacyl-CoA dehydrogenase IPB002135: D118-Q150 BLIMPS_BLOCKS	DEHYDROGENASE PROTEIN 3HYDROXY-ACYLCOA BLAST_PRODOM FATTY ACID METABOLISM OXIDOREDUCTASE 3HYDROXY-BUTYRYLCOA INCLXIDES: ENOYLCOA	PD001126: V46-K140	3-HYDROXYACYL-COA DEHYDROGENASE DM00842[P00348]11-306: V46-E144	3-HYDROXYACYL-COA DEHYDROGENASE DM00842[P41938]18-308: V46-D139		Signal Peptide: M1-G29	Quinolinate phosphoribosyl transferase, C-terminal domain: HMMER_PFAM A62-K190	Quinolinate phosphoribosyl transferase, N-terminal domain: HMMER_PFAM C27-S110	Nicotinate-nucleotide pyrophosphoryl a (nadC): L18-L191 HMMER_TIGRFAM
Potential Glycosylation Sites														
rylation				S70 S77 S145 T10 T106						S110 S188				
Amino Acid Potential Residues Phospho Sites				149						1203				
SEQ Incyte ID Polypeptide NO: ID				7512877CD1 149						7512782CD1				
S E SE				33						34				

Analytical Methods	and Databases	BLAST_PRODOM		BLAST_DOMO	SPSCAN	HMMER	HMMBR_PFAM	HMMER_TIGREAM	BLAST_PRODOM			BLAST_DOMO	BLIMPS_BLOCKS	BLIMPS_PRODOM
Signature Sequences, Domains and Motifs			GLYCOSYL-TRANSFERASE PROBABLE PD003988: W22-R95, V87-P147	PYROPHOSPHORYLASE; NICOTINATE; NUCLEOTIDE; TRANSPORT; DM02806[P43619]1-274; V10-D83 V87-X177	Signal cleavage: M1-A44		syl transferase, C-te: T14-K153	Nicotinate-nucleotide pyrophosphoryl a (nadC); L18-L154 HMMER_TTGRFAM	TRANSFERASE NICOTINATENUCLEOTIDE PYROPHOSPHORYLASE QUINOLINATE CARDONYM ATHIC PHOSPHORIBORY	TRANSFERASE DECARBOXYLATING QAPRTASE GLYCOSYLTRANSFERASE PROBABLE PD003988:	A53-P110	PYROPHOSPHORYLASB; NICOTINATE; NUCLEOTIDB; TRANSPORT; DMD2806fb436191-274; V19-G52, L46-V135	5: F7-F18	DECARBOXYLASE BIOSYNTHESIS S-ADBNOSYLMETHIONINE PD02625: F7-F18
	Glycosylation Sites													
Potential	Phosphorylation Sites				873.8151								T10	
Amino Acid Potential	Residues				166								84	
	ypeptide				7512784CD1								7512794CD1 48	
SEQ Incyte	回 日 日 日 日				35								36	

Analytical Methods and Databases	BLAST_DOMO	SPSCAN	HMMER	HMMER	HMMER	HWMER	HMMER	TIMEMINER			BLIMPS_BLOCKS	BLAST_PRODOM		SPSCAN	HMMER	HMMER	HMMER	HMMER	HMMER	TMHMMER			PROFILESCAN		BLIMPS_PRINTS	
Signature Sequences, Domains and Motifs	S-ADENOSYLMETHIONINE DECARBOXYLASE BETA BLAST_DOMO CHAIN DM01913 48237 1-333: M1-L42	Signal_cleavage: MI-G17	Signal Peptide: M1-G17	Signal Peptide: M1-L19	Signal Peptide: M1-W20	Signal Peptide: M1-D23	Signal Peptide: M4-D23	Cytosolic domain: M1-M4	Transmembrane domain: W5-W22	Non-cytosolic domain: D23-P244	Sulfatase IPB000917: M55-R66	PIGL PROTEIN CATSATP13 INTERGENIC REGION	URF2 PD039078: L45-R199	Signal_cleavage: M1-G35	Signal Peptide: M18-S32	Signal Peptide: M18-G35	Signal Peptide: R16-G35	Signal Peptide: N14-G35	Signal Peptide: M1-G35	Cytosolic domain: M1-K19	Transmembrane domain: L20-V38	Non-cytosolic domain: N39-H115	Tau and MAP proteins tubulin-binding domain signature:	V10-166	nine ADP-ribosyltransferase signature PR00970: M18-	832
Potential Glycosylation Sites																										
Potential Phosphorylation Sites		S25 S172 T87 T207												S42 S53 S84 T74												
Amino Acid Potential Residues Phospho Sites		244												115												
Incyte Polypeptide ID		7512886CD1												7512929CD1												
SEQ NO.		37												38												

Polynucleotide	Sequence Fragments
SEQ ID NO:/	
Incyte ID/ Sequence	
39/7511289CB1	1-105, 1-126, 1-210, 1-471, 1-644, 1-1792, 15-219, 18-219, 18-297, 24-579, 54-320, 105-544, 130-714, 152-353, 168-438, 174-433, 175-
1798	436, 179-559, 182-615, 196-708, 199-461, 202-697, 208-771, 225-721, 227-711, 227-713, 227-714, 228-523, 236-714, 244-444, 244-451,
	247-770, 257-729, 266-714, 279-577, 305-614, 305-713, 315-621, 346-807, 346-865, 403-694, 434-530, 453-715, 476-715, 490-694, 518-
	715. 533-715. 713-963. 722-1154, 723-1154, 735-1154, 767-1038, 767-1155, 770-1223, 771-1297, 791-1068, 803-1056, 812-1241, 822-
	1088, 822-1292, 842-1154, 845-1489, 860-1311, 879-1122, 881-1470, 886-1148, 912-1187, 949-1280, 952-1096, 989-1231, 1010-1620,
	1029-1225, 1031-1778, 1036-1624, 1042-1274, 1079-1349, 1080-1638, 1097-1414, 1126-1400, 1145-1798, 1162-1421, 1163-1389, 1167-
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75	7512886CB1	SINTBST01
76	7512929CB1	LUNGNOT12

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ADRETUT01	E	Infarty recent design RNA isolated from right adrenal tumor tissue removed from a 50-year-old Turkish male during amainteen admenteered with exteriors that demote a tiemmerical spough. Amainteen admenteer great electricom that demote a tiemmerical spough. Benoching in the region of the medular. The patient presented with corticoadrenal insufficiency, inclicomal hermin, and non-alcoholic steato hepatitis. Patient history included renal cell carcinoma. Family history included renal cell carcinoma. Family history included renal cell carcinoma. Family history included itwee cancer.
BRAINON01	PSPORT1	Library was constructed and normalized from 4.88 million independent closes from a britin issue library, KNA was made from vania sissue unovered from a 26-year-cold cusasian made during cannelogary and excision of a cerebral memigeal liston. Pathology for the associated tumor issue inclinated a grade 4 oligosateo/roman in the right tomo-parietal part of the brain. The normalization and hydridization conditions were adapted from Source et al., PNAS (1964) 91;9228, except that a significantly longer (48-bour) rearmenting hydridization was used.
BRAINOT03	PSPORTI	Library was constructed using RNA isolated from brain tissue removed from a 26-year-old Caucasian male during cramophasty man decision meningal siston. Pathology for the associated tumor tissue indicated a grade 4 oligoastrocyforna in the right fromto-partied part of the brain.
BRAZNOT01	pINCY	Library was constructed using RNA signated from straining, globus pullisa and posterior pulment issue encrowed from 4.5- year-old Caucasian female who died from a dissecting acrdis amongsman and isohamic bowel disease. Pathology indicated mild arteriosaciousis involving the execting cortic amongsman and isohal among the was mild menalinged filtronis and mild focal attention bitsue in middle exected rarcy, as well as vertical attention bitsuely, filtronis and mild focal attention bitsue, but mindle exected front areas in the white matter that contain blood vessels that were burned stapped, bytalizined, with themostedicari-laten mancrophages in the "richtoo-Nobin store. In addition, there were scattered neuroribrillary tangles whitm the basolateral under of the sunyguiah. Pattent tissury richtede mild atheomatosis of sort and coronary atteries, bowel and liver infarct due to aneurysm, physiologic fatty liver associated with obesity, mild diffuse emphysema, thrombosis of
BRSTNOT02	PSPORT1	inescanticis and potal veins, cardiomegaly due to hypercopyloy of lest venirole, arterial hypertension, acute pulmonary odenna, spetemengaly, obesity (300 lb.), leionyona of utens, sheps gamea, and iron deficiency anamia. In the state of the castan fermale during Library was constructed using RNA isolated from diseased treast tissue removed from a 55-year-old Caucasian fermale during a millicated, and canded simple masterousy. Pathology inficient de profiterates fibrogycale, changes characterized by upocition mengplasia, electrodist gamea experiency, and experiency and applicate the properties of the associated unnor tissue indicated an investive grade 4 mammary adenocarcinoma. Patient history included artial tachycardia and a benign propisam. Family history included cardiovascular and cerebrovascular disease.

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#### Caucasian male during a sigmoidectomy and permanent colostomy. Pathology indicated invasive grade 2 adenocarcinoma. One termatitis. Family history included benign hypertension, atherosclerotic coronary artery disease, hyperlipidemia, breast cancer, epithelial cell tissue library. Starting RNA was made from untreated prostatic epithelial cell issue removed from a 17-year-old Hispanic male. The library was normalized in two rounds using conditions adapted from Soares et al., PNAS (1994) 91:9228 prostate fibroblast library. Starting RNA was made from fibroblasts of prostate stroma removed from a male fetus, who died Estrogen and progesterone immunostains were positive in the neoplastic cells. Patient history included cirrhosis of the liver, containing clones as follows: plasmid DNA was prepped from approximately 1 million clones from the normalized prostate ymph node contained metastasis with extranodal extension. Patient history included hyperlipidemia, cataract disorder, and This normalized prostate epithelial cell tissue library was constructed from 2.36 million independent clones from a prostate esophageal ulcer, hyperlipidemia, and neuropathy caused by an unspecified disease. Previous surgeries included segmental after 26 weeks' gestation. The libraries were normalized in two rounds using conditions adapted from Soares et al., PNAS Library was constructed using RNA isolated from right breast tumor tissue removed from a 59-year-old Caucasian female napillomatosis. Prior right breast biopsy indicated invasive grade 3, nuclear grade 3, invasive and in situ ductal carcinoma. lung resection and a liver transplant. Patient medications included Prograf, prednisone, Imuran, Lozol, Zantac, Estraderm The normalized prostate stromal fibroblast tissue libraries were constructed from 1.56 million independent clones from a forming an ill-defined mass situated in the right breast biopsy cavity site. The non-neoplastic breast parenchyma displays during unilateral extended simple mastectomy. Pathology indicated invasive lobular carcinoma with extension into ducts Library was constructed using RNA isolated from colon tumor tissue obtained from the sigmoid colon of a 62-year-old hours/round)reannealing hybridization was used. The library was then linearized and recircularized to select for insert and Bonaldo et al., Genome Research (1996) 6:791, except that a significantly longer (48-hours/round) reannealing 1994) 91:9228 and Bonaldo et al., Genome Research (1996) 6:791, except that a significantly longer (48stromal fibroblast tissue libraries following soft agar transformation. nybridization was used. atches, and Provera. Library Description and prostate cancer. pINCY Vector PINCY SINCY PINCY COLNTUTOS BRSTTUT22 3PIPNON05 FIBPFEN06

Library	Vector	Library Description
FTUBTUT02	pINCY	Liberay was constructed using RNA isolated from fallopian tube tumor issue removed from an 85-year-old cuestain fermale during bilateral sulpingo-coplorectomy and hysterectomy. Pathology indicated poorly differentiated mixed endometrioid (80%) and serous (20%) adenorection and which was confined to the microsa without mural involvement. Endometrioid experience in serious in situ was also present endometriology for the associated tuents unnor indicated food endometrioid adenocarcinoma in situ an noderately differentiated in visely acknowner/more in an endometrial polyp. Metastatic endometrioid and serous adenocarcinoma was present a the cul-de-sac tumor. Patient history included medullary carcinoma of the thyroid and myocardial infrarction.
GBLADIT02	pINCY	The library was constructed using RNA isolated from diseased guilhidder tissue removed from a 16-year-old Camesisian femic holms during release to the control of the contr
HNT2RAT01	PBLUESCRIPT	PBI_UESCRIPT interry was constructed at Stratagene (STR937231), using RNA isolated from the hNT2 cell line (derived from a human leture. An extra constituted properties characteristic of a committed neuronal precursor). Cells were treated with retinoic acid for 24 hours.
ISLTNOT01 LIVRBCT01	pINCY PBLUESCRIPT	pINCY Lithrary was constructed using RNA isolated from a pooled collection of paracratic islet cells.  PBIJUES/GRIPT Library was constructed using RNA isolated from the liver tissue of a patient with primary biliary circlosis who had a liver transplant.  Introplant.
LIVRTUBOL	PCDNA2.1	This 5' biased random printed library was constructed using RNA isolated from liver tumor tissue emboved from a 72-year-old forestain and education partial betatement. Pathogo included methods and every continuous forestain and education partial betatement. Pathogo included betated for expension, uppe 1 diaches, prestation by perputation, type 1 diaches, prostation by perputation processing of with mensatatic liver cancer. Patient history included benign hypertension, type 1 diaches, prostation by perputation and processing of with mensatatic liver cancer. Patient history included descriptions of partial processing of programments and total appearation of a processing proce

Library	Vector	Library Description
4ON07	pINCY	This normalized lung tissue library was constructed from 5.1 million independent clones from a lung tissue library. Starting NNA was medic from RNA stoaked from lung tissue. The library was normalized in two rounds using conditions adapted from SNA 2000 set al., PNAS (1994) 91:9228-9223 and Bornaloc at al., Genome Research (1996) 67/91, except that a significantly longer (48 hours/nound) remaining hybridization was used.
LUNGNOTI2	pINCY	Library was constructed using RNA isolated from lung issue removed from a 78-year-old Caucasian male during a segmental lung resection and regional lymph tode essection. Plankogy indicated fibrosis plears was protected, but not invaded. Pathology for the associated unmer issue indicated an invasive plutnosing grade 3 denoncarionen. Patient bistory included corebronsecular disease, arterioscierotio coronary attery disease, thrombophilebitis, chronic obstructive pulmonary disease, and saffran. Femily bistory included intractualla hematoma, cerebrovascular disease, arterioscierotic coronary artery disease, and saffran. Femily bistory included intractualla hematoma, cerebrovascular disease, arterioscierotic coronary artery disease, and publishess.
MENITUT03	pINCY	Library was constructed using RNA isolated from brain meningtoma tissue removed from a 55-year-old Caucasian female during excision of a carebal meningsal lesion. Pathology indicated a bening mospiasm in the right corebelloporatine ungle of the brain. Patient history included hypothyroidism. Family history included myocardial infarction and breast cancer.
MYOMNOT01 PSPORT1	PSPORT1	Library was constructed using RNA isolated from uterine myometrial tissue removed from a 43-year-old Caucasian female during a vaginal hysteocriousy and removal of the failopian tubes and overless. Family history included lung cancer, stroke, type II diabetes, hepatic lesion, chronic liver disease, hyperlipidemia, congenital heart anomaly, and mitral valve prolupes.
OVARNOT03 PSPORTI	PSPORTI	Library was constructed using RN4 sloaked from overation sisses removed from a 45-yacoled funcation from bed unity grounval for the fallowing these and overable bed from the fall of the fallowing these and overable Public for the exociated tumor tissure indicated grade 2 manious cystalenocurcinoms. Patient listory included mintal valve disorder, preumonia, and viral hepatitis. Family history included attentosclerotic coronary arresty disease, present cancer, and uterine cancer, stress reaction, cerebrovascular disease, breast cancer, and uterine cancer.
OVARTUT04	pINCY	Library was constructed using RNA isolated from ovarian tumor tissue removed from 12-yea-old Cunstain framile during a real absormatel pysterectory, removed of the hillogine more statement of the right oversy. The left oversy that is branchagies compared in the maniform of the right oversy. The left oversy had is branchagies compared the maniform of the right oversy. The left oversy that is branchagies compared and multiple fellowances (I statemental), and the endometrium was interior. Fix end-de-sace contained abundant historys and rare clusters of mesofhelial colls. Patient history included breast fibrosclerosis and chronic stomach ulcer. Family history included acute stomach ulcer with perforation, breast cancer, heading more, hearing hypertension, coronny angioplasty, and hypertipidemia.

Library	Vector	Library Description
0NV10	PCR2-TOPOTA	PCR2.TOPOTA Library was constituted using pooled cDNA from different donors. cDNA was generated using mRNA isolated from pooled steeled make its extensor from text to 10.5 7-year-old Caustain make and female notions who died from sudden darti; from pooled thymus itssue removed from into 18 to 32-year-old Caustain make and female donors who died from sudden death; from pooled thymus itssue removed from into 18 to 32-year-old Caustain make and female choras who died from sudden death; from pooled liver itssue removed from 32 Caucasian male and female fetuses who died at 18-24 weeks gestation due to spontaneous abortion; from kidney tissue removed from 50 Caucasian male and female fetuses who died at 20-33 weeks gestation due to apontaneous abortion; and from brain itssue removed from a Caucasian male fetuse who died at 23 weeks gestation due to fetal demise.
PHOSDNV44	PCR2-TOPOTA	PEIOSDNV44 PCR2_TOPOTO, Il https://www.net.org/net/points/
PLACNOT02	pINCY	Library was constructed using RNA isolated from the placental issue of a Hispanic formale fetus, who was prematurely delivered at 21 weeks' gestation. Serologies of the mother's blood were positive for CMV (cytomegalovirus).
PROSNOT16	pINCY	Library was constructed using RNA isolated from diseased prostate tissue removed from a 68-year-old Caucusian male during arrange arrange in the associated through indicated adendromous hyperplicits. Pathogogy indicated adendromous hyperplicits. Pathogogy indicated and adenocationous (Glescon gade 3-4). The patient presented with devasted prostate specific antigen (PSA). During this hospitalization, the patient was diagnosed with myasthenia gravis. Patient history included osteoarthritis, and type II diabetes. Physical physperension, acute myocardial infraction, hyperlipidemia, and arteriosclerotic coronary artery disease.

Library	Vector	Library Description
MT05	pINCY	The library was constructed using RNA isolated from diseased prostate tissue removed from a 55-year-old Caucasian male defining a netale, prostate course, regional trapil node of succession and testing a netale, prostate course, regional trapil node of succession and contained adenoifmentations hyperplasts, a thublogy for the associated tumor tissue indicated adenocationna. Glesson grade 51-4, forming a predominant mass involving the left side peripherally with extension into the right posterior superior region. The term of the more riveded and performed the capsule to involve protostate tissue in helder josterior superior region. The left indicated and superior posterior superior region. The left indicates a superior region and margins were positive. One (of 9) left perior by mpt nodes was measuratedly involved. The patient presented with elevated prostate specific antigen (PSA). Patient history included calculus of the kidney. Family history included breast cancer and hung cancer.
PROTDNV21	PCR2-TOPOTA	PCR2-TOPOTA, Library was constructed using pooled QDNA from different donors, cDNA was generated using mRNA isolated from pooled small insertine itseiser emoved from a Cancasian male feats (donor B) who died from feat demise; from plean tumor frisate from prename birth; from ling lisase removed from a 2-tunesian male feats (donor B) who died from feat demise; from plean tumor fissue removed from a 2-tunesian male feats (donor B) who died from feat demise; from plean tumor fissue removed from a 2-tunesian featale (donor D) during a complete penamonecutory; from frontigherited Itsnin tumor tissue removed from a 2-typer-old Cancasian male (donor E) during partial beparentemy; from produptient lettale morner from converted from a 2-typer-old Cancasian male feats (donor P) during partial beparentemy; from produptient lettale tumor from from issue removed from a 2-typer-old Cancasian male feats (donor P) who died at 22 weeks' gestation from promature birth; from pooled feat lettiney tissue removed from a 2-typer-old (donor P) who died at 22 weeks' gestation from premature birth; from pooled feat listiney tissue removed from a 2-typer-old.
		inale and female fetures who died from aponianeous abortion; from pooled thymus tistase removed from \$7.82-year-old male and female denotes who died from saided eaths, and from pooled fetal live istuse removed from \$2.18.24 weak-old male and female female fetaless. For donor 4, sologies were regarive. Farmly history included diabetes in the mother. For donor 8, secropies were regarive. Farmly history included diabetes in the mother. For donor 8, secropies forming a bosellated mass replacing the right lower lobe and a portion of the middle lobe. Multiple notalise comprising the immor above near total necrosis. Smooth massle actin was positive. Eatogen neeptor was regarive and progesterone receptor was positive. The Anexone are sufficient to require the surface of the pool in the surface of t

	X74	Literary Decomposition
And y	Action	Interpretation of hing beloin, and appendectionty. Patient medications included Megace, tamoxifier, and Pepcid Family history included multiple sciencis in the modern, alterosterio coronary artery discuss and type I dichest in the father, and release concerning the sciences in the modern alterosterio coronary artery giesses and type I dichest in the father, and the camer in the grampheme(b). For donor D, publicing inferior temocrated multi-mor with advanced ganglicanic camer in the grampheme(b). For donor D, publicing inferior temocrated multiple and some productive control between the productive control between the productive control and the productive control
		robacco abuse in remission. Previous surgeries included destruction of a pancreatic leston, closed prostatic biopsy, transardent postsubectony, removed of bilatest lesses and total spleadourly. Pleatest included bisinchi, Bytrin, Proseas, Econim, and insulial. Easily sistory included attenselectoric concury ratery disease and earner mouther, attentocaterotic concury artery disease and seaten spocearful infraction in the mother; attentocaterotic concury artery disease and type II diabetes in the father. For donor F and G, Serologies were negative for both donors and family history for donor O included diabetes in the mother.
SCOMDIT01	pINCY	Library was constructed using RNA isolated from diseased spinal cord tissue removed from the base of the medulia of a 57- year-old Causasian male who died from a corebrovascular accident. Patient history included Huntington's disease and emphysema.
SININOT04	pINCY	Library was constructed using RNA isolated from diseased item tissue obtained from a 26-year-old Cancasian male during a partial coloratory, permanent cobstancy, and an includent appendencion; Pathology indicated moderately to severely active (Cohn's disease, Family ingroy include enterities of the small insestine.
SINTBST01	pINCY	Library was constructed using RNA isolated from item tesse obtained from an 18-year-old Cataussian female during bowel mansionness, britchiegh Undersid Catobri 8 disease of the item, involving 15 cm of the small bowel. Family history included creebyovascular disease and alterosed-corie conomy artery disease.
THP1NOB01	PBLUESCRIPT	PBLUESCRIPT Library was constructed using RNA isolated from cultured, unstimulated THP-1 cells. THP-1 is a human promonoxypte line derived from the peripheral blood of a L-year-old Caucasian male with acute monocypte leukemia (sef. Int. J. Cancer (1980) 26; I71.

Library	Vector	Library Description
THYRNOT03 pINCY	pINCY	Library was constructed using RNA isolated from thyroid tissue removed from the left thyroid of a 28-year-old Caucasian
		female during a complete thyroidectomy. Pathology indicated a small nodule of adenomatous hyperplasia present in the left
		thyroid. Pathology for the associated tumor tissue indicated dominant follicular adenoma, forming a well-encapsulated mass in
		the left thymid.
THYRNOT10 pINCY	pINCY	Library was constructed using RNA isolated from the diseased left thyroid tissue removed from a 30-year-old Caucasian
		female during a unilateral thyroid lobectomy and parathyroid reimplantation. Pathology indicated lymphocytic thyroiditis.

Program	Description	Reference	Parameter Threshold
ABI FACTURA	A program that removes vector sequences and masks Applied Biosystems, Foster City, CA ambiguous bases in nucleic acid sequences.	Applied Biosystems, Foster City, CA.	
ABI/PARACEL FDF	A Fast Data Finder useful in comparing and annotating amino acid or nucleic acid sequences.	Applied Biosystems, Foster City, CA; Paracel Inc., Pasadena, CA.	Mismatch <50%
ABI AutoAssembler	A program that assembles nucleic acid sequences.	Applied Biosystems, Foster City, CA.	
BLAST	A Basic Local Aligument Search Tool useful in Additional, S.F. et al. (1990) J. Mol. Biol. sequence suitality each rea mainto stand an medical 215.405-410; Altschul S.F. et al. (1997) and sequences. BLAST includes free functions: Nucleic Additional Search Searc		ESTs: Probability value = 1.0B-8 or less; Pull Length sequences: Probability value = 1.0B-10 or less
FASTA	A Pearson and Lipram algorithm that searches for similarity between a query sequence and a group of sequences of the same type. FASTA comprises as least five functions: fasta, ffasta, fastx, tfastx, and ssearch.	Pearson, W.R. and D.J. Lipman (1988) Proc. EST's: fasts B' value = 1.00E-6; Natl. Acad Sci. USA 88:244-2448; Pearson, Ascembled ESTs: fixet Identity W.R. (1990) Methods Enzymol. 185:62-98; = 95% or genter and March and Smith, T.F. and M.S. Waterman (1981) longth = 200 bases or geneter; Adv. Appl. Math. 2-482-489.  Fig. 1. Acad Sci. USA 8 Sci. 1981 Sci. 1982 Sci. 1982 Sci. 1982 Sci. 1983 Full Length sequences: fatx score = 100 or geneter	ESTS: fasta E value = 1.06E-6; Assembled ESTS: fasta Identity = 95% or greater and Match length = 200 bases or greater; fastx E value = 1.0E-8 or less; Full Length sequences: fastx score = 100 or greater
BLMPS	A BLocks IMProved Searcher that manches a Heniverft. S. and f.O. Henizhoff (1991) sequence against those in BLOCKS, PRINTS, Nucleic Acids Res. 196565-6732. Henixoff DOMO, PRODOM, and PFAM detabases to search   I.G. and S. Henixoff (1996) Methods for gene families, sequence homology, and structural Enzymol. 26688-105; and Attwood, T.K. et fingerprint regions.	Henkerff, S. and J.G. Henkerf (1991) [Nucleic Acids Res. 196505-6572; Henkerff, Ises J.G. and S. Henkloff (1996) Methods [Enzymol, 266:88-105; and Altwood, T.K. et el. al. (1997) J. Chem. Inf. Compatt Sci. 37/417- 424.	Probability value = 1.0B-3 or less

Program	Description	Reference	Parameter Threshold
HMMER	An algorithm for searching a query sequence against	Krogh, A. et al. (1994) J. Mol. Biol.	PFAM, INCY, SMART or
	mudeli Markov model (Anama)-based databases of protein family consensus sequences, such as PFAM,	(1988) Nucleic Acids Res. 26:320-322;	value = 1.0E-3 or less; Signal
	INCY, SMART and TIGRFAM.	Durbin, R. et al. (1998) Our World View, in	peptide hits: Score = 0 or greater
		a Nutshell, Cambridge Univ. Press, pp. 1-	
Destilation	An alreadistry that canadian for eterocent and	Geibebon M at al (1988) CABIOS 4-61-66: Normalized quality score > GCG	Normalized amplity score > GCG
riomescan	sequence motify in protein sequences that match	Gribskov, M. et al. (1989) Methods	specified "HIGH" value for that
	sequence patterns defined in Prosite.	et al.	particular Prosite motif.
		(1997) Nucleic Acids Res. 25:217-221.	Generally, score = 1.4-2.1.
Phred	A base-calling algorithm that examines automated	Ewing, B. et al. (1998) Genome Res. 8:175-	
	sequencer traces with high sensitivity and probability. 185; Ewing, B. and P. Green (1998) Genome	185; Ewing, B. and P. Green (1998) Genome	
		Res. 8:186-194.	
Phrap	A Phils Revised Assembly Program including	Smith, T.F. and M.S. Waterman (1981) Adv. Score = 120 or greater; Match	Score = 120 or greater; Match
	SWAT and CrossMatch, programs based on efficient Appl. Math. 2:482-489; Smith, T.F. and	Appl. Math. 2:482-489; Smith, T.F. and	length = 56 or greater
	implementation of the Smith-Waterman algorithm,	M.S. Waterman (1981) J. Mol. Biol. 147:195-	
	useful in searching sequence homology and	197; and Green, P., University of	
	assembling DNA sequences.	Washington, Seattle, WA.	
Consed	A graphical tool for viewing and editing Phrap	Gordon, D. et al. (1998) Genome Res. 8:195-	
	assemblies.	202.	
SPScan	A weight matrix analysis program that scans protein	Nielson, H. et al. (1997) Protein Engineering Score = 3.5 or greater	Score = 3.5 or greater
	sequences for the presence of secretory signal	10:1-6; Claverie, J.M. and S. Audic (1997)	
	peptides.	CABIOS 12:431-439.	
TMAP	A program that uses weight matrices to delineate	Persson, B. and P. Argos (1994) J. Mol. Biol.	
	transmembrane segments on protein sequences and	237:182-192; Persson, B. and P. Argos	
	determine orientation.	(1996) Protein Sci. 5:363-371.	

Program	Description	Reference	Parameter Threshold
TMHMMBR	A program ultar usa in dated makeyo model (IRMA) Standamaner, EL, et al. (1998) Prec. Sixth of defined transmendrane segments on protein ind. Conf. On intelligence (Sperms for Mon.) sequences and determine orientation.    Ind. Conf. On intelligence (AAAI) Press, for Artificial Intelligence (AAAI) Press, Monto Park, CA, and MIT Press, Cambridge Monto Park, CA, and MIT Press, Cambridge MA, pp. 175-182.	Sonnhammer, E.L. et al. (1998) Proc. Sixth lintl. Conf. On Intelligency Systems for Mol. Biol., Clasgow et al., eds. The Am. Assoc. for Artificial Intelligence (AAAI) Press, Mento Park, CA, and MIT Press, Cambridge, MA, pp. 175-182.	
Motifs	A program that searches amino acid sequences for patterns that matched those defined in Prosite.	Bairoch, A. et al. (1997) Nucleic Acids Res. 25:217-221; Wisconsin Package Program Manual, version 9, page M51-59, Genetics Computer Group, Madison, WI.	

Table 8

			_		_					_	_	_	_						_	_				-	_	_	_	_
Hispanic Allele 1 frequency	p/u	n/a	n/a	p/u	p/u	p/u	0.92	n/a	0.92	n/a	n/a	n/a	0.31	n/a	n/a	n/a	n/a	n/a	n/a	n/a	n/a	0.93	n/a	n/a	n/a	n/a	n/a	n/a
Asian Allele 1 frequency	p/u	n/a	n/a	p/u	p/u	p/u	0.93	n/a	0.93	n/a	n/a	n/a	0.31	n/a	n/a	n/a	n/a	n/a	n/a	n/a	n/a	96'0	n/a	n/a	n/a	n/a	n/a	u/a
African Allele 1 frequency	p/u	n/a	n/a	p/u	p/u	p/u	86'0	n/a	86.0	n/a	n/a	n/a	0.49	n/a	n/a	n/a	n/a	n/a	n/a	n/a	n/a	6.0	n/a	n/a	n/a	n/a	n/a	1/0
Caucasian Allele 1 frequency	p/u	n/a	p/a	p/u	p/u	p/u	66'0		66'0	n/a	n/a	n/a	0.36		p/u	n/a	p/u	n/a	0.36	8.0	n/a	76.0	n/a	n/a	n/a	n/a	p/u	n/a
Allele Amino Acid	noncoding	P202	G105	noncoding	noncoding	noncoding		L106	1	noncoding		G468	L488	D345	L488	D521	Y489	L347	noncoding	noncoding	66A	A179	L123	A158	175	SS 6S	noncoding	411
Allele 2	A	Ü	ı	A	A	V V	A	L	Ā	T	T	C	T	C	o o	Ē	ر ن	C	A I		/ V	Ą		g G	ົ	C	H	-
Allele 1	U	Ĺ	ر د	C	O	C	G	U	b	٥	٥	ī	ر د	T	T	U	T	_ L	G	Ü	G	C	T	C	ī	T	ن	c
EST Allele	U	ī	٥	S	U	o	V		b	c	C	Ü	Ü	L	ī	Ü	T	C	A		G	U	T	C	T	T	U	ر
SNP	1470	199	376	1471	1476	1472	915	377	916	13	904	1454	1514	1085	1514	1613	1515	1089	9/	42	318	557	409	514	265	99	1081	73
SNP	10	178	202	258	10	241	147	415	156	13	26	86	158	195	95	130	57	143	30	24	83	134	53	158	27	28	382	35
SNP ID	SNP00039681	SNP00115026	SNP00111942	SNP00039681	SNP00039681	SNP00039681	SNP00060558	SNP00111942 415	SNP00060558 156	SNP00124167	SNP00061413	SNP00009248 98	SNP00057378 158	SNP00037263	SNP00011634	SNP00068404 130	SNP00011634 57	SNP00037263 143	SNP00009036	SNP00036880 24	SNP00044235 83	SNP00044236	SNP00132159	SNP00132160	SNP00003186	SNP00132158	SNP00092454	SNP00003185
ESTID	1260704H1	1391688H1	1399006H1	1849722T6	2132554R6	2829206T6	3412913H1	7423309T1	8038111H1	1443673F6	1443673R1		1721376H1	1809703F6	1919212H1	2372004H1	7385231H1	776158131	1672333H1	3448924H1	2377009H1	2851653H1	1225412H1	1225412H1	1282158F6	1321651F6	1326847F6	1444147H1
PID	7511289	7511289	7511289	7511289	7511289	7511289	7511289	7511289	7511289	7511056	7511056	7511567	7511567	7511567	7511567	7511567	7511567	7511567	7511651	7511651	7512181	7512181	7511726	7511726	7511726	7511726	7511726	7511726
S B S	39	39	39	93	88	39	39	39	39	40	04	141	4	41	14	41	4	41	42	42	4	4	45	5	45	45	45	45

Table 8

	_		_			_																						
Hispanic Allele 1 frequency	n/a	n/a	n/a	n/a	n/a	n/a	n/a	n/a	n/a	n/a	n/a	n/a	n/a	p/u	n/a	n/a	p/u	n/a	n/a	n/a	p/u	n/a	7.0	n/a	n/a	n/a	n/a	2/4
Asian Allele I frequency	n/a	n/a	n/a	n/a	n/a	n/a	n/a	n/a	n/a	n/a	n/a	n/a	n/a	p/u	n/a	n/a	p/u	n/a	n/a	n/a	p/u	n/a	0.44	n/a	n/a	n/a	n/a	4
African Allele 1 frequency	n/a	n/a	n/a	n/a	n/a	n/a	n/a	n/a	n/a	n/a	n/a	n/a	n/a	p/u	n/a	n/a	p/u	n/a	n/a	n/a	p/u	n/a	0.51	n/a	n/a	n/a	n/a	n/a
Caucasian Allele 1 frequency	n/a	p/u	p/u	n/a	p/u	p/u	n/a	n/a	n/a	n/a	n/a	n/a		p/u	n/a		p/u	n/a	n/a	n/a	p/u	n/a	0.57	n/a	n/a	n/a	n/a	9/4
Allele Amino Acid	noncoding	noncoding	noncoding	D79	noncoding	noncoding	V81	L129	A164	A11	175	8	L123	1260	noncoding	Г	A259	noncoding		P165	P261	G392	oding	1213	1.205	noncoding	noncoding	D354
Allele 2	Γ	T	T			F		CI		T.		C		A	A	T	A A	A L	Г		A			T	C	A	A n	
Allele 1	H	Ü	C		c	٥		T	ن		T		T	Ü	Ö	Ü	C		V		် ပ	T (	U		T	5	G.	۲
EST	F	C	٥	۷	C	د	. I	T		T		T	T	C	A		c	Ö	' V	Ü	C	L	U		T	Ü	G	٦
SN GB	1075	1077	1097	277	1082	1084			532	74	2		410	062	1029	202	788	_		206	794	1403	2040	998	841	1678		1280
SNP	38	82	24	238	38	38		348	453	13	203	9	349	42	82	186	362		130	240	228	69		244	219	23	252	217
O ANS	SNP00003187	SNP00092454	SNP00092454	SNP00147540	SNP00092454	SNP00092454		SNP00132159	SNP00132160	SNP00003185	. 1	SNP00132158	SNP00132159	SNP00035960		SNP00035958	SNP00035960		SNP00035959	SNP00035958	SNP00035960	SNP00011025	SNP00011026 423		SNP00137093	SNP00041731	SNP00041731	SNP00054031
ESTID	1927771T6	3074414T6		3781813HI	4549457T1				762752411	770706211			770706231	1461423H1		2960723F6				5652256H1	_	031545H1	1269946T6		1288103H1 S	1377666F6		1477055H1
ON O	7511726	7511726	7511726	7511726	7511726	7511726	7511726	7511726	7511726	7511726	7511726	7511726	7511726	7511078	7511078	7511078	7511078	7511078	7511078	7511078	7511078	7511505	7511505	7511505	7511505	7511505	7511505	7511505
ğ a ÿ	45		_	45	45							45			47 7	47 7	Ť			47 7			48 7		48	48 7		48 7

Table 8

		-	_	_	_	-		_	_	_		_	_	-	_	_	-	_	_	_	-	_	_	_	_	_	_	_
Hispanic Allele 1 frequency	n/a	0.7	n/a	n/a	n/a	0.86	n/a	n/a	n/a	n/a	n/a	n/a	0.86	n/a	p/u	p/u	p/u	p/u	n/d	n/a	n/d	n/a						
Asian Allele 1 frequency	n/a	0.44	n/a	n/a	n/a	6.0	n/a	n/a	n/a	n/a	n/a	n/a	6.0	n/a	n/d	p/u	, p/u	p/u	p/u	n/a	p/u	n/a						
African Allele 1 frequency	n/a	0.51	n/a	n/a	n/a	0.87	n/a	n/a	n/a	n/a	n/a	n/a	0.87	n/a	66.0	p/u	66.0	0.99	66.0	n/a	p/u	u/a						
Caucasian Allele 1 frequency	n/a	0.57	n/a	n/a	n/a	0.98	n/a	n/a	n/a	n/a	n/a	n/a	86.0	n/a	n/a	n/a	e/u	n/a	n/a	n/a	p/u	p/u	p/u	p/u	p/u	n/a	p/u	1/0
Allele Amino Acid 2	noncoding	noncoding	noncoding	R31	T45	N114	noncoding	R31	1214	L205	T23	T37	A106	S75	Y154	V64	L143	D158	R155	Y96	noncoding	R87	noncoding	noncoding	noncoding	E193	L140	02120
	T	Ŀ	c	G	Ţ	T	T	G	T	c	G	Т	T		၁	C	ပ	T	T	G	G	T	9	Ğ	G	A	Ģ	C
Allele 1	U	C	T	A	ပ	Ü	С	٧	ပ	Ţ	4	C	c	T	T	T	L	U	C	Α	Т	Ü	T	ī	Ţ	Ö	T	c
EST	C	c	ī	G	T	ပ	၁	Ą	ပ	Ţ	4	ပ	)	F	ī	Ŧ	F	U	C	¥	F	ပ	T	Ţ	T	D	H	
SR GB	2162	2039	205	319	362	569	2161	320	867	842	294	337	544	424	629	391	627	477	466	290	791	262	810	208	790	582	422	417
SNP	294		205	49	91	298	298	13	248	273	86	140	347	226	176	185	421	55	316	140	20	113	260	257	10	279	399	212
SNP ID	SNP00011026 294	SNP00011026 214	SNP00041727	SNP00041728	SNP00041729 91	SNP00041730 298	SNP00011026	SNP00041728	SNP00054030	SNP00137093	SNP00041728	SNP00041729	SNP00041730	SNP00011392	SNP00042220	SNP00011392	SNP00042220	SNP00008678	8L9800004NS	SNP00132026	SNP00105006	SNP00035683	SNP00105006 260	SNP00105006	SNP00105006	SNP00058604	SNP00096536	CATDO0120102
ESTID	1649779T6	1737071H1	2484384H1	2512592F6	2512592F6	2512592F6	Г	7255731H2	765024912	765024912	776324601	7763246J1	776324601	1224235H1	1284532H1	7455113H2	7455113H2	1210857R1	1709352F6	1709352H1	1709764T6	1710463H1	1841101H1	1841101R6	1841101T6	6214728H1	6216796H1	021/02/02/03/
OM.	7511505	7511505	7511505	7511505	7511505	7511505	7511505	7511505	7511505	7511505	7511505	7511505	7511505	7511552	7511552	7511552	7511552	7511722	7511722	7511722	7511722	7511722	7511722	7511722	7511722	7511722	7511722	20011200
SEQ ID NO:	84	48	48	84	84	48	48	48	48	84	84	84	84	49	46	49	49	20	20	20	20	20	20	20	20	20	20	T

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Hispanic Allele 1 frequency																												
⊞ < ଞ	n/a	n/a	n/a	n/a	n/a	n/a	n/a	n/a	n/a	n/a	n/a	n/a	n/a	n/a	n/a	n/a	n/a	n/a	0.67	n/a	n/a	0.67	n/a	n/a	n/a	n/a	n/a	n/a
Asian Allele 1 frequency	n/a	n/a	n/a	n/a	n/a	n/a	n/a	n/a	n/a	n/a	n/a	n/a	n/a	n/a	n/a	n/a	n/a	n/a	0.95	n/a	n/a	0.95	n/a	n/a	n/a	n/a	n/a	n/a
African Allele 1 frequency	n/a	n/a	n/a	n/a	n/a	n/a	n/a	n/a	n/a	n/a	n/a	n/a	n/a	n/a	n/a	n/a	n/a	n/a	n/a	n/a	n/a	n/a	n/a	n/a	n/a	n/a	n/a	n/a
Caucasian Allele 1 frequency	n/a	p/u	n/a	n/a	n/a	n/a	n/a	n/a	n/a	n/a	n/a	n/a	n/a	n/a	n/a	n/a	n/a	n/a	0.4	n/a	n/a	0.4	n/a	n/a	n/a	n/a	0.85	n/a
Allele Amino Acid	A204	L333	1265	noncoding	G182	K45	7.7	H47	H9	V286	noncoding	G208	N46	8/8	K45	ల	noncoding	T42	A285	L183	T42	A277	noncoding	noncoding	noncoding		noncoding	noncodina
Allele 2	0	ບ	Ü	Ü	L		ī	ß	T		Ü		Ö		Ö				T		G		C	υ	C		C	
Allele 1	Ą	Ţ	C	H	၁	A	Ç	¥	C	C	Ţ	Ą	A	C	Ą	Ü	T	C	င	C	V	S	¥	Ą	L	A	G.	E
EST Allele	A	Ţ	G	Ŧ	c	A	Т	A	C	G	Ţ	G	A	Ţ	A	T	Т	C	c	c	A	C	Ą	Ą	C	Ą	G	ر
SNP SNP	615	1136	932	1449	683	271	158	712	164	993	1448	759	273	160	272	159	1447	262	166	684	261	296	1403	1409	1604	1630	576	1601
EST	273	232	221	63	19	216	103	190		224		26	196	83	165	52	110	103	238	167	22	63	152	158	62	36	10	~
SNP ID	SNP00140687	SNP00123117	SNP00003188	SNP00003189	SNP00123116	SNP00065489 216	SNP00072434 103	SNP00065489   190	SNP00072434	SNP00003188 224	SNP00003189 19	SNP00152940	SNP00065489	SNP00072434 83	SNP00065489   165	SNP00072434 52	SNP00003189	SNP00152939   103	SNP00105147 238	SNP00123116   167	SNP00065489	SNP00105147	SNP00154888  152	SNP00154889 158	SNP00154890	SNP00154891	SNP00002753 10	SNP00154890
ESTID	6738488H1	000278H1	020177H1	1274759T6	1291827H1	1624459F6	1624459F6	2112792R6	2112792R6	2112792T6	3120473T6	4425660H1	4805658HI	4805658H1	493625R6	493625R6	493625T6	5301835F6	551634T6	1	679957831	7243569H1	125846H1	125846H1	1259618R6	1259618R6	1669737F6	1030120T6
	7511722	7511489	7511489	7511489	7511489	7511489	7511489	7511489	7511489	7511489	7511489	7511489	7511489	7511489	7511489	7511489	7511489	7511489	7511489	7511489	7511489	7511489	7511612	7511612	7511612	7511612	7511612	7511612
g a g	20	51	51	51	51 7	51	51	51	51	51	51	51	51	51 7	51 7	51 7	51	51 7	51 7	51 7	51 7	51	54	54	54	54	54	54

Table 8

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## Hispanic frequency Allele 1 0.93 0.93 n/a n/a n/a n/a n/a a a 8 8 n/a n/a n/a n/a n/a n/a n/a requency Allele 1 Asian 10/a 10/a 10/a 10/a 10/a 10/a n/a 0.91 n/a p/u n/a n/a n/a N/a frequency African Allele 1 98.0 0.97 0.97 n/a n/a ş n/a ₽ a n/a n/a ďa ďa a a a a ďa n/a n/a n/a n/a Caucasian requency Allele 1 0.85 187 0.97 0.81 0.97 0.78 n/a n/a β a a a Na Na n/a n/a ₽ Na n/a n/a Allele Amino Acid noncoding noncoding noncoding noncoding noncoding noncoding stop374 2214 A206 G292 C218 G303 C229 S145 1169 P117 9/./ 9/7 N71 25 X10 P14 N71 Z, A2 υ <u>-</u>-HU < H F Q Q A 4 ⊱ ح <u>ن</u> Allele A A F F D O Allele EST ŒΗ 니이이나 O OH SNP GB 2447 519 363 575 96142 29 29 3 8 2 3 435 299 314 EST SNP 13 8 13 8 8 9 209 224 209 224 388 93 SNP00116769 122 8 22 SNP00123257 SNP00012037 SNP00012036 SNP00044775 SNP00116769 SNP00058613 SNP00058614 SNP00002753 SNP00128389 SNP00013069 SNP00055466 SNP00013069 SNP00101209 SNP00013068 SNP00154889 SNP00154887 SNP00128388 SNP00013067 SNP00055468 SNP00055467 SNP00013067 SNP00055468 SNP00055466 SNP00055467 SNP00123257 SNP ID 3522462R6 631132F6 2101062T6 3803815H1 6604874H1 6604874H1 273292F6 1334850F6 508329F6 2636474H1 2636474H1 273292F6 334850F6 508329F6 2568901H1 2636474H1 2636474H1 3014210H1 2373340T6 3407727H1 3712320F6 004612T6 256663F6 1321148F6 1321148F6 3523116H1 EST] 7511626 7511965 7511965 7511965 7511612 7511624 7511624 7511626 7511626 7511626 7512885 7512403 7512403 7512403 7512403 7511612 7511612 7511612 7511612 7511612 7511612 7511624 7511624 7511624 7511626 7511626 7511626 7512885 ДÖ 8 8 8 8 8 8 8 8 8 8 8 8 8 8 જ્ 20 57

## Table 8

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Hispanic Allele 1 frequency	p/u	n/a	p/u	n/a	n/a	n/a	n/a	n/a	n/a	n/a	n/a	n/a	n/a	n/a	n/a	n/a	n/a	n/a	n/a	n/a	9/11							
Asian Allele 1 frequency	p/u	n/a	p/u	n/a	n/a	n/a	n/a	n/a	n/a	n/a	n/a	n/a	n/a	n/a	n/a	n/a	n/a	n/a	n/a	n/a	n/a							
African Allele 1 frequency	p/u	n/a	p/u	n/a	n/a	n/a	n/a	n/a	n/a	n/a	n/a	n/a	n/a	n/a	n/a	n/a	n/a	n/a	n/a	n/a	-/-							
Caucasian Allele 1 frequency	p/u	n/a	p/u	n/a	n/a	n/a	n/a	n/a	n/a	n/a	n/a	n/a	n/a	n/a	n/a	n/a	n/a	n/a	n/a	n/a	-7-							
Allele Amino Acid Caucasian  2 Allele 1 frequency	noncoding	S311	noncoding	Y375	noncoding	noncoding		ding	DI24	H190	H277	D367	E361		H232	K386	K372	H145	noncoding	noncoding	noncoding	The same of						
Allele J	5	Y S	G I	c l	C	C	υ υ	G	C	5	T	5	C	G	C	T	T	A I	V I	A	T	A	V I	T	T	T	G	E
Allele 1	A	C	Ą	T	Т	Ţ	H	Ą	T	Ą	Ü	Ą	T	V	T	၁	٥	Ð	Ð	Ö	2	Ð	Ð	C	U	Ü	Ą	
EST	A	C	Ą	T	I	Ţ	Ü	Ą	T	Ą	Ü	Ą	T	V	C	T	C	Ð	Y	G	၁	V	A	T	C	C	A	,
SS G	2495	1138	2497	1328	1350	1342	1639	1500	1635	1496	1411	1406	1644	2021	395	597	928	1114	1097	1096	602	1172	1129	450	211	1595	1153	010
SNP	11	232	52	59	147	122	139	114	13	151	132	19	283	145	951	223	38	216	248	132	06	156	194	223	98	219	53	
SNP ID	SNP00058614	SNP00025602 232	SNP00058614	SNP00116769	697911004NS	SNP00116769	SNP00006647	SNP00135814	SNP00006647	SNP00135814	SNP00149378 132	SNP00006646	SNP00006647	SNP00135814 145	SNP00048181 156	SNP00002073	SNP00149574	SNP00002074	SNP00002074	SNP00002074	SNP00149574	SNP00002074	SNP00002074 194	SNP00002073	SNP00009024	SNP00036860	SNP00107326 53	A 000000000000000000000000000000000000
ESTID	1321148H1	1816079F6	9T6116961	2962164T7	961757R6	961757T6	1HL/26100	1385137F6	1385138T6	1385138T6	4111577T6	4594352F6	5764290H1	5764290H1	2044743H1	3595491H1	5845804H1	1832824T6	2079512T6	2434429H1	2654610H1	3152185T6	3187495T6	3595491H1	088573H1	1740626R6	2050911H1	2000112000
OH.	7512403	7512403	7512403	7512403	7512403	7512403	7512564	7512564	7512564	7512564	7512564	7512564	7512564	7512564	7512646	7512707	7512707	7512710	7512710	7512710	7512710	7512710	7512710	7512710	7512884	7512884	7512884	10001
S B S	65	. 65	29	29	65	65	09	8	9	9	9	9	8	9	19	63	8	49	4	64	45	4	2	8	33	59	ß	

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## Table 8

Hispanic	Allele 1	frequency																												
H	7	ag L	p/u	n/a	p/u	n/a	n/a	n/a	n/a	n/a	n/a	n/a	n/a	β	n/a	n/a	n/a	n/a	p/u	n/a										
Asian	Allele 1	frequency	p/u	n/a	p/u	n/a	n/a	n/a	n/a	n/a	n/a	n/a	n/a	p/u	n/a	n/a	n/a	n/a	p/u	n/a										
African	Allele 1	frequency	p/u	п/а	n/a	n/a	n/a	n/a	n/a	n/a	п/а	п/а	п/а	n/a	p/u	n/a	n/a	п/а	п/а	п/а	n/a	n/a	n/a	p/u	n/a	n/a	n/a	n/a	p/u	n/a
Caucasian	Allele 1	frequency	p/u	p/u	n/a	86.0	n/a	n/a	n/a	86.0	n/a	86.0	n/a	p/u	26.0	n/a	n/a	n/a	p/u	р/u	n/a	p/u	p/u	p/u						
Allele Amino Acid			noncoding	noncoding	Г	noncoding	noncoding	R211	T357	noncoding	noncoding	noncoding	Г		noncoding	V43	P14	D223	noncoding	noncoding	noncoding		noncoding	noncoding	noncoding		P90	noncoding	noncoding	
Allele	7		ر د	ī	ر د	T.	G		υ	T I	O	Ţ	G	g				9		Ţ	T		T	T	H		T	T	T.	T
Allele	-		T	ບ	T	G	C	ī	Ą	G	Ü	G	S	Ü	၁	G	o	A	Ţ	G	Ð	c	Ü	ບ	G	C	ပ	ບ	ပ	G
EST	Allele		Ţ	Ţ	Т	G	c	ß	C	G	C	b	C	ບ	ວ	<sub>D</sub>	9	V	Ţ	G	G	၁	ບ	ပ	G	ပ	C	Ţ	ပ	G
BI	SNP		855	808	735	52	75	835	1273	52	75	36	59	372	1946	197	112	738	1249	1745	1739	1478	1449	1403	1760	1499	538	1448	1402	1729
EST	SNP		246	485	13	9	67	17	455	9	29	9	29	160	257	219	299	251	170	111	112	373	204	158	107	368	∞	203	157	121
SNP ID			SNP00036859 246	SNP00048574	SNP00059656	SNP00028270	SNP00028271	SNP00059657	SNP00059658	SNP00028270	SNP00028271	SNP00028270	SNP00028271	SNP00039384	SNP00003729  257		SNP00119494	SNP00119495	SNP00058761	SNP00033300	SNP00033300	SNP00036178	SNP00036178	SNP00105496	SNP00033300	SNP00036178	SNP00075867	SNP00036178	SNP00105496	SNP00033300
ESTID			3247348H1	5512625F6	2257046H1	2615823HI	2615823H1	5970391HI	5970391H1	2615823H1	2615823H1	2615823H1	2615823H1	181246H1	4275007H1	6705322F6	6705322H1	7359891H1	1308817H1	1441063T6	1550727T6	1550727T6	1627381F6	1627381F6	1627381T6	1627381T6	1665743H1	1726046F6	1726046F6	1726046T6
PID			7512884	7512884	7512931	7512931	7512931	7512931	7512931	7512933	7512933	7512942	7512942	7512779	7512779	7512779	7512779	7512779	7512877	7512877	7512877	7512877	7512877	7512877	7512877	7512877	7512877	7512877	7512877	7512877
SEO	А	ö	8		99	99	99	99	-			89	89	70	. 0/	. 0/	0,	20	71	71	71	7.1	71	7.1	7.1	7.1	7.1	71	71	7.1

Table 8

			_	_	_			_	_	_	_		_	_	_	_
Hispanic Allele 1 frequency	n/a	n/a	n/a	n/a	n/a	p/u	p/u	p/u	n/a	p/u	n/a	n/a	n/a	n/a	n/a	n/a
Asian Allele 1 frequency	n/a	n/a	n/a	n/a	n/a	p/u	p/u	p/u	n/a	p/u	n/a	n/a	n/a	n/a	n/a	n/a
African Allele 1 frequency	n/a	n/a	n/a	n/a	n/a	p/u	p/u	p/u	n/a	p/u	n/a	n/a	n/a	n/a	n/a	n/a
Caucasian Allele 1 frequency	p/u	p/u	p/u	p/u	p/u	p/u	p/u	p/u	n/a	p/u	n/a	n/a	n/a	n/a	n/a	n/a
BST Allele Allele Amino Acid Caucasian Allele 1 2 Allele 1 Allele 1 Aries Allele 1 A	noncoding	noncoding	noncoding	noncoding	noncoding	noncoding	W17	noncoding	noncoding	noncoding	noncoding	noncoding	noncoding	noncoding	T97	noncoding
Allele 2	ı	ı.	T	T	ī	T	b	C	Ð	C	Т	Ü	C	C	ī	c
Allele 1	b	Ü	G	G	ن	ر	T	T	Ą	T	C	T	T	T	ن	Т
EST	b	Ü	G	G	ت	C	T	T	Ü	ı	C	T	T	T	C	T
SN CB1	1738	1477	1710	1737	1476	1431	354	1016	781	1015	771	1420	1414	1481	339	1403
SNP	115	376	183	135	396		Г	30	257	138	222	378	384	316		
SNP ID	SNP00033300 115	SNP00036178 376	SNP00033300 183	SNP00033300 135	302 SVID00036178	SNP00105496   441	SNP00069120 359	SNP00014852 30	SNP00014851 257	SNP00014852 138	SNP00151594   222	SNP00028980 378	SNP00028980 384	316 316 316	SNP00069824 129	SNP00028980 17
ESTID	1729440T6	1729440T6	1808254H1	781216T6	781216T6	781216T6	7585776H2	1752587T6	2555028F6	391692R6	1363313F6	1363313T6	1364310T6	1984112T6	2109361H1	2909762H1
GIA _	7512877	7512877	7512877	7512877	7512877	7512877	7512794	7512886	7512886	7512886	7512929	7512929	7512929	7512929	7512929	7512929
SEQ B G	71	7.	11	7.1	7.1	71	4	75	75	75	26	92	92	9/	2/2	92

## What is claimed is:

5

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- 1. An isolated polypeptide selected from the group consisting of:
- a) a polypeptide comprising an amino acid sequence selected from the group consisting of SEO ID NO:1-38.
- b) a polypeptide comprising a naturally occurring amino acid sequence at least 90% identical to an amino acid sequence selected from the group consisting of SEQ ID NO:1, SEQ ID NO:3, SEQ ID NO:5, SEQ ID NO:8, SEQ ID NO:11, SEQ ID NO:14-16, SEO ID NO:30, SEO ID NO:32-33, and SEO ID NO:36-37,
- a polypeptide consisting essentially of a naturally occurring amino acid sequence at least 90% identical to an amino acid sequence selected from the group consisting of SEQ ID NO:4, SEQ ID NO:9-10, SEQ ID NO:12-13, SEQ ID NO:17-19, SEQ ID NO:21-22, SEQ ID NO:24, SEQ ID NO:26, SEQ ID NO:28, SEQ ID NO:31, SEQ ID NO:34-35, and SEQ ID NO:38.
- d) a polypeptide comprising a naturally occurring amino acid sequence at least 95% identical to the amino acid sequence of SEQ ID NO:6,
  - a polypeptide comprising a naturally occurring amino acid sequence at least 98% identical to an amino acid sequence selected from the group consisting of SEQ ID NO:7, SEO ID NO:20, SEO ID NO:25, and SEQ ID NO:29,
- 20 f) a polypeptide comprising a naturally occurring amino acid sequence at least 94% identical to an amino acid sequence selected from the group consisting of SEQ ID NO:23 and SEO ID NO:27.
  - a biologically active fragment of a polypeptide having an amino acid sequence selected from the group consisting of SEQ ID NO:1-38, and
  - an immunogenic fragment of a polypeptide having an amino acid sequence selected from the group consisting of SEQ ID NO:1-38.
    - An isolated polypeptide of claim 1 comprising an amino acid sequence selected from the group consisting of SEQ ID NO:1-38.
      - 3. An isolated polynucleotide encoding a polypeptide of claim 1.
        - 4. An isolated polynucleotide encoding a polypeptide of claim 2.

 An isolated polynucleotide of claim 4 comprising a polynucleotide sequence selected from the group consisting of SEO ID NO:39-76.

- A recombinant polynucleotide comprising a promoter sequence operably linked to a
   nolynucleotide of claim 3.
  - 7. A cell transformed with a recombinant polynucleotide of claim 6.
  - 8. A transgenic organism comprising a recombinant polynucleotide of claim 6.

A method of producing a polypeptide of claim 1, the method comprising:

- a) culturing a cell under conditions suitable for expression of the polypeptide, wherein said cell is transformed with a recombinant polynucleotide, and said recombinant polynucleotide comprises a promoter sequence operably linked to a polynucleotide encoding the polypeptide of claim 1, and
- b) recovering the polypeptide so expressed.

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 A method of claim 9, wherein the polypeptide comprises an amino acid sequence selected from the group consisting of SEQ ID NO:1-38.

11. An isolated antibody which specifically binds to a polypeptide of claim 1.

- 12. An isolated polynucleotide selected from the group consisting of:
- a) a polynucleotide comprising a polynucleotide sequence selected from the group consisting of SEQ ID NO:39-76,
- a polynucleotide consisting essentially of a naturally occurring polynucleotide sequence at least 90% identical to a polynucleotide sequence selected from the group consisting of SEQ ID NO:39-40, SEQ ID NO:45-48, SEQ ID NO:50, SEQ ID NO:52-53. SEO ID NO:57, SEO ID NO:59-69, and SEQ ID NO:74-76,
- a polynucleotide comprising a naturally occurring polynucleotide sequence at least
   99% identical to a polynucleotide sequence selected from the group consisting of SEQ
   ID NO:41, SEQ ID NO:44, and SEQ ID NO:55-56,
  - d) a polynucleotide comprising a naturally occurring polynucleotide sequence at least

98% identical to a polynucleotide sequence selected from the group consisting of SEQ ID NO:42 and SEQ ID NO:70,

- a polynucleotide comprising a naturally occurring polynucleotide sequence at least
   93% identical to the polynucleotide sequence of SEO ID NO:43.
- 5 f) a polynucleotide comprising a naturally occurring polynucleotide sequence at least 91% identical to the polynucleotide sequence of SEO ID NO:49.
  - a polynucleotide comprising a naturally occurring polynucleotide sequence at least
     97% identical to the polynucleotide sequence of SEQ ID NO:51,
  - a polynucleotide comprising a naturally occurring polynucleotide sequence at least
     96% identical to a polynucleotide sequence selected from the group consisting of SEQ
     ID NO:54 and SEQ ID NO:58.
    - a polynucleotide comprising a naturally occurring polynucleotide sequence at least 90% identical to a polynucleotide sequence selected from the group consisting of SEQ ID NO:71-73,
    - j) a polynucleotide complementary to a polynucleotide of a),
    - a polynucleotide complementary to a polynucleotide of b),
    - a polynucleotide complementary to a polynucleotide of c),
  - m) a polynucleotide complementary to a polynucleotide of d),
    - a polynucleotide complementary to a polynucleotide of e),
  - a polynucleotide complementary to a polynucleotide of f),
    - p) a polynucleotide complementary to a polynucleotide of g),
    - q) a polynucleotide complementary to a polynucleotide of h),
    - r) a polynucleotide complementary to a polynucleotide of i), and
  - s) an RNA equivalent of a)-r).

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- An isolated polynucleotide comprising at least 60 contiguous nucleotides of a polynucleotide of claim 12.
- A method of detecting a target polynucleotide in a sample, said target polynucleotide
   having a sequence of a polynucleotide of claim 12, the method comprising:
  - hybridizing the sample with a probe comprising at least 20 contiguous nucleotides comprising a sequence complementary to said target polynucleotide in the sample, and which probe specifically hybridizes to said target polynucleotide, under conditions

whereby a hybridization complex is formed between said probe and said target polynucleotide or fragments thereof, and

 detecting the presence or absence of said hybridization complex, and, optionally, if present, the amount thereof.

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- 15. A method of claim 14, wherein the probe comprises at least 60 contiguous nucleotides.
- 16. A method of detecting a target polynucleotide in a sample, said target polynucleotide having a sequence of a polynucleotide of claim 12, the method comprising:
  - amplifying said target polynucleotide or fragment thereof using polymerase chain reaction amplification, and
    - detecting the presence or absence of said amplified target polynucleotide or fragment thereof, and, optionally, if present, the amount thereof.
- 15 17. A composition comprising a polypeptide of claim 1 and a pharmaceutically acceptable excipient.
  - A composition of claim 17, wherein the polypeptide comprises an amino acid sequence selected from the group consisting of SEQ ID NO:1-38.

- 19. A method for treating a disease or condition associated with decreased expression of functional ENZM, comprising administering to a patient in need of such treatment the composition of claim 17.
- 25 20. A method of screening a compound for effectiveness as an agonist of a polypeptide of claim 1. the method comprising:
  - a) contacting a sample comprising a polypeptide of claim 1 with a compound, and
  - detecting agonist activity in the sample.
- 30 21. A composition comprising an agonist compound identified by a method of claim 20 and a pharmaceutically acceptable excipient.
  - 22. A method for treating a disease or condition associated with decreased expression of

functional ENZM, comprising administering to a patient in need of such treatment a composition of claim 21.

- A method of screening a compound for effectiveness as an antagonist of a polypeptide of
   claim 1, the method comprising:
  - a) contacting a sample comprising a polypeptide of claim 1 with a compound, and
  - detecting antagonist activity in the sample.

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- A composition comprising an antagonist compound identified by a method of claim 23 and
   a pharmaceutically acceptable excipient.
  - 25. A method for treating a disease or condition associated with overexpression of functional ENZM, comprising administering to a patient in need of such treatment a composition of claim 24.
- 15 ... 26. A method of screening for a compound that specifically binds to the polypeptide of claim 1. the method comprising:
  - a) combining the polypeptide of claim 1 with at least one test compound under suitable conditions, and
  - detecting binding of the polypeptide of claim 1 to the test compound, thereby identifying a compound that specifically binds to the polypeptide of claim 1.
  - 27. A method of screening for a compound that modulates the activity of the polypeptide of claim 1, the method comprising:
    - a) combining the polypeptide of claim 1 with at least one test compound under conditions
      permissive for the activity of the polypeptide of claim 1,
    - assessing the activity of the polypeptide of claim 1 in the presence of the test compound, and
    - c) comparing the activity of the polypeptide of claim 1 in the presence of the test compound with the activity of the polypeptide of claim 1 in the absence of the test compound, wherein a change in the activity of the polypeptide of claim 1 in the presence of the test compound is indicative of a compound that modulates the activity of the polypeptide of claim 1.

28. A method of screening a compound for effectiveness in altering expression of a target polynucleotide, wherein said target polynucleotide comprises a sequence of claim 5, the method comprising:

- contacting a sample comprising the target polynucleotide with a compound, under conditions suitable for the expression of the target polynucleotide,
  - b) detecting altered expression of the target polynucleotide, and

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- comparing the expression of the target polynucleotide in the presence of varying amounts of the compound and in the absence of the compound.
- 29. A method of screening for potential toxicity of a test compound, the method comprising:
  - a) treating a biological sample containing nucleic acids with the test compound,
  - b) hybridizing the nucleic acids of the treated biological sample with a probe comprising at least 20 contiguous nucleotides of a polynucleotide of claim 12 under conditions whereby a specific hybridization complex is formed between said probe and a target polynucleotide in the biological sample, said target polynucleotide comprising a polynucleotide sequence of a polynucleotide of claim 12 or fragment thereof,
  - c) quantifying the amount of hybridization complex, and
  - d) comparing the amount of hybridization complex in the treated biological sample with the amount of hybridization complex in an untreated biological sample, wherein a difference in the amount of hybridization complex in the treated biological sample indicates potential toxicity of the test compound.
- 30. A method for a diagnostic test for a condition or disease associated with the expression of ENZM in a biological sample, the method comprising:
- 25 a) combining the biological sample with an antibody of claim 11, under conditions suitable for the antibody to bind the polypeptide and form an antibody:polypeptide complex, and
  - detecting the complex, wherein the presence of the complex correlates with the presence of the polypeptide in the biological sample.
  - 31. The antibody of claim 11, wherein the antibody is:
  - a) a chimeric antibody,
  - b) a single chain antibody,

c) a Fab fragment,

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- d) a F(ab'), fragment, or
- e) a humanized antibody.
- 5 32. A composition comprising an antibody of claim 11 and an acceptable excipient.
  - 33. A method of diagnosing a condition or disease associated with the expression of ENZM in a subject, comprising administering to said subject an effective amount of the composition of claim 32.

34. A composition of claim 32, further comprising a label.

- 35. A method of diagnosing a condition or disease associated with the expression of ENZM in a subject, comprising administering to said subject an effective amount of the composition of claim 15 34.
  - 36. A method of preparing a polyclonal antibody with the specificity of the antibody of claim 11, the method comprising:
    - immunizing an animal with a polypeptide consisting of an amino acid sequence selected from the group consisting of SEQ ID NO:1-38, or an immunogenic fragment thereof, under conditions to elicit an antibody response,
      - b) isolating antibodies from the animal, and
    - c) screening the isolated antibodies with the polypeptide, thereby identifying a polyclonal antibody which specifically binds to a polypeptide comprising an amino acid sequence selected from the group consisting of SEQ ID NO:1-38.
      - 37. A polyclonal antibody produced by a method of claim 36.
      - 38. A composition comprising the polyclonal antibody of claim 37 and a suitable carrier.
  - 39. A method of making a monoclonal antibody with the specificity of the antibody of claim 11, the method comprising:
    - a) immunizing an animal with a polypeptide consisting of an amino acid sequence

selected from the group consisting of SEQ ID NO:1-38, or an immunogenic fragment thereof, under conditions to elicit an antibody response,

- b) isolating antibody producing cells from the animal,
- fusing the antibody producing cells with immortalized cells to form monoclonal antibody-producing hybridoma cells,
- d) culturing the hybridoma cells, and
- e) isolating from the culture monoclonal antibody which specifically binds to a
  polypeptide comprising an amino acid sequence selected from the group consisting of
  SEO ID NO:1-38.

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- 40. A monoclonal antibody produced by a method of claim 39.
- 41. A composition comprising the monoclonal antibody of claim 40 and a suitable carrier.
- 42. The antibody of claim 11, wherein the antibody is produced by screening a Fab expression library.
  - 43. The antibody of claim 11, wherein the antibody is produced by screening a recombinant immunoglobulin library.

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- 44. A method of detecting a polypeptide comprising an amino acid sequence selected from the group consisting of SEQ ID NO:1-38 in a sample, the method comprising:
  - incubating the antibody of claim 11 with the sample under conditions to allow specific binding of the antibody and the polypeptide, and
  - detecting specific binding, wherein specific binding indicates the presence of a
    polypeptide comprising an amino acid sequence selected from the group consisting of
    SEO ID NO:1-38 in the sample.
- 45. A method of purifying a polypeptide comprising an amino acid sequence selected from 30 the group consisting of SEQ ID NO:1-38 from a sample, the method comprising:
  - a) incubating the antibody of claim 11 with the sample under conditions to allow specific binding of the antibody and the polypeptide, and
  - b) separating the antibody from the sample and obtaining the purified polypeptide

comprising an amino acid sequence selected from the group consisting of SEQ ID NO:1-38.

- 46. A microarray wherein at least one element of the microarray is a polynucleotide of claim 5 13.
  - 47. A method of generating an expression profile of a sample which contains polynucleotides, the method comprising:
    - a) labeling the polynucleotides of the sample,

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- contacting the elements of the microarray of claim 46 with the labeled polynucleotides
  of the sample under conditions suitable for the formation of a hybridization complex,
  and
  - c) quantifying the expression of the polynucleotides in the sample.
- 48. An array comprising different nucleotide molecules affixed in distinct physical locations on a solid substrate, wherein at least one of said nucleotide molecules comprises a first oligonucleotide or polynucleotide sequence specifically hybridizable with at least 30 contiguous nucleotides of a target polynucleotide, and wherein said target polynucleotide is a polynucleotide of claim 12.
- 49. An array of claim 48, wherein said first oligonucleotide or polynucleotide sequence is completely complementary to at least 30 contiguous nucleotides of said target polynucleotide.
  - 50. An array of claim 48, wherein said first oligonucleotide or polynucleotide sequence is completely complementary to at least 60 contiguous nucleotides of said target polynucleotide.
  - 51. An array of claim 48, wherein said first oligonucleotide or polynucleotide sequence is completely complementary to said target polynucleotide.
    - 52. An array of claim 48, which is a microarray.
  - 53. An array of claim 48, further comprising said target polynucleotide hybridized to a nucleotide molecule comprising said first oligonucleotide or polynucleotide sequence.

54. An array of claim 48, wherein a linker joins at least one of said nucleotide molecules to said solid substrate.

- 55. An array of claim 48, wherein each distinct physical location on the substrate contains multiple nucleotide molecules, and the multiple nucleotide molecules at any single distinct physical location have the same sequence, and each distinct physical location on the substrate contains nucleotide molecules having a sequence which differs from the sequence of nucleotide molecules at another distinct physical location on the substrate.
  - 56. A polypeptide of claim 1, comprising the amino acid sequence of SEQ ID NO:1.

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- 57. A polypeptide of claim 1, comprising the amino acid sequence of SEQ ID NO:2.
- 58. A polypeptide of claim 1, comprising the amino acid sequence of SEQ ID NO:3.
- 59. A polypeptide of claim 1, comprising the amino acid sequence of SEQ ID NO:4.
- 60. A polypeptide of claim 1, comprising the amino acid sequence of SEQ ID NO:5.
- 61. A polypeptide of claim 1, comprising the amino acid sequence of SEQ ID NO:6.
  - 62. A polypeptide of claim 1, comprising the amino acid sequence of SEQ ID NO:7.
  - 63. A polypeptide of claim 1, comprising the amino acid sequence of SEQ ID NO:8.
  - 64. A polypeptide of claim 1, comprising the amino acid sequence of SEQ ID NO:9.
  - 65. A polypeptide of claim 1, comprising the amino acid sequence of SEQ ID NO:10.
- 66. A polypeptide of claim 1, comprising the amino acid sequence of SEQ ID NO:11.
  - 67. A polypeptide of claim 1, comprising the amino acid sequence of SEQ ID NO:12.

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68. A polypeptide of claim 1, comprising the amino acid sequence of SEQ ID NO:13. 69. A polypeptide of claim 1, comprising the amino acid sequence of SEQ ID NO:14. 70. A polypeptide of claim 1, comprising the amino acid sequence of SEQ ID NO:15. 71. A polyneptide of claim 1, comprising the amino acid sequence of SEO ID NO:16. 72. A polypeptide of claim 1, comprising the amino acid sequence of SEQ ID NO:17. 73. A polypeptide of claim 1, comprising the amino acid sequence of SEQ ID NO:18. 74. A polypeptide of claim 1, comprising the amino acid sequence of SEQ ID NO:19. A polypeptide of claim 1, comprising the amino acid sequence of SEQ ID NO:20. 76. A polypeptide of claim 1, comprising the amino acid sequence of SEQ ID NO:21. 77. A polypeptide of claim 1, comprising the amino acid sequence of SEQ ID NO:22. 78. A polypeptide of claim 1, comprising the amino acid sequence of SEQ ID NO:23. 79. A polypeptide of claim 1, comprising the amino acid sequence of SEQ ID NO:24. 80. A polypeptide of claim 1, comprising the amino acid sequence of SEQ ID NO:25. 81. A polypeptide of claim 1, comprising the amino acid sequence of SEO ID NO:26. 82. A polypeptide of claim 1, comprising the amino acid sequence of SEO ID NO:27. 83. A polypeptide of claim 1, comprising the amino acid sequence of SEQ ID NO:28.

84. A polypeptide of claim 1, comprising the amino acid sequence of SEQ ID NO:29.

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85. A polypeptide of claim 1, comprising the amino acid sequence of SEQ ID NO:30. 86. A polypeptide of claim 1, comprising the amino acid sequence of SEQ ID NO:31. 87. A polypeptide of claim 1, comprising the amino acid sequence of SEQ ID NO:32. 88. A polypeptide of claim 1, comprising the amino acid sequence of SEO ID NO:33. 89. A polypeptide of claim 1, comprising the amino acid sequence of SEQ ID NO:34. 90. A polypeptide of claim 1, comprising the amino acid sequence of SEQ ID NO:35. 91. A polypeptide of claim 1, comprising the amino acid sequence of SEQ ID NO:36. 92. A polypeptide of claim 1, comprising the amino acid sequence of SEQ ID NO:37. 93. A polypeptide of claim 1, comprising the amino acid sequence of SEQ ID NO:38. 94. A polynucleotide of claim 12, comprising the polynucleotide sequence of SEQ ID NO:39. 95. A polynucleotide of claim 12, comprising the polynucleotide sequence of SEQ ID NO:40. 96. A polynucleotide of claim 12, comprising the polynucleotide sequence of SEQ ID NO:41.

97. A polynucleotide of claim 12, comprising the polynucleotide sequence of SEQ ID NO:42.

98. A polynucleotide of claim 12, comprising the polynucleotide sequence of SEQ ID NO:43.

99. A polynucleotide of claim 12, comprising the polynucleotide sequence of SEQ ID NO:44.

100. A polynucleotide of claim 12, comprising the polynucleotide sequence of SEQ ID NO:45.

 $101.\,$  A polynucleotide of claim 12, comprising the polynucleotide sequence of SEQ ID NO:46.

- $$102.\,$  A polynucleotide of claim 12, comprising the polynucleotide sequence of SEQ ID  $$5\,$  NO:47.
  - 103. A polynucleotide of claim 12, comprising the polynucleotide sequence of SEQ ID NO:48.
- 10 104. A polynucleotide of claim 12, comprising the polynucleotide sequence of SEQ ID NO:49.
  - A polynucleotide of claim 12, comprising the polynucleotide sequence of SEQ ID NO:50.

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- $106.\,$  A polynucleotide of claim 12, comprising the polynucleotide sequence of SEQ ID NO:51.
- $$107.\,$  A polynucleotide of claim 12, comprising the polynucleotide sequence of SEQ ID  $$20\,$  NO:52.
  - 108. A polynucleotide of claim 12, comprising the polynucleotide sequence of SEQ ID NO:53.
- 109. A polynucleotide of claim 12, comprising the polynucleotide sequence of SEQ ID NO:54.
  - A polynucleotide of claim 12, comprising the polynucleotide sequence of SEQ ID NO:55.

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 A polynucleotide of claim 12, comprising the polynucleotide sequence of SEQ ID NO:56.

112. A polynucleotide of claim 12, comprising the polynucleotide sequence of SEQ ID NO:57.

- 113. A polynucleotide of claim 12, comprising the polynucleotide sequence of SEQ ID  $_{\odot}$  NO:58.
  - 114. A polynucleotide of claim 12, comprising the polynucleotide sequence of SEQ ID NO:59.
- 10 115. A polynucleotide of claim 12, comprising the polynucleotide sequence of SEQ ID NO:60.
  - A polynucleotide of claim 12, comprising the polynucleotide sequence of SEQ ID NO:61.

 A polynucleotide of claim 12, comprising the polynucleotide sequence of SEQ ID NO:62.

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- $118. \ \ A polynucleotide of claim 12, comprising the polynucleotide sequence of SEQ ID \\ 20 \quad NO:63.$ 
  - 119. A polynucleotide of claim 12, comprising the polynucleotide sequence of SEQ ID NO:64.
- 25 120. A polynucleotide of claim 12, comprising the polynucleotide sequence of SEQ ID NO:65.
  - 121. A polynucleotide of claim 12, comprising the polynucleotide sequence of SEQ ID NO:66.

122. A polynucleotide of claim 12, comprising the polynucleotide sequence of SEQ ID  $\,$  NO:67.

123. A polynucleotide of claim 12, comprising the polynucleotide sequence of SEQ ID NO:68.

- $124. \ \ A polynucleotide of claim 12, comprising the polynucleotide sequence of SEQ ID \\ 5 \quad NO:69.$ 
  - A polynucleotide of claim 12, comprising the polynucleotide sequence of SEQ ID NO:70.
- 10 126. A polynucleotide of claim 12, comprising the polynucleotide sequence of SEQ ID NO:71.
  - 127. A polynucleotide of claim 12, comprising the polynucleotide sequence of SEQ ID NO:72.
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  128. A polynucleotide of claim 12, comprising the polynucleotide sequence of SEQ ID NO:73.
- $129. \ \ A polynucleotide of claim 12, comprising the polynucleotide sequence of SEQ ID \\ 20 \quad NO:74.$ 
  - 130. A polynucleotide of claim 12, comprising the polynucleotide sequence of SEQ ID NO:75.
- 25 131. A polynucleotide of claim 12, comprising the polynucleotide sequence of SEQ ID NO:76.

```
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Gly Arg Thr Arg Ile Arg Trp Ser Arg Cys Cys Thr Met Pro Ser
                                    565
Ser Ser Ala Glu Thr Ala Thr Arg Leu Trp Ser Thr Ser Ser Leu
                575
                                    580
Gly Gly Arg Thr Ser Ala Met Ala Pro Leu Leu Tyr Arg Ala Leu
                590
                                    595
Trp Val Arg Thr Leu Gly Cys Leu Pro Ala Pro Gly Leu Cys Pro
                605
                                    610
Ala Gly Leu Glu Ser His His Cys Leu Pro Trp Ala Tyr Pro Met
                620
                                    625
Arg Pro Ser Asp Ser Thr Thr Ser Leu Arg Arg Gly Trp Arg Gly
                635
                                    640
Gln Ile Ser Glu Arg Leu Ser Cys Glu Leu Leu Asp Pro Pro Leu
                650
                                    655
His Gly Pro Asn Cys Ala Val Cys Pro Leu Ser Tyr Gly Asp
                665
                                    670
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<sup>&</sup>lt;210> 4

<sup>&</sup>lt;211> 237

<sup>&</sup>lt;212> PRT <213> Homo sapiens

<sup>&</sup>lt;220>

<sup>&</sup>lt;221> misc\_feature

<sup>&</sup>lt;223> Incyte ID No: 7511651CD1

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Met Lys Val Leu Leu Leu Lys Asp Ala Lys Glu Asp Asp Cys Gly
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Gln Asp Pro Tyr Ile Arg Leu Ser His Pro Glu Asp Tyr Gly Gly
                 20
Leu Ile Phe Thr Ser Pro Arg Ala Val Glu Ala Ala Glu Leu Cys
                 35
                                     40
Leu Glu Gln Asn Asn Lys Thr Glu Val Trp Glu Arg Ser Leu Lys
                 50
                                     55
Glu Lys Trp Asn Ala Lys Ser Val Tyr Val Val Gly Asn Ala Thr
                 65
                                     70
Ala Ser Leu Val Ser Lys Ile Gly Leu Asp Thr Glu Gly Glu Thr
                                     85
Cys Gly Asn Ala Glu Lys Leu Ala Glu Tyr Ile Cys Ser Arg Glu
                                    100
Ser Ser Ala Leu Pro Leu Leu Phe Pro Cys Gly Asn Leu Lys Arg
                110
                                    115
Glu Ile Leu Pro Lys Ala Leu Lys Asp Lys Gly Ile Ala Met Glu
                125
                                    130
Ser Ile Thr Val Tyr Gln Thr Val Ala His Pro Gly Ile Gln Gly
                140
                                    145
Asn Leu Asn Ser Tyr Tyr Ser Gln Gln Gly Val Pro Ala Ser Ile
                155
                                    160
Thr Phe Phe Ser Pro Ser Gly Leu Thr Tyr Ser Leu Lys His Ile
                170
                                    175
Gln Glu Leu Ser Gly Asp Asn Ile Asp Gln Ile Lys Phe Ala Ala
                                    190
Ile Gly Pro Thr Thr Ala Arg Ala Leu Ala Ala Gln Gly Leu Pro
                200
                                    205
Val Ser Cvs Thr Ala Glu Ser Pro Thr Pro Gln Ala Leu Ala Thr
                215
Gly Ile Arg Lys Ala Leu Gln Pro His Gly Cys Cys
                230
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Met Lys Asp Glu Val Ala Leu Leu Ala Ala Val Thr Leu Leu Gly
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                                     10
Val Leu Leu Gln Ala Tyr Phe Ser Leu Gln Val Ile Ser Ala Arg
                 20
                                     25
Arg Ala Phe Arg Val Ser Pro Pro Leu Thr Thr Gly Pro Pro Glu
                                     40
                 35
Phe Glu Arg Val Tyr Arg Ala Gln Val Asn Cys Ser Glu Tyr Phe
                                     55
                 50
Pro Leu Phe Leu Ala Thr Leu Trp Val Ala Gly Ile Phe Phe His
                                     7.0
                 65
Glu Gly Arg Gly Val Gly Gln Gly Arg Thr Arg Trp Thr Pro Gly
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Thr Arg Ala Gly Arg Ser Pro Gly Pro Cys Val Pro Leu Ala Gly
                                    100
Ala Ala Ala Leu Cys Gly Leu Val Tyr Leu Phe Ala Arg Leu Arg
                110
                                    115
Tyr Phe Gln Gly Tyr Ala Arg Ser Ala Gln Leu Arg Leu Ala Pro
                125
                                    130
Leu Tyr Ala Ser Ala Arg Ala Leu Trp Leu Leu Val Ala Leu Ala
                140
                                    145
Ala Leu Gly Leu Leu Ala His Phe Leu Pro Ala Ala Leu Arg Ala
                155
                                    160
Ala Leu Leu Gly Arg Leu Arg Thr Leu Leu Pro Trp Ala
                                    175
<21.0> 6
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Met Gly Lys Leu Val Ala Leu Val Leu Leu Gly Val Gly Leu Ser
                                     10
Leu Val Gly Glu Met Phe Leu Ala Phe Arg Glu Arg Val Asn Ala
                 20
                                     25
Ser Arg Glu Val Glu Pro Val Glu Pro Glu Asn Cys His Leu Ile
                 35
                                     40
Glu Glu Leu Glu Ser Gly Ser Glu Asp Ile Asp Ile Leu Pro Ser
                 50
                                     55
Gly Leu Ala Phe Ile Ser Ser Gly Leu Lys Tyr Pro Gly Met Pro
                                     70
                 65
Asn Phe Ala Pro Asp Glu Pro Gly Lys Ile Phe Leu Met Asp Leu
                 80
                                     85
Asn Glu Gln Asn Pro Arg Ala Gln Ala Leu Glu Ile Ser Gly Gly
                                    100
                 95
Phe Asp Lys Glu Leu Phe Asn Pro His Gly Ile Ser Ile Phe Ile
                110
                                    115
Asp Lys Asp Asn Thr Val Tyr Leu Tyr Val Val Asn His Pro His
                                    130
                125
Met Lys Ser Thr Val Glu Ile Phe Lys Phe Glu Glu Gln Gln Arg
                                    145
Ser Leu Val Tyr Leu Lys Thr Ile Lys His Glu Leu Leu Lys Ser
                155
                                    160
Val Asn Asp Ile Val Val Leu Gly Pro Glu Gln Phe Tyr Ala Thr
                170
                                    175
Arg Asp His Tyr Phe Thr Asn Ser Leu Leu Ser Phe Phe Glu Met
                185
                                    190
Ile Leu Asp Leu Arg Trp Thr Tyr Val Leu Phe Tyr Ser Pro Arg
                200
                                    205
Glu Val Lys Val Val Ala Lys Gly Phe Cys Ser Ala Asn Gly Ile
                                    220
                215
Thr Val Ser Ala Asp Gln Lys Tyr Phe Ala Ser Arg Met Phe Cys
                                    235
                230
                                                        240
Leu Arg Ser Pro Gly
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                  5
 1
                                     10
Arg Pro Pro Val Arg Cys Pro Ala Trp Arg Pro Phe Ala Ser Gly
                                     25
                 20
Ala Asn Phe Glu Tyr Ile Ile Ala Glu Lys Arg Gly Lys Asn Asn
                 35
                                    40
Thr Val Glv Leu Ile Gln Leu Asn Arg Pro Lvs Ala Leu Asn Ala
                                    55
Leu Cys Asp Gly Leu Ile Asp Glu Leu Asn Gln Ala Leu Lys Ile
                 65
                                    70
Phe Glu Glu Asp Pro Ala Val Gly Ala Ile Val Leu Thr Gly Gly
                 80
                                     85
Asp Lys Ala Phe Ala Ala Gly Ala Asp Ile Lys Glu Met Gln Asn
                                   100
                 95
Leu Ser Phe Gln Asp Cys Tyr Ser Ser Lys Phe Leu Lys His Trp
                110
                                    115
Asp His Leu Thr Gln Val Lys Lys Pro Val Ile Ala Ala Val Asn
                                    130
Gly Tyr Ala Phe Gly Gly Gly Cys Glu Leu Ala Met Met Cys Asp
                                    145
                140
Ile Ile Tyr Ala Gly Glu Lys Ala Gln Phe Ala Gln Pro Glu Ile
                155
                                   160
Leu Ile Gly Thr Ile Pro Gly Ala Gly Gly Thr Gln Arg Leu Thr
                170
                                   175
Arg Ala Val Gly Lys Ser Leu Ala Met Glu Met Val Leu Thr Gly
                185
                                   190
Asp Arg Ile Ser Ala Gln Asp Ala Lys Gln Ala Ala Phe Glu Met
                                    205
                200
Thr Leu Thr Glu Gly Ser Lys Leu Glu Lys Lys Leu Phe Tyr Ser
                215
                                    220
Thr Phe Ala Thr Asp Asp Arg Lys Glu Gly Met Thr Ala Phe Val
                230
                                    235
                                                        240
Glu Lys Arg Lys Ala Asn Phe Lys Asp Gln
                245
<210> 8
<211> 145
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Met Ala Lys Ser Leu Leu Lys Thr Ala Ser Leu Ser Gly Arg Thr
                                    10
                  5
Lys Leu Leu His Gln Thr Gly Leu Ser Leu Tyr Ser Thr Ser His
                 20
                                    25
Gly Phe Tyr Glu Glu Glu Val Lys Lys Thr Leu Gln Gln Phe Pro
                 35
                                    40
Gly Gly Ser Ile Asp Leu Gln Lys Glu Asp Asn Gly Ile Gly Ile
                 50
                                     55
Leu Thr Leu Asn Asn Pro Ser Arg Met Asn Ala Phe Ser Gly Val
                                    70
Met Met Leu Gln Leu Leu Glu Lys Val Ile Glu Leu Glu Asn Trp
                 ٩n
                                     85
Thr Glu Gly Lys Gly Leu Ile Val Arg Gly Ala Lys Asn Thr Phe
                 95
                                    100
Ser Ser Gly Ser Asp Leu Asn Ala Val Lys Ser Leu Gly Thr Pro
                                    115
                110
Glu Thr Ser Phe Asn Lys Cys Cys Ala Gly Ser Arg Leu Gly Ile
                125
                                   130
Gly Trp Arg Ser Arg Ile Tyr Tyr Ser Met
                140
                                    145
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<211> 333
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Met Gly Ser Gln Val Ser Val Glu Ser Gly Ala Leu His Val Val
                                     10
Ile Val Gly Gly Phe Gly Gly Ile Ala Ala Ala Ser Gln Leu
                                     25
                 20
Gln Ala Leu Asn Val Pro Phe Met Leu Val Asp Met Lys Asp Ser
                 35
                                     40
Phe His His Asn Val Ala Ala Leu Arg Ala Ser Val Glu Thr Gly
                                     55
                 50
Phe Ala Lys Lys Thr Phe Ile Ser Tyr Ser Val Thr Phe Lys Asp
                                     70
Asn Phe Arg Gln Gly Leu Val Val Gly Ile Asp Leu Lys Asn Gln
                                     85
                 80
Met Val Leu Leu Gln Gly Gly Glu Val Gln Arg Ser Arg Phe Ile
                                    100
Val Val Val Gly Gly Ser Ala Gly Val Glu Met Ala Ala Glu
                110
                                    115
Ile Lys Thr Glu Tyr Pro Glu Lys Glu Val Thr Leu Ile His Ser
                                    130
                125
Gln Val Ala Leu Ala Asp Lys Glu Leu Leu Pro Ser Val Arg Gln
                                    145
                140
Glu Val Lys Glu Ile Leu Leu Arg Lys Gly Val Gln Leu Leu Leu
                                    160
                155
Ser Glu Arg Val Ser Asn Leu Glu Glu Leu Pro Leu Asn Glu Tyr
                                                        180
                                    175
                 170
```

Arg Glu Tyr Ile Lys Val Gln Thr Asp Lys Gly Thr Glu Val Ala 185 190 Thr Asn Leu Val Ile Leu Cys Thr Gly Ile Lys Ile Asn Ser Ser 200 205 Ala Tyr Arg Lys Ala Phe Glu Ser Arg Leu Ala Ser Ser Gly Ala 215 220 Leu Arg Val Asn Glu His Leu Gln Val Glu Gly His Ser Asn Val 230 235 Tyr Ala Ile Gly Asp Cys Ala Asp Val Arg Thr Pro Lys Met Ala 250 Tvr Leu Ala Glv Leu His Ala Asn Ile Ala Val Ala Asn Ile Val 265 260 Asn Ser Val Lys Gln Arg Pro Leu Gln Ala Tyr Lys Pro Gly Ala 275 Leu Thr Phe Leu Leu Ser Met Gly Arg Asn Asp Gly Val Gly Gln 290 295 Ile Ser Gly Phe Tyr Val Gly Arg Leu Met Val Arg Leu Thr Lys 305 310 Ser Arg Asp Leu Phe Val Ser Thr Ser Trp Lys Thr Met Arg Gln 330 320 325 Ser Pro Pro

<210> 10

<211> 443

<212> PRT

<213> Homo sapiens

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Met Ala Ala Lys Ser Gln Pro Asn Ile Pro Lys Ala Lys Ser Leu 10 Asp Gly Val Thr Asn Asp Arg Thr Ala Ser Gln Gly Gln Trp Gly Arg Ala Cys Cys Ala Leu Thr Gly Pro Val Val Asp Ile Val Thr 40 35 Gly His Ala Arg Leu Ser Asp Ile Trp Ala Lys Thr Pro Pro Ile 50 Thr Arg Lys Ala Ala Gln Leu Tyr Thr Leu Trp Val Thr Phe Gln 70 Val Leu Leu Tyr Thr Ser Leu Pro Asp Phe Cys His Lys Phe Leu 80 85 Pro Gly Tyr Val Gly Gly Ile Gln Glu Gly Ala Val Thr Pro Ala 95 100 Gly Val Val Asn Lys Tyr Gln Ile Asn Gly Leu Gln Ala Trp Leu 110 115 Leu Thr His Leu Leu Trp Phe Ala Asn Ala His Leu Leu Ser Trp 125 130 135 Phe Ser Pro Thr Ile Ile Phe Asp Asn Trp Ile Pro Leu Leu Trp 140 145 150 Cys Ala Asn Ile Leu Gly Tyr Ala Val Ser Thr Phe Ala Met Val 155 160 Lys Gly Tyr Phe Phe Pro Thr Ser Ala Arg Asp Cys Lys Phe Thr

```
175
                170
Gly Asn Phe Phe Tyr Asn Tyr Met Met Gly Ile Glu Phe Asn Pro
                185
                                    190
Arg Ile Gly Lys Trp Phe Asp Phe Lys Leu Phe Phe Asn Gly Arg
                200
                                    205
Pro Gly Ile Val Ala Trp Thr Leu Ile Asn Leu Ser Phe Ala Ala
                215
                                    220
Lys Gln Arg Glu Leu His Ser His Val Thr Asn Ala Met Val Leu
                230
                                    235
Val Asn Val Leu Gln Ala Ile Tyr Val Ile Asp Phe Phe Trp Asn
                245
                                    250
Glu Thr Trp Tyr Leu Lys Thr Ile Asp Ile Cys His Asp His Phe
                260
Gly Trp Tyr Leu Gly Trp Gly Asp Cys Val Trp Leu Pro Tyr Leu
                                    280
Tyr Thr Leu Gln Gly Leu Tyr Leu Val Tyr His Pro Val Gln Leu
                290
                                    295
Ser Thr Pro His Ala Val Gly Val Leu Leu Gly Leu Val Gly
                305
                                    310
TVr Tvr Ile Phe Arg Val Ala Asn His Gln Lys Asp Leu Phe Arg
                320
                                    325
Arg Thr Asp Gly Arg Cys Leu Ile Trp Gly Arg Lys Pro Lys Val
                335
                                    340
Ile Glu Cys Ser Tyr Thr Ser Ala Asp Gly Gln Arg His His Ser
                350
                                    355
Lys Leu Leu Val Ser Gly Phe Trp Gly Val Ala Arg His Phe Asn
                                    370
Tyr Val Gly Asp Leu Met Gly Ser Leu Ala Tyr Cys Leu Ala Cys
                380
                                    385
Gly Gly Gly His Leu Leu Pro Tyr Phe Tyr Ile Ile Tyr Met Ala
                395
                                    400
                                                        405
Ile Leu Leu Thr His Arg Cys Leu Arg Asp Glu His Arg Cys Ala
                410
                                    415
Ser Lys Tyr Gly Arg Asp Trp Glu Arg Tyr Thr Ala Ala Val Pro
                425
                                    430
Tyr Arg Leu Leu Pro Gly Ile Phe
                440
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Met Ser Ser Ser Gly Thr Pro Asp Leu Pro Val Leu Leu Thr Asp
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Leu Lys Ile Gln Tyr Thr Lys Ile Phe Ile Asn Asn Glu Trp His
                 20
                                     25
Asp Ser Val Ser Gly Lys Lys Phe Pro Val Phe Asn Pro Ala Thr
                 35
                                     40
Glu Glu Glu Leu Cys Gln Val Glu Glu Gly Asp Lys Glu Asp Val
                 50
                                      55
```

```
Asp Lys Ala Val Lys Ala Ala Arg Gln Ala Phe Gln Ile Gly Ser
Pro Trp Arg Thr Met Asp Ala Ser Glu Arg Gly Arg Leu Leu Tyr
                80
                                     85
Lys Leu Ala Asp Leu Ile Glu Arg Asp Arg Leu Leu Ala Thr
                95
                                   100
Met Glu Ser Met Asn Gly Gly Lys Leu Tyr Ser Asn Ala Tyr Leu
                110
                                    115
Asn Asp Leu Ala Gly Cys Ile Lys Thr Leu Arg Tyr Cys Ala Gly
                                    130
                125
Trp Ala Asp Lys Ile Gln Gly Arg Thr Ile Pro Ile Asp Gly Asn
                                    145
Phe Phe Thr Tyr Thr Arg His Glu Pro Ile Gly Val Cys Gly Gln
                                    160
                155
Ile Ile Pro Trp Asn Phe Pro Leu Val Met Leu Ile Trp Lys Ile
                                    175
Gly Pro Ala Leu Ser Cys Gly Asn Thr Val Val Val Lys Pro Ala
                185
                                    190
Glu Gln Thr Pro Leu Thr Ala Leu His Val Ala Ser Leu Ile Lys
                200
                                    205
Glu Ala Arg Gly Gly Ile Lys Ala Thr Leu Ser Ser Pro Gln Cys
                215
                                    220
Ser Leu Met Leu Gln Met Arg Cvs Ala Leu Pro Lys Arg Arg Phe
                230
                                    235
Leu Asp Gln Cys Ser Lys Ser
                245
<210> 12
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<212> PRT
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Met Ala Pro Leu Arg Phe Ser Ala Asn Leu Ser Trp Leu Phe Pro
Asp Leu Ser Gly Leu Pro Ala Arg Val Arg Ala Ala Gly Ser Ser
                 2.0
                                     25
Gly Phe Glu Ala Val Glu Val Ala Trp Pro Tyr Ala Glu Thr Pro
                                     40
Glu Ala Leu Ala Arg Ala Ala Arg Glu Ala Gly Leu Arg Leu Val
Leu Ile Asn Thr Pro Pro Gly Asp Gln Glu Lys Gly Glu Met Gly
                 65
Leu Gly Ala Val Pro Gly Arg Gln Ala Ala Phe Arg Glu Gly Leu
                 80
                                     85
Glu Gln Ala Val Arg Tyr Ala Lys Ala Leu Gly Cys Pro Arg Ile
                 95
                                    100
His Leu Met Ala Gly Arg Val Pro Gln Gly Ala Asp Arg Ile Ala
                110
                                    115
Val Lys Ala Glu Met Glu Ala Val Phe Leu Glu Asn Leu Arg His
                125
                                    130
Ala Ala Gly Val Leu Ala Gln Glu Asp Leu Val Gly Leu Leu Glu
```

```
140
                                   145
Pro Ile Asn Thr Arg Ile Thr Asp Pro Gln Tyr Phe Leu Asp Thr
               155
                                   160
Pro Gln Gln Ala Ala Ala Ile Leu Gln Lys Asp Ile Phe His Trp
               170
                                   175
Gln Ile Met Asp Gly Asn Leu Thr Gly Asn Ile Arg Glu Phe Leu
               185
                                   190
Pro Ile Val Gly His Val Gln Val Ala Gln Val Pro Gly Arg Gly
               200
                                   205
Glu Pro Ser Ser Pro Gly Glu Leu Asn Phe Pro Tyr Leu Phe Gln
               215
                                   220
Leu Leu Glu Asp Glu Gly Tyr Lys Gly Phe Val Gly
               230
                                   235
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Leu Val Arg Phe Arg Ile Gly Gly Lys Arg Lys Val Met Pro Lys
                 20
Met Asp Gln Asp Ser Leu Ser Ser Ser Leu Lys Thr Cys Tyr Lys
                 35
Tyr Leu Asn Gln Thr Ser Arg Ser Phe Ala Ala Val Ile Gln Ala
                50
                                     55
Leu Asp Gly Glu Met Arg Asn Ala Val Cys Ile Phe Tyr Leu Val
                                     70
                65
Leu Arg Ala Leu Asp Thr Leu Glu Asp Asp Met Thr Ile Ser Val
                 80
                                     85
Glu Lys Lys Val Pro Leu Leu His Asn Phe His Ser Phe Leu Tyr
                 95
                                   100
Gln Pro Asp Trp Arg Phe Met Glu Ser Lys Glu Lys Asp Arg Gln
                                   115
                110
Val Leu Glu Asp Phe Pro Thr Ile Ser Leu Glu Phe Arg Asn Leu
Ala Glu Lys Tyr Gln Thr Val Ile Ala Asp Ile Cys Arg Arg Met
                140
Gly Ile Gly Met Ala Glu Phe Leu Asp Lys His Val Thr Ser Glu
                155
                                    160
Gln Glu Trp Asp Lys Tyr Cys His Tyr Val Ala Gly Leu Val Gly
                170
                                    175
Ile Gly Leu Ser Arg Leu Phe Ser Ala Ser Glu Phe Glu Asp Pro
                185
                                    190
Leu Val Gly Glu Asp Thr Glu Arg Ala Asn Ser Met Gly Leu Phe
                200
                                   205
Leu Gln Lys Thr Asn Ile Ile Arg Asp Tyr Leu Glu Asp Gln Gln
                215
                                    220
Gly Gly Arg Glu Phe Trp Pro Gln Glu Val Met Ala Ile Ala Thr
                230
                                    235
                                                        240
```

```
Leu Ala Ala Cys Tyr Asn Asn Gln Gln Val Phe Lys Gly Ala Val
                                    250
                245
Lys Ile Arg Lys Gly Gln Ala Val Thr Leu Met Met Asp Ala Thr
                                    265
Asn Met Pro Ala Val Lys Ala Ile Ile Tyr Gln Tyr Met Glu Glu
                                    280
                275
'Ile Tyr His Arg Ile Pro Asp Ser Asp Pro Ser Ser Ser Lys Thr
                                    295
                290
Arg Gln Ile Ile Ser Thr Ile Arg Thr Gln Asn Leu Pro Asn Cys
                305
                                    310
Gln Leu Ile Ser Arg Ser His Tyr Ser Pro Ile Tyr Leu Ser Phe
                                    325
                320
Val Met Leu Leu Ala Ala Leu Ser Trp Gln Tyr Leu Thr Thr Leu
                                    340
Ser Gln Val Thr Glu Asp Tyr Val Gln Thr Gly Glu His
<210> 14
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 Met Ala Ser Arg Gly Arg Arg Pro Glu His Gly Gly Pro Pro Glu
                   5
                                      10
 Leu Phe Tyr Asp Glu Thr Glu Ala Arg Lys Tyr Val Arg Asn Ser
                  20
                                      25
 Arg Met Ile Asp Ile Gln Thr Arg Met Ala Gly Arg Ala Leu Glu
                  35
                                      40
 Leu Leu Tyr Leu Pro Glu Asn Lys Pro Cys Tyr Leu Leu Asp Ile
                  50
                                      55
 Gly Cys Gly Thr Gly Leu Ser Gly Ser Tyr Leu Ser Asp Glu Gly
                  65
                                      70
 His Tyr Trp Val Gly Leu Asp Ile Ser Pro Ala Met Leu Ala Phe
                  80
                                     85
 Leu Leu Cys Ser Gly Ser Val Met Leu Thr Arg Ser Leu Lys Thr
                  95
                                    100
 Leu Pro Ser Ala Cys Thr Ala Phe Leu Leu Phe Phe Leu Phe
                 110
                                     115
 Ser Ser Gly Asp Pro Glu Leu Ser Cys Ser Cys Thr Leu Arg Thr
                                     130
                                                         135
 Gln Ser Ser Trp Ser
                 140
 <210> 15
 <211> 248
 <212> PRT
 <213> Homo sapiens
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 <223> Incvte ID No: 7511498CD1
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Met Ala Ser Arg Gly Arg Arg Pro Glu His Gly Gly Pro Pro Glu
                 5
                                    10
Leu Phe Tyr Asp Glu Thr Glu Ala Arg Lys Tyr Val Arg Asn Ser
                20
                                    25
Arg Met Ile Asp Ile Gln Thr Arg Met Ala Gly Arg Ala Leu Glu
                35
                                    40
Leu Leu Tyr Leu Pro Glu Asn Lys Pro Cys Tyr Leu Leu Asp Ile
                50
Gly Cys Gly Thr Gly Leu Ser Gly Ser Tyr Leu Ser Asp Glu Gly
                 65
                                    70
His Tyr Trp Val Gly Leu Asp Ile Ser Pro Ala Met Leu Asp Glu
                 80
                                    85
Ala Val Asp Arg Glu Ile Glu Gly Asp Leu Leu Leu Gly Asp Met
                 95
                                    100
Gly Gln Gly Ile Pro Phe Lys Pro Gly Thr Phe Asp Gly Cys Ile
                110
                                    115
Ser Ile Ser Ala Val Gln Trp Leu Cvs Asn Ala Asn Lys Lys Ser
                125
                                    130
Glu Asn Pro Ala Lys Arg Leu Tyr Cys Phe Phe Ala Ser Leu Phe
                140
                                    145
Ser Val Leu Val Arg Gly Ser Arg Ala Val Leu Gln Leu Tyr Pro
                155
                                    160
Glu Asn Ser Glu Gln Leu Glu Leu Ile Thr Thr Gln Ala Thr Lys
                170
                                    175
Ala Gly Phe Ser Gly Gly Met Val Val Asp Tyr Pro Asn Ser Ala
                185
                                    190
Lys Ala Lys Lys Phe Tyr Leu Cys Leu Phe Ser Gly Pro Ser Thr
                200
                                    205
Phe Ile Pro Glu Val Pro Ile Lys Asp Val Glu Ala Gly Asn Gly
                215
                                    220
                                                        225
Glu Glu Glu Ser Gly Met Gly Ala Gly Glu Glu Gly Ala Ala Gln
                230
                                    235
Ala Pro Gly Gln Gly Ser Gln Thr
                245
<210> 16
<211> 153
<212> PRT
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Val Asn Leu Arc Pro Glu Gln Leu Glv Asp Ile Cvs Val Glv Asn 80 85 Val Leu Gln Pro Gly Ala Gly Ala Ile Met Ala Arg Ile Ala Gln 95 100 Phe Leu Ser Asp Ile Pro Glu Thr Val Pro Leu Ser Thr Val Asn 110 115 Arg Gln Cys Ser Ser Gly Leu Gln Ala Val Ala Ser Ile Ala Gly 125 130 Trp Ser Pro Cys Pro Trp Leu Thr Glu Gly Thr Leu Glu Ile Leu 140 145 Leu Arg Ala

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<213> Homo sapiens

<220> <221> misc\_feature

<223> Incyte ID No: 7511624CD1

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Met Ala Leu Leu His Ser Gly Arg Val Leu Pro Gly Ile Ala Ala 10 Ala Phe His Pro Gly Leu Ala Ala Ala Ala Ser Ala Arg Ala Ser 20 25 Ser Trp Trp Thr His Val Glu Met Gly Pro Pro Asp Pro Ile Leu 40 Gly Val Thr Glu Ala Phe Lys Arg Asp Thr Asn Ser Lys Lys Met 50 55 Asn Leu Gly Val Gly Ala Tyr Arg Asp Asp Asn Gly Lys Pro Tyr 70 Val Leu Pro Ser Val Arg Lys Ala Glu Ala Gln Ile Ala Ala Lys 80 85 Asn Leu Asp Lys Glu Tyr Leu Pro Ile Gly Gly Leu Ala Glu Phe 95 100 Cys Lys Ala Ser Ala Glu Leu Ala Leu Gly Glu Asn Ser Glu Val 110 115 Leu Lvs Ser Glv Arg Phe Val Thr Val Gln Thr Ile Ser Glv Thr 125 130 Gly Ala Leu Arg Ile Gly Ala Ser Phe Leu Lys Ile Pro Glu Gln 140 145 Ser Val Leu Leu His Ala Cys Ala His Asn Pro Thr Gly Val 155 160 Asp Pro Arg Pro Glu Gln Trp Lys Glu Ile Ala Thr Val Val Lys 170 175 Lys Arg Asn Leu Phe Ala Phe Phe Asp Met Ala Tyr Gln Gly Phe 185 190 Ala Ser Gly Asp Gly Asp Lys Asp Ala Trp Ala Val Arg His Phe 200 205

Ile Glu Gln Gly Ile Asn Val Cys Leu Cys Gln Ser Tyr Ala Lys 215 220 Asn Met Gly Leu Tyr Gly Glu Arg Val Gly Ala Phe Thr Met Val 230 235

Cys Lys Asp Ala Asp Glu Ala Lys Arg Val Glu Ser Gln Leu Lys

```
250
                245
Ile Leu Ile Arg Pro Met Tyr Ser Asn Pro Pro Leu Asn Gly Ala
                260
                                    265
Arg Ile Ala Ala Ala Ile Leu Asn Thr Pro Asp Leu Arg Lys Gln
                                    280
                275
Trp Leu Gln Glu Val Lys Gly Met Ala Asp Arg Ile Ile Gly Met
                                    295
                290
Arg Thr Gln Leu Val Ser Asn Leu Lys Lys Glu Gly Ser Thr His
                                    310
                305
Asn Trp Gln His Ile Thr Asp Gln Ile Gly Met Phe Cys Phe Thr
                                    325
                320
Gly Leu Lys Pro Glu Gln Val Glu Arg Leu Ile Lys Glu Phe Ser
                                    340
Ile Tyr Met Thr Lys Asp Gly Arg Ile Ser Val Ala Gly Val Thr
                                    355
                350
Ser Ser Asn Val Gly Tyr Leu Ala His Ala Ile His Gln Val Thr
                                    370
                365
Lys
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Met Ala Leu Leu His Ser Gly Arg Val Leu Pro Gly Ile Ala Ala
Ala Phe His Pro Gly Leu Ala Ala Ala Ala Ser Ala Arg Ala Ser
                 20
Ser Tro Tro Thr His Val Glu Met Gly Pro Pro Asp Pro Ile Leu
                 35
Gly Val Thr Glu Ala Phe Lys Arg Asp Thr Asn Ser Lys Lys Met
                                      55
Asn Leu Gly Val Gly Ala Tyr Arg Asp Asp Asn Gly Lys Pro Tyr
                                      70
                  65
Val Leu Pro Ser Val Arg Lys Phe Val Thr Val Gln Thr Ile Ser
                                     85
                  80
Gly Thr Gly Ala Leu Arg Ile Gly Ala Ser Phe Leu Gln Arg Phe
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                                     100
Phe Lys Phe Ser Arg Asp Val Phe Leu Pro Lys Pro Thr Trp Gly
                                     115
                 110
Asn His Thr Pro Ile Phe Arg Asp Ala Gly Met Gln Leu Gln Gly
                                     130
                 125
Tyr Arg Tyr Tyr Asp Pro Lys Thr Cys Gly Phe Asp Phe Thr Gly
                140
                                     145
Ala Val Glu Asp Ile Ser Lys Ile Pro Glu Gln Ser Val Leu Leu
                                     160
                 155
Leu His Ala Cys Ala His Asn Pro Thr Gly Val Asp Pro Arg Pro
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                                     175
Glu Gln Trp Lys Glu Ile Ala Thr Val Val Lys Lys Arg Asn Leu
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                 185
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Phe Ala Phe Phe Asp Met Ala Tyr Gln Gly Phe Ala Ser Gly Asp
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Gly Asp Lys Asp Ala Trp Ala Val Arg His Phe Ile Glu Gln Gly
                                    220
Ile Asn Val Cys Leu Cys Gln Ser Tyr Ala Lys Asn Met Gly Leu
                230
                                    235
Tyr Gly Glu Arg Val Gly Ala Phe Thr Met Val Cys Lys Asp Ala
                245
                                    250
Asp Glu Ala Lys Arg Val Glu Ser Gln Leu Lys Ile Leu Ile Arg
                                    265
                260
Pro Met Tyr Ser Asn Pro Pro Leu Asn Gly Ala Arg Ile Ala Ala
                275
                                    280
Ala Ile Leu Asn Thr Pro Asp Leu Arg Lys Gln Trp Leu Gln Glu
                290
                                    295
Val Lys Gly Met Ala Asp Arg Ile Ile Gly Met Arg Thr Gln Leu
                305
                                    310
Val Ser Asn Leu Lys Lys Glu Gly Ser Thr His Asn Trp Gln His
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                                    325
Ile Thr Asp Gln Ile Gly Met Phe Cys Phe Thr Gly Leu Lys Pro
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                                    340
Glu Gln Val Glu Arg Leu Ile Lys Glu Phe Ser Ile Tyr Met Thr
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Lys Asp Gly Arg Ile Ser Val Ala Gly Val Thr Ser Ser Asn Val
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Gly Tyr Leu Ala His Ala Ile His Gln Val Thr Lys
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Cys Leu Gly Val Phe Gly Leu Phe Arg Leu Leu Gln Trp Val Arg
                                     40
                 35
Gly Lys Ala Tyr Leu Arg Asn Ala Val Val Val Ile Thr Gly Ala
                 50
                                     55
Thr Ser Gly Leu Gly Lys Glu Cys Ala Lys Val Phe Tyr Ala Ala
                                     70
                 65
Gly Ala Lys Leu Val Leu Cys Gly Arg Asn Gly Gly Ala Leu Glu
Glu Leu Ile Arg Glu Leu Thr Ala Ser His Ala Thr Lys Val Gln
                                    100
                 95
Thr His Lys Pro Tyr Leu Val Thr Phe Asp Leu Thr Asp Ser Gly
                110
                                    115
Ala Ile Val Ala Ala Ala Glu Ile Leu Gln Cys Phe Gly Tyr
                                    130
                125
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Val Asp Ile Leu Val Asn Asn Ala Gly Ile Ser Tyr Arg Gly Thr

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145
Ile Met Asp Thr Thr Val Asp Val Asp Lys Arg Val Met Glu Thr
                155
                                   160
Asn Tyr Phe Gly Pro Val Ala Leu Thr Lys Ala Leu Leu Pro Ser
                                   175
Met Ile Lys Arg Arg Gln Gly His Ile Val Ala Ile Ser Ser Ile
                                   190
                185
Gln Gly Lys Met Ser Ile Pro Phe Arg Ser Ala Phe Met Asp Thr
                                   205
                200
Thr Thr Ala Gln Gly Arg Ser Pro Val Glu Val Ala Gln Asp Val
                                    220
                215
Leu Ala Ala Val Gly Lys Lys Lys Asp Val Ile Leu Ala Asp
                230
Leu Leu Pro Ser Leu Ala Val Tyr Leu Arg Thr Leu Ala Pro Gly
                                    250
Leu Phe Phe Ser Leu Met Ala Ser Arg Ala Arg Lys Glu Arg Lys
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Ser Lvs Asn Ser
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Ala Val Lys Val Ala Ile Asp Ala Gly Tyr Arg His Ile Asp Cys
                 35
                                     40
Ala Tyr Val Tyr Gln Asn Glu His Glu Val Gly Glu Ala Ile Gln
                 50
                                     55
Glu Lys Ile Gln Glu Lys Ala Val Met Arg Glu Asp Leu Phe Ile
                                     70
                 65
Val Ser Lys Val Trp Pro Thr Phe Phe Glu Arg Pro Leu Val Arg
                                     85
Lys Ala Phe Glu Lys Thr Leu Lys Asp Leu Lys Leu Ser Tyr Leu
                                    100
                 95
Asp Val Tyr Leu Ile His Trp Pro Gln Gly Phe Lys Thr Gly Asp
                110
                                    115
Asp Phe Phe Pro Lys Asp Asp Lys Gly Asn Met Ile Ser Gly Lys
                125
                                    130
Gly Thr Phe Leu Asp Ala Trp Glu Ile Glu Arg Leu Leu Asn Lys
                                    145
                140
Pro Gly Leu Lys Tyr Lys Pro Val Thr Asn Gln Val Glu Cys His
                                    160
                155
Pro Tyr Leu Thr Gln Glu Lys Leu Ile Gln Tyr Cys His Ser Lys
                                    175
                170
Gly Ile Thr Val Thr Ala Tyr Ser Pro Leu Gly Ser Pro Asp Arg
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                                    190
                 185
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Pro Trp Ala Lvs Pro Glu Asp Pro Ser Leu Leu Glu Asp Pro Lvs
                200
                                   205
Ile Lys Glu Ile Ala Ala Lys His Lys Lys Thr Thr Ala Gln Val
                215
                                    220
Leu Ile Arg Phe His Ile Gln Arg Asn Val Thr Val Ile Pro Lys
               230
                                   235
Ser Met Thr Pro Ala His Ile Val Glu Asn Ile Gln Val Phe Asp
               245
                                   250
Phe Lys Leu Ser Asp Glu Glu Met Ala Thr Ile Leu Ser Phe Asn
               260
                                   265
Arg Asn Trp Arg Ala Phe Asp Phe Lys Glu Phe Ser His Leu Glu
               275
                                   280
Asp Phe Pro Phe Asp Ala Glu Tyr
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Arg Arg Met Val Gln Glu Arg Glu Lys Asp Ile Leu Thr Ala Ile
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                                     40
Ala Ala Asp Leu Cys Lys Ser Glu Phe Asn Val Tyr Ser Gln Glu
                 50
                                    55
Val Ile Thr Val Leu Glv Glu Ile Asp Phe Met Leu Glu Asn Leu
                 65
                                    70
Pro Glu Trp Val Thr Ala Lys Pro Val Lys Lys Asn Val Leu Thr
                80
                                    85
Met Leu Asp Glu Ala Tyr Ile Gln Pro Gln Pro Leu Gly Val Val
                95
                                   100
Leu Ile Ile Gly Ala Trp Asn Tyr Pro Phe Val Leu Thr Ile Gln
                110
                                   115
Pro Leu Ile Gly Ala Ile Ala Ala Gly Asn Ala Val Ile Ile Lys
               125
                                    130
Pro Ser Glu Leu Ser Glu Asn Thr Ala Lys Ile Leu Ala Lys Leu
               140
                                   145
Leu Pro Gln Tyr Leu Asp Gln Glu Phe Tyr Gly Glu Asn Ile Lys
                155
                                    160
Glu Ser Pro Asp Tvr Glu Arg Ile Ile Asp Leu Arg His Phe Lys
                170
                                    175
Arg Ile Leu Ser Leu Leu Glu Glv Gln Lvs Ile Ala Phe Glv Glv
                185
                                   190
Glu Thr Asp Glu Ala Thr Arg Tyr Ile Ala Pro Thr Val Leu Thr
                200
                                   205
Asp Val Asp Pro Lys Thr Lys Val Met Gln Glu Glu Ile Phe Gly
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                                   220
Pro Ile Leu Pro Ile Val Pro Val Lys Asn Val Asp Glu Ala Ile
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235
               230
Asn Phe Ile Asn Glu Arg Glu Lys Pro Leu Ala Leu Tyr Val Phe
                                    250
               245
Ser His Asn His Lys Leu Ile Lys Arg Met Ile Asp Glu Thr Ser
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                                    265
Ser Gly Gly Val Thr Gly Asn Asp Val Ile Met His Phe Thr Leu
                                    280
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Asn Ser Phe Pro Phe Gly Gly Val Gly Ser Ser Gly Met Gly Ala
                                    295
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Tyr His Gly Lys His Ser Phe Asp Thr Phe Ser His Gln Arg Pro
                                    310
                305
Cys Leu Leu Lys Ser Leu Lys Arg Glu Gly Ala Asn Lys Leu Arg
                                    325
                320
Tyr Pro Pro Asn Ser Gln Ser Lys Val Asp Trp Gly Lys Phe Phe
                                    340
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Leu Leu Lys Arg Phe Asn Lys Glu Lys Leu Gly Leu Leu Leu
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Thr Phe Leu Gly Ile Val Ala Ala Val Leu Val Lys Ala Glu Tyr
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                365
Tyr
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                                      40
                 35
Val Thr Gly Ala Asn Thr Gly Ile Gly Lys Glu Thr Ala Lys Glu
                 50
                                      55
Leu Ala Gln Arg Gly Ala Arg Val Tyr Leu Ala Cys Arg Asp Val
                                     70
                 65
Glu Lys Gly Glu Leu Val Ala Lys Glu Ile Gln Thr Thr Thr Gly
                                      85
 Asn Gln Gln Val Leu Val Arg Lys Leu Asp Leu Ser Asp Thr Lys
                                     100
                 95
 Ser Ile Arg Ala Phe Ala Lys Gly Phe Leu Ala Glu Glu Lys His
                                     115
                 110
 Leu His Val Leu Ile Asn Asn Ala Gly Val Met Met Cys Pro Tyr
                 125
                                     130
 Ser Lys Thr Ala Asp Gly Phe Glu Met His Ile Gly Val Asn His
                                     145
                 140
 Leu Gly Ser Gly Val Thr Thr Tyr Ser Val His Pro Gly Thr Val
                 155
                                    160
 Gln Ser Glu Leu Val Arg His Ser Ser Phe Met Arg Trp Met Trp
                                                         180
                 170
                                     175
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Trp Leu Phe Ser Phe Phe Ile Lys Thr Pro Gln Gln Gly Ala Gln
                                    190
                185
Thr Ser Leu His Cys Ala Leu Thr Glu Gly Leu Glu Ile Leu Ser
                200
                                    205
Gly Asn His Phe Ser Asp Cys His Val Ala Trp Val Ser Ala Gln
                                    220
                215
Ala Arg Asn Glu Thr Ile Ala Arg Arg Leu Trp Asp Val Ser Cys
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Asp Leu Leu Gly Leu Pro Ile Asp
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His Val Ser Ala Ile Pro Trp Gly Ile Ser Ala Thr Ala Ala Leu
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                                     25
Arg Pro Ala Ala Cys Gly Arg Thr Ser Leu Thr Asn Leu Leu Cys
                 35
                                     40
Ser Gly Ser Ser Gln Ala Lys Leu Phe Ser Thr Ser Ser Ser Cys
                 50
                                     55
His Ala Pro Ala Val Thr Gln His Ala Pro Tyr Phe Lys Gly Thr
                 65
                                     70
Ala Val Val Asn Gly Glu Phe Lys Asp Leu Ser Leu Asp Asp Phe
                                     85
Lys Gly Lys Tyr Leu Val Leu Phe Phe Tyr Pro Leu Asp Phe Thr
                 95
                                    100
Phe Val Cys Pro Thr Glu Ile Val Ala Phe Ser Asp Lys Ala Asn
                110
                                    115
Glu Phe His Asp Val Asn Cys Glu Val Val Ala Val Ser Val Asp
                125
                                    130
Ser His Phe Ser His Leu Ala Trp Ile Asn Thr Pro Arg Lys Asn
                140
                                    145
Gly Gly Leu Gly His Met Asn Ile Ala Leu Leu Ser Asp Leu Thr
                155
                                    160
Lys Gln Ile Ser Arg Asp Tyr Gly Val Leu Leu Glu Gly Ser Gly
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Leu Ala Leu Arg Ser Ser Gln Val Gln Leu Leu Pro Lys Ser Thr
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Phe Arg Arg
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Ser Leu Trp Ala Thr Asn His Val Pro Glu Met Val Arg Pro Thr
                                    40
                 35
Leu Glu Arg Thr Leu Arg Val Leu Gln Leu Asp Tyr Val Asp Leu
                                     55
                 50
Tyr Ile Ile Glu Val Pro Met Ala Phe Lys Pro Gly Asp Glu Ile
                 65
                                     70
Tyr Pro Arg Asp Glu Asn Gly Lys Trp Leu Tyr His Lys Ser Asn
                 80
                                     85
Leu Cys Ala Thr Trp Glu Ala Met Glu Ala Cys Lys Asp Ala Gly
                 95
                                   100
Leu Val Lys Ser Leu Gly Val Ser Asn Phe Asn Arg Arg Gln Leu
                110
                                    115
Glu Leu Ile Leu Asn Lys Pro Gly Leu Lys His Lys Pro Val Ser
                125
                                   130
Asn Gln Val Glu Cys His Pro Tyr Phe Thr Gln Pro Lys Leu Leu
                140
                                   145
Lys Phe Cys Gln Gln His Asp Ile Val Ile Thr Ala Tyr Ser Pro
                155
                                   160
Leu Gly Thr Ser Arg Asn Pro Ile Trp Val Asn Val Ser Ser Pro
                170
                                    175
Pro Leu Leu Lys Asp Ala Leu Leu Asn Ser Leu Gly Lys Arg Tyr
                                    190
Asn Lys Thr Ala Ala Gln Ile Val Leu Arg Phe Asn Ile Gln Arg
                200
                                    205
Gly Val Val Val Ile Pro Lys Ser Phe Asn Leu Glu Arg Ile Lys
                215
                                    220
Glu Asn Phe Gln Ile Phe Asp Phe Ser Leu Thr Glu Glu Glu Met
                                    235
                230
Lys Asp Ile Glu Ala Leu Asn Lys Asn Val Arg Phe Val Glu Leu
                245
                                    250
Leu Met Trp Arg Asp His Pro Glu Tyr Pro Phe His Asp Glu Tyr
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15
Pro His Ser Gly Pro Leu Val Ser Trp Arg Arg Trp Ser Gly Ala
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25
30

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Ser Thr Asp Thr Val Tvr Asp Val Val Val Ser Gly Gly Gly Leu
Val Glv Ala Ala Met Ala Cvs Ala Leu Glv Tvr Asp Ile His Phe
                 50
                                     55
His Asp Lys Lys Ile Leu Leu Leu Glu Ala Gly Pro Lys Lys Val
                65
                                     70
Leu Glu Lys Leu Ser Glu Thr Tyr Ser Asn Arg Val Ser Ser Ile
                 80
                                     85
Ser Pro Gly Ser Ala Thr Leu Leu Ser Ser Phe Gly Ala Trp Asp
                 95
                                    100
His Ile Cys Asn Met Arg Tyr Arg Ala Phe Arg Arg Met Gln Val
                                    115
                110
Trp Asp Ala Cys Ser Glu Ala Leu Ile Met Phe Asp Lys Asp Asn
                125
Leu Asp Asp Met Gly Tyr Ile Val Glu Asn Asp Val Ile Met His
                140
                                    145
Ala Leu Thr Lys Gln Leu Glu Ala Val Ser Asp Arg Val Thr Val
                155
                                    160
Leu Tyr Arg Ser Lys Ala Ile Arg Tyr Thr Trp Pro Cys Pro Phe
                170
                                    175
Pro Met Ala Asp Ser Ser Pro Trp Val His Ile Thr Leu Gly Asp
                185
                                    190
Gly Ser Thr Phe Gln Thr Lys Leu Leu Ile Gly Ala Asp Gly His
                200
                                    205
Asn Ser Gly Val Arg Gln Ala Val Gly Ile Gln Asn Val Ser Trp
                                    220
                215
Asn Tyr Asp Gln Ser Ala Val Val Ala Thr Leu His Leu Ser Glu
                                    235
Ala Thr Glu Asn Asn Val Ala Trp Gln Arg Phe Leu Pro Ser Gly
                245
                                    250
Pro Ile Ala Leu Leu Pro Leu Ser Asp Thr Leu Ser Ser Leu Val
                                    265
Trp Ser Thr Ser His Glu His Ala Ala Glu Leu Val Ser Met Asp
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                                    280
Glu Glu Lys Phe Val Asp Ala Val Asn Ser Ala Phe Glu Gln Ile
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                                    295
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Met Ala Phe Ala Ser Lys
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His	ITe	Cys	Asn	Met 65	Arg	Tyr	Arg	Ala	Phe 70	Arg	Arg	Met	GIn	75
Tro	Asp	Ala	Cvs		Glu	Ala	Leu	Ile		Phe	Asp	Lvs	Asp	
-	-		•	80					85		_	-	-	90
Leu	Asp	Asp	Met	${\tt Gly}$	Tyr	Ile	Va1	Glu		Asp	Val	Ile	Met	His
				95					100					105
Ala	Leu	Thr	Lys		Leu	G1u	Ala	Va1		Asp	Arg	Val	Thr	
Lou	m rx	Arg	cor	110	21.	т1 о	) ra	(II)s roc	115	m-rn	Dro	Circ	Dro	120
Бец	TÄT	ALG	per	125	ALG	TTE	ALG	TAT	130	TTD	FLU	Cys	FIU	135
Pro	Met	A1a	Asp		Ser	Pro	Trp	Va1		I1e	Thr	Leu	G1y	
				140			-		145					150
G1y	Ser	Thr	Phe		Thr	Lys	Leu	Leu		Gly	Ala	Asp	G1y	
_	_			155					160		_		_	165
Asn	ser	G1y	Val	Arg 170	GIn	Ala	Val	GIĀ	175	GIn	Asn	Val	Ser	Trp
Asn	Tur	Asp	G1n		M1a	Va1	Va1	λla		T.em	His	Len	Ser	
	-1-	1100		185			•		190	204				195
Ala	Thr	Glu	Asn	Asn	Val	Ala	Trp	G1n	Arg	Phe	Leu	Pro	Ser	G1y
				200					205					210
Pro	I1e	A1a	Leu		Pro	Leu	Ser	Asp		Leu	Ser	Ser	Leu	
m	0	Thr		215	a1	*** _	31-	21-	220	·	17-1		Mak	225
TTD	ser	THE	ser	230	GIU	nrs	Ala		235	ren	vai	ser	met	240
Glu	Glu	Lys	Phe		Asp	Ala	Va1			Ala	Phe	Trp	Ser	
		-		245	-				250			-		255
Ala	Asp	His	Thr		Phe	Ile	Asp	Thr		G1y	A1a	Met	Leu	
_			_	260	_	_	_		265		_			270
Tyr	Ala	Va1	Ser	275	Leu	Lys	Pro	Thr	Lys 280	Val	Ser	Ala	Arg	285
T.em	Pro	Pro	Ser		λla	Ara	Va1	Aen		Tare	Ser	Ara	va1	
Doa		110	DUL	290		9	****	, mp	295	23.5		, a.g		300
Phe	Pro	Leu	Gly	Leu	G1y	His	Ala	Ala	G1u	Tyr	Val	Arg	Pro	Arg
				305					310					315
Val	Ala	Leu	I1e		Asp	Ala	A1a	His		Va1	His	Pro	Leu	
C3.r	C1 n	Gly	17.7	320	Mot	C3.11	Dho	C1.,	325	т1.	0	e	Tou	330
GIY	GIII	GIĀ	Val	335	met	GTĀ	Pne	GIY	340	TTE	ser	ser	Leu	345
His	His	Leu	Ser		A1a	Ala	Phe	Asn		Lvs	Asp	Leu	Glv	
				350					355	-	-		_	360
Va1	Ser	His	Leu		Gly	Tyr	G1u	Thr		Arg	Gln	Arg	His	
				365					370					375
Thr	Ala	Leu	Leu	Ala 380	Ala	Thr	Asp	Leu	Leu 385	Lys	Arg	Leu	Tyr	Ser 390
Thr	Ser	Ala	Sor		Lon	17m3	Lon	Len		Thr	Tra	G117	Lon	
			~ 63.	395	2-cu	var	Zeu		400		12.10	CLY	ou	405
Ala	Thr	Asn	Ala	Val	Ser	Pro	Leu	Lys	Glu	Gln	I1e	Met	Ala	
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Ala	Ser	Lys												

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Glu Glu Val Gln Ala Ile Met Glu Ala Tyr Glu Ser Asp Pro Thr
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                                     40
Glu Trp Ala Met Tyr Ala Lys Phe Asp Gln Tyr Arg Tyr Thr Arg
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Asn Leu Val Asp Gln Gly Asn Gly Lys Phe Asn Leu Met Ile Leu
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Cys Trp Gly Glu Gly His Gly Arg Cys Tyr Arg Glu Ile
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Ala Ala Ser Tyr His Leu Ser Arg Ala Pro Cys Pro Pro Lys Val
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Val Leu Val Glu Ser Ser Glu Arg Leu Gly Gly Trp Ile Arg Ser
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Val Arg Gly Pro Asn Gly Ala Ile Phe Glu Leu Gly Pro Arg Gly
                                     55
                 50
Ile Arg Pro Ala Gly Ala Leu Gly Ala Arg Thr Leu Leu Leu Val
                                     70
                 65
Ser Glu Leu Gly Leu Asp Ser Glu Val Leu Pro Val Arg Gly Asp
                                     85
                 80
His Pro Ala Ala Gln Asn Arg Phe Leu Tyr Val Gly Gly Ala Leu
                                     100
                 95
His Ala Leu Pro Thr Gly Leu Arg Glu Leu Thr Lys Pro Arg Gly
                                     115
                 110
Lys Glu Pro Asp Glu Thr Val His Ser Phe Ala Gln Arg Arg Leu
                 125
Gly Pro Glu Val Ala Ser Leu Ala Met Asp Ser Leu Cys Arg Gly
                                    145
                 140
Val Phe Ala Gly Asn Ser Arg Glu Leu Ser Ile Arg Ser Cys Phe
                                     160
                 155
 Pro Ser Leu Phe Gln Ala Glu Gln Thr His Arg Ser Ile Leu Leu
                                     175
                 170
 Gly Leu Leu Gly Ala Gly Arg Thr Pro Gln Pro Asp Ser Ala
                                    190
                 185
 Leu Ile Arg Gln Ala Leu Ala Glu Arg Trp Ser Gln Trp Ser Leu
                                     205
                                                         210
                 200
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Arg Gly Gly Leu Glu Met Leu Pro Gln Ala Leu Glu Thr His Leu 215 220 Thr Ser Arg Gly Val Ser Val Leu Arg Gly Gln Pro Val Cys Gly 230 235 Leu Ser Leu Gln Ala Glu Gly Arg Trp Lys Val Ser Leu Arg Asp 250 Ser Ser Leu Glu Ala Asp His Val Ile Ser Ala Ile Pro Ala Ser 265 260 Val Leu Ser Glu Leu Leu Pro Ala Glu Ala Ala Pro Leu Ala Arg 275 280 Ala Leu Ser Ala Ile Thr Ala Val Ser Val Ala Val Val Asn Leu 295 290 Gln Tyr Gln Gly Ala His Leu Pro Val Gln Gly Phe Gly His Leu 305 310 Val Pro Ser Ser Glu Asp Pro Gly Val Leu Gly Ile Val Tyr Asp 325 Ser Val Ala Phe Pro Glu Gln Asp Gly Ser Pro Pro Gly Leu Arg 340 335 Val Thr Val Met Leu Gly Gly Ser Trp Leu Gln Thr Leu Glu Ala 355 Ser Gly Cys Val Leu Ser Gln Glu Leu Phe Gln Gln Arg Ala Gln 370 365 Glu Ala Ala Ala Thr Gln Leu Gly Leu Lys Glu Met Pro Ser His 380 385 Cys Leu Val His Leu His Lys Asn Cys Ile Pro Gln Tyr Thr Leu 395 400 Gly His Trp Gln Lys Leu Glu Ser Ala Arg Gln Phe Leu Thr Ala 410 415 His Arg Leu Pro Leu Thr Leu Ala Gly Ala Ser Tyr Glu Gly Val 430 Ala Val Asn Asp Cvs Ile Glu Ser Gly Arg Gln Ala Ala Val Ser 440 445 Val Leu Gly Thr Glu Pro Asn Ser 455

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                                   115
His Leu His Lys Asn Cys Ile Pro Gln Tyr Thr Leu Gly His Trp
                125
                                   130
Gln Lys Leu Glu Ser Ala Arg Gln Phe Leu Thr Ala His Arg Leu
                                   145
Pro Leu Thr Leu Ala Gly Ala Ser Tyr Glu Gly Val Ala Val Asn
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Asp Cys Ile Glu Ser Gly Arg Gln Ala Ala Val Ser Val Leu Gly
                                   175
Thr Glu Pro Asn Ser
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Ala Ala Ser Tyr His Leu Ser Arg Ala Pro Cys Pro Pro Lys Val
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Met Leu Gly Gly Ser Trp Leu Gln Thr Leu Glu Ala Ser Gly Cys
                 35
                                     40
Val Leu Ser Gln Glu Leu Phe Gln Gln Arg Ala Gln Glu Ala Ala
                                     55
                 50
Ala Thr Gln Leu Gly Leu Lys Glu Met Pro Ser His Cys Leu Val
                                     70
                 65
His Leu His Lys Asn Cys Ile Pro Gln Tyr Thr Leu Gly His Trp
                                     85
                 80
Gln Lys Leu Glu Ser Ala Arg Gln Phe Leu Thr Ala His Arg Leu
                                    100
                 95
 Pro Leu Thr Leu Ala Gly Ala Ser Tyr Glu Gly Val Ala Val Asn
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 Asp Cys Ile Glu Ser Gly Arg Gln Ala Ala Val Ser Val Leu Gly
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 Thr Glu Pro Asn Ser
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Arg Thr Gly Asn Asp Val Leu Lys Ala Leu Ala Leu Gly Ala Lys
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Cys Ile Phe Leu Gly Arg Pro Ile Leu Trp Gly Leu Ala Cys Lys
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Gly Glu His Gly Val Lys Glu Val Leu Asn Ile Leu Thr Asn Glu
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Lys Ala Gly Asp Glu Phe Val Glu Lys Thr Leu Ser Thr Ile Ala
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Thr Ser Thr Asp Ala Ala Ser Val Val His Ser Thr Asp Leu Val
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Leu Trp Ala Lys Ser Pro Gly Val Leu Ala Gly Gln Pro Phe Phe
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                                   160
Cys Gly Pro His Ile Asp Val Ile Ser Met Gly Met Leu Thr Gln
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Thr Leu Asp Asn Leu Pro Gln Phe Cys Gly Pro His Ile Asp Val
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Pro Leu Lys Pro

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